

A new species of *Moaciria* (Nematoda, Heterakidae) and other helminths in the red Mawatta frog, *Hylophorbus* cf. *rufescens* (Anura, Microhylidae) from Papua New Guinea

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Abstract

Moaciria moraveci sp. nov. (Nematoda, Heterakidae) from the large intestine of Hylophorbus cf. rufescens from Papua New Guinea is described. Moaciria moraveci sp. nov. represents the 9th species assigned to the genus and the 5th from the Australo-Papuan region. It is distinguished from congeners by the distribution pattern of the caudal papillae of the male, spicule length and vulvar position.

Key words

Nematoda, Moaciria moraveci sp. nov., Anura, Hylophorbus cf. rufescens, Papua New Guinea

Introduction

In a helminthological survey of frogs of Papua New Guinea, 10 of 21 (48%) red Mawatta frogs, Hylophorbus cf. rufescens Macleay (Microhylidae), were found to harbor an undescribed species of *Moaciria* Freitas, 1956. The systematics for the genus Hylophorbus Macleay from eastern New Guinea are unresolved; however, several unnamed species are involved and members of the genus occupy the whole of Papua New Guinea from sea level to 3570 m as well as the offshore d'Entrecasteaux and Louisiade archipelagos and Woodlark Island (Zweifel 1972, Menzies 2006). Type locality for *Hylophorbus* rufescens is Katow in southwestern Papua New Guinea (Zweifel 1972) and the host specimens discussed herein represent a closely related but yet-to-be determined species. To our knowledge, there are no helminthological reports for this frog. Moaciria was established by Freitas (1956) to receive Moaciria alvarengai Freitas, 1956, a newly described nematode from the lizard Mabuya maculata (Gray) collected at Ilha Fernando de Noronha, Brazil. The purpose of this paper is to describe the 9th species assigned to *Moaciria* and to provide an initial parasite list for this host.

Materials and methods

Twenty-one adult *Hylophorbus* cf. *rufescens* from Fergusson Island, Milne Bay Province, Papua New Guinea in the collec-

tion of the Bernice P. Bishop Museum, Honolulu, Hawaii (BPBM) were examined for helminths: 12 (BPBM 16181, 16184-16187, 16192-16194, 16196, 16197, 16199, 16204) collected on Oya Tabu, 12-26 August 2002; 9 (BPBM 16206-16214), collected 9-14 September 2002 on Oya Waka. The abdominal cavity of each frog was opened and the gastrointestinal tract was removed, opened longitudinally and searched for helminths using a dissecting microscope. The coelom was also searched. The helminths, fixed in situ, were cleared in lactophenol on a glass slide and examined using a compound microscope. Illustrations were made with the aid of a microprojector. Measurements are given in micrometers, unless otherwise stated, as mean \pm 1 SD with range in parentheses. Type specimens were deposited in the United States National Parasite Collection (USNPC), Beltsville, Maryland; voucher specimens in USNPC and BPBM.

Results

Twenty-six nematodes, 3 mature males and 23 mature females, of a species of *Moaciria* were found in the large intestines of 10 of 21 *Hylophorbus* cf. *rufescens*. Mature individuals of three species of Nematoda, *Cosmocerca novaeguineae* Moravec et Sey, 1990, *Cosmocerca tyleri* Bursey, Goldberg et Kraus, 2006, and *Oswaldocruzia bakeri* Moravec et Sey, 1986, and larvae (in cysts) of another nematode, *Physocephalus* sp., as well as acanthocephalan cystacanths were also

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Table I. Site of infection, number of helminths, prevalence, mean intensity, range of infection and USNPC and BPBM accession numbers	;
for voucher specimens of 6 helminth species from Hylophorbus cf. rufescens from Papua New Guinea	

Helminth species	Site of infection	Number	Prevalence	Mean intensity	Range	Accession number	
				± SD		SNPC	BPBM
Cosmocerca novaeguineae	large intestine	24	5/21 (23%)	4.8 ± 2.7	1–7	99596	H205
Cosmocerca tyleri	large intestine	274	9/21 (42%)	30.4 ± 24.6	4-73	99597	H206
Moaciria moraveci sp. nov.	large intestine	26	10/21 (48%)	2.6 ± 3.0	1-11	99598	H207
Oswaldocruzia bakeri	large intestine	18	6/21 (28%)	3.0 ± 2.2	1–7	99599	H208
Physocephalus sp. (cysts)	stomach wall	18	2/21 (10%)	9.0 ± 11.3	1 - 17	99600	H209
Acanthocephalan cystacanths	body cavity	14	4/21 (19%)	3.5 ± 1.3	2-5	99601	H210

found. Numbers of individuals, prevalence and mean intensity for each helminth species are presented in Table I. Description of the new species follows.

