

### Central European Journal of Medicine

# Increased oxidative stress status in rat serum after five minutes treadmill exercise

#### Research Article

Florin-Petrut Trofin<sup>1</sup>, Alin Ciobica<sup>1,2</sup>, Dumitru Cojocaru<sup>1</sup>, Marin Chirazi<sup>1</sup>, Cezar Honceriu<sup>1</sup>, Laurentiu Trofin<sup>3</sup>, Dragomir Serban\*<sup>4</sup>, Daniel Timofte<sup>¥4</sup>, Sabina Ioana Cojocaru<sup>¥1</sup>, Emil Anton<sup>¥4</sup>

 $^{
mathcal{T}}$ These authors equally contributed to this work (as last and coordinating author).

1 "Alexandru Ioan Cuza" University, Iasi, 700506, Romania

2 Center of Biomedical Research of the Romanian Academy, Iasi Branch, Romania

3 Colegiul Tehnic "Ion Creanga", Tg. Neamt, Romania

4 "Gr. T. Popa" University of Medicine and Pharmacy, 700115, Iasi, Romania

#### Received 1 June 2013; Accepted 10 January 2014

Abstract: Although it is accepted that an important correlation exists between the physical exercise and the oxidative stress status, the data regarding the levels of the main oxidative stress markers after physical training have been difficult to interpret and a subject of many controversies. There are also very few studies regarding the effects of short-time exercise on the oxidative stress status modifications. Thus, in the present report we were interested in studying the modifications of some oxidative stress markers (two antioxidant enzymes- superoxide dismutase and glutathione peroxidase, a lipid peroxidation parameter - malondyaldehide, the total antioxidant status and protein carbonyl levels), from the serum of rats that were subject to one bout of five minutes exercise on a treadmill, when compared to a control sedentary group. In this way, we observed a decrease of superoxide dismutase specific activity in the rats which performed the exercises. Still, no modifications of glutathione peroxidase specific activity were found between groups. In addition, increased levels of malondyaldehide and protein carbonyls were observed in the rats subjected to exercises. In conclusion, our data provides new evidence regarding the increase of the oxidative stress status, as a result of a 5-minutes bout of treadmill exercising in rats, expressed through a decrease in the SOD specific activity and the total antioxidant status and also an increase of the lipid peroxidation and protein oxidation processes.

**Keywords:** Exercise • Rat • Oxidative stress • Treadmill

© Versita Sp. z o.o

## 1. Introduction

Today is well established that there is an important correlation between physical exercise and the oxidative stress process, which is defined as the imbalance between the organism antioxidant capacity on one hand and the overproduction of the reactive oxygen species (ROS) on the other hand [1].

In this way, there are previous reports describing a direct relationship between the muscle oxidative stress status and the isometric force production [2]. Thus, it seems that a perfectly balanced equilibrium between ROS production and the enzymatic/non-enzymatic antioxidants will result in an ideal skeletal muscle force [3,4].

However, while some authors demonstrated an increase in the oxidative stress status as a result of physical exercise [4-6], there are also reports describing a so-called adaptation of the antioxidant system to physical exercise and training [7], which will result in an increase of the various types of antioxidant defenses.

These aspects led to numerous contradictions in the current literature regarding the levels of some oxidative stress markers after physical training. In this way, the swimming effort in rats has been reported to result in a decrease of glutathione (GSH) levels [8], which is an important antioxidant, while other studies showed that some different physical exercise generated increased levels of plasma GSH [9,10] or increased GSH concentration in several muscles after endurance exercises in dogs [11]. Similar aspects were also reported in the case of superoxide dismutase-SOD (the first line of defense against the nocive effects of ROS), which was showed to express both increased [12-14] and decreased [15, 16] specific activity after the physical exercise was performed, as well as in the cases of the glutathione peroxidase - GPX [17,18] and catalase - CAT [7,19], the other two important antioxidant enzymes.

Still, there are very few studies regarding the effects of very short-time exercise on the oxidative stress status modifications. In the present study we were interested to see the modifications of some oxidative stress markers (two antioxidant enzymes- superoxide dismutase and glutathione peroxidase, a lipid peroxidation parameter – malondyaldehide, the Total Antioxidant Status-TAS and protein carbonyl levels), from the serum of rats that were subject to one bout of 5 minutes exercise on a treadmill. In fact, according to the best of our knowledge, this is the first time when these markers have been determined after only one bout of 5 minutes exercise.

### 2. Material and methods

#### 2.1. Animals

Adult male Wistar (n=14) rats, weighing 200-250 g, were kept in a room with controlled temperature (22°C) and a 12:12-h light/dark cycle (starting at 08:00 h), with food and water ad libitum. The animals were treated in accordance with the guidelines of animal bioethics from the Act on Animal Experimentation and Animal Health and Welfare Act from Romania and all procedures were in compliance with the European Communities Council Directive of 24 November 1986 (86/609/EEC). This study was approved by the local Ethics Committee and also efforts were made to minimize animal suffering and to reduce the number of animals used.

#### 2.2. Experimental design

All the rats (n=14) were acclimatized to the experimental room for 1 week before the exercise testing.

They performed one bout of driven rodent treadmill running 5 minutes in one day, 0% grade, at 10:00 am