Moaciria moraveci sp. nov. (Figs 1-8)

General: Heterakidae Railliet et Henry, 1912; Spinicaudinae Travassos, 1920; *Moaciria* Freitas, 1956. Small, stout nematodes, females larger than males. Body cylindrical, fusiform, however posteriorly truncated in males. Cuticle with transverse striations. Narrow lateral alae extending along entire length of body. Mouth with three lips offset from body, dorsal lip with 2 sessile papillae, each ventrolateral lip with a sessile papilla and a stalked amphid. Oesophagus divisible into anterior pharynx, corpus and posterior valved bulb. Somatic papillae present. Males with caudal sucker and caudal alae supported by papillae.

Male (holotype and 2 paratypes): Length 2.75 ± 0.14 mm (2.59-2.85 mm), width at midbody 267 ± 55 (204–306). Narrow alae beginning at base of lips and ending at caudal truncation. Length of lips 15 ± 3 (12–18); pharynx 47 ± 3 (43–49) long; corpus 433 ± 18 (415–451) long; bulb 102 ± 7 (98–110) long, 100 ± 15 (82–110) wide. Nerve ring 266 ± 19 (250–287) and excretory pore 467 ± 85 (395–561) from anterior end. Eight longitudinal rows of equally spaced somatic papillae. Twelve pairs of caudal papillae: 4 pairs precloacal, 2 pairs anterior to sucker, 2 pairs lateral to sucker; 2 pairs adcloacal, 1 pair lateral, 1 sublateral; 6 pairs postcloacal, 2 pairs on truncation, pair adjacent to cloaca small, posterior pair largest of the papillae present; 4 pairs on caudal filament, 2 pairs lateral in position, 2 pairs ventral. Spicules, similar, equal, 246 ± 3 (244–250) long, proximal end truncate, distal end with blunt point; gubernaculum $169 \pm 9 (159-177)$ in length. Caudal end with well-developed, sclerified sucker, 33 ± 2 (31–34) in diameter, $35 \pm 2 (33-37)$ deep. Cloaca $124 \pm 12 (110-134)$ from posterior end of body; caudal filament conical, 78 ± 8 (68–84) in length, terminating in sharp point and supporting 4 pairs of the caudal papillae.

Female (allotype and 9 paratypes): Length 3.64 ± 0.47 mm (2.94–4.16), width at level of vulva 384 ± 52 . Narrow alae extending from base of lips to near middle of tail. Cuticle thick, fine transverse striations approximately 6 apart, equally spaced somatic papillae in longitudinal 8 rows. Lips 21 ± 3

(15–24) in length; pharynx 54 ± 4 (49–61) in length; corpus 601 ± 47 (537–671) in length; bulb 139 ± 12 (122–159) long, 150 ± 23 (116–183) wide. Nerve ring 293 ± 11 (275–305), excretory pore 542 ± 33 (485–599) from anterior end, respectively. Vulva, slightly post-equatorial, transverse slit, lips slightly salient, 1.91 ± 0.26 mm (1.60–2.27; 52–54% of body length) from anterior end. Ovijector muscular, directed posteriorly, joining 1 anteriorly and 1 posteriorly directed uterus. Uteri containing a small number of eggs. Eggs in ovijector with thick, punctate shell, cylindrical in profile, 82 ± 3 (79–85) × 30 ± 2 (27–34). Conical tail 350 ± 31 (299–415) in length, terminating in sharp point. Phasmids just posterior to middle of tail.

Type host: *Hylophorbus* cf. *rufescens* Macleay, 1878, red Mawatta frog; symbiotype, BPBM 16199; collected 21 August 2002, Oya Tabu.

Type locality: E. slope Oya Tabu, (9°27′S, 150°47′E, 1100 m), Fergusson Island, Milne Bay Province, Papua New Guinea.

Site of infection: Large intestine.

Type specimens: Holotype male, USNPC 99593; allotype female, USNPC 99594; paratypes USNPC 99595; voucher specimens USNPC 99598, BPBM H207.

Etymology: The new species is named for František Moravec, Institute of Parasitology, Academy of Sciences of the Czech Republic, in recognition of his contributions on the helminths of Papuan anurans and as the first to report (Moravec 1990) the presence of Spinicaudinae in amphibians.

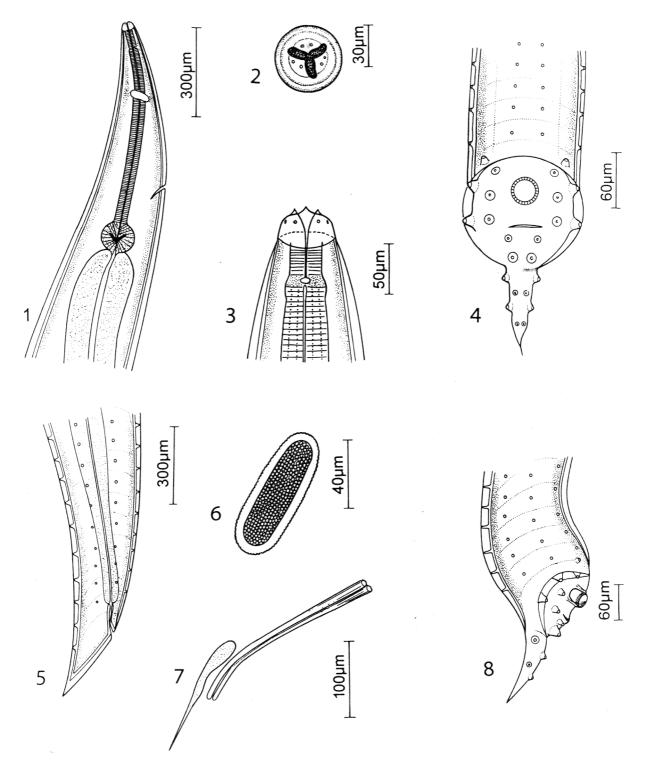
Remarks

Moravec (1990) collected 3 female specimens, which he assigned to Spinicaudinae, 2 from the large intestine of *Callulops humicola* (Zweifel) and 1 from *C. stictogaster* (Zweifel) (Microhylidae). Although these specimens are larger than ours, 4.73–6.85 mm vs. 2.94–4.16 mm, the ratio of distance of vulva to total body length is in agreement (50–52% vs. 52–54%). The only species of Spinicaudinae reported from New Guinea is *Moaciria chondropythonis* Gibbons, 1979, which has a much larger body size (Gibbons 1979) (see Table II). Three additional species, *Moaciria butleri* Jones, 1979, *M. etnae* Jones, 1979, and *M. sphenomorphi* Jones, 1979, are found in the Australo-Papuan region, but at this time are known only

from Australia and only from reptiles. Morphological characters of these 3 species are different from *M. moraveci* sp. nov. (see Table II).

Discussion

All helminths found during this study represent new host records for *Hylophorbus* cf. *rufescens*. *Cosmocerca novaegui*-



Figs 1–8. Moaciria moraveci sp. nov.: 1. Female, anterior extremity, lateral view. 2. Female, anterior extremity, en face view. 3. Female, anterior extremity, ventral view. 4. Male, posterior end, ventral view. 5. Female, posterior end, lateral view. 6. Egg. 7. Spicules and gubernaculum, lateral view. 8. Male, posterior end, lateral view

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Table II. Selected characters of males of species of Moaciria

Male characters	Length (mm)	Spicule (µm)	Gubernaculum (µm)	Sucker diameter (µm)	Caudal papillae
M. alvarengai Freitas, 1956	3.01–3.93	230–290	150–160	42–50	7 pairs, 3 single
M. butleri Jones, 1979	6.37	450	290	80	19 pairs, 2 single
M. chondropythonis Gibbons, 1979	10.89–12.06	633–912	524–599	117–152	15 pairs, 1 single
M. etnae Jones, 1979	4.95	350	220	60	16 pairs
M. freitasi Chabaud et Brygoo, 1960	4.2	260	150	25	11 pairs, 1 single
M. icosiensis (Seurat, 1917)	4.95	360	180	not stated	7 pairs
= A. pharyngeodentata Belle, 1957	3.9-4.5	380-420	180	_	11 pairs
M. komodoensis (Pinnell et Schmidt, 1977)	5.73	457	295	79	9 pairs
M. moraveci sp. nov.	2.59-2.85	244-250	159-177	31–34	12 pairs
M. sphenomorphi Jones, 1979	4.65-4.95	290-300	180-200	40	12 pairs, 1 single

Table II (continued). Selected characters of females of species of Moaciria

Female characters	Length (mm)	Vulva from anterior		Egg (µm)	Shell	Reference	
		mm	% body length				
M. alvarengai	4.12-5.49	1.91-2.41	44–46	76×42	smooth	Freitas (1956)	
M. butleri	7.12-9.52	_	44–47	84×56	smooth	Jones (1979)	
M. chondropythonis	11.73-16.25	not stated	_	none present	_	Gibbons (1979)	
M. etnae	6.37-7.35	_	44–47	87×59	smooth	Jones (1979)	
M. freitasi	undescribed	_	-	-	_	Chabaud and Brygoo (1960)	
M. icosiensis	7.20	3.66	51	85×56	not stated	Seurat (1917)	
= Aplectana pharyngeodentata	4.7 - 6.1	about mi	ddle of body	160×120	not stated	Belle (1957)	
M. komodoensis	undescribed	_	_	-	-	Pinnell and Schmidt (1977)	
M. moraveci sp. nov. M. sphenomorphi	2.94–4.16 undescribed	1.60–2.27	52–54 –	82 × 30 -	punctate –	present paper Jones (1979)	

neae was originally described by Moravec and Sey (1990) from specimens taken from the intestine of Platymantis papuensis Meyer (Ranidae) collected in the Torricelli Mountains, Sepik Province, Papua New Guinea and was previously known only from that host. Cosmocerca tyleri was originally described by Bursey et al. (2006) from specimens taken from the large intestine of Genyophryne thomsoni Boulenger (Microhylidae) collected in the nearby Cloudy Mountains, Milne Bay Province, Papua New Guinea and was previously known only from that host. Oswaldocruzia bakeri was originally described by Moravec and Sey (1986) from specimens taken from Phrynomantis stictogaster (currently, Callulops stictogaster) (Microhylidae) collected in the Eastern Highlands Province of Papua New Guinea. It is also known from the frogs Callulops humicola, C. wilhelmanus (Loveridge) and Platymantis pelewensis Peters and the lizards Cyrtodactylus louisiadensis (de Vis) and Sphenomorphus jobiensis (Meyer) (Moravec and Sey 1986; Bursey and Goldberg 2004; Bursey et al. 2005a, b). The thick stomach worm Physocephalus sexalatus (Molin, 1860), a cosmopolitan species, is a common parasite of wild and domestic pigs and is occasionally found in tapirs, equines, bovines, and lagomorphs (Anderson 2000). Infective larvae have been recovered from at least 20 different genera of beetles, and encapsulated larvae have been found in amphibians, reptiles, birds, and mammals that typically feed on beetles (Anderson 2000). Third-stage larvae are characterized by a digitiform knob at the tip of the tail (Alicata 1935). To our knowledge this is the first report of *Physocephalus* in Papua New Guinea and *H. cf. rufescens* may be an accidental host. The life cycle of acanthocephalans generally requires 2 hosts; an arthropod intermediate host and an avian or mammalian definitive host (Near 2002). Should an inappropriate host eat the infected arthropod, the cystacanth excysts then migrates from the digestive tract into the body cavity and again encysts to await an appropriate host. *Hylophorbus* cf. *rufescens* most likely serves as a paratenic host for acanthocephalans.

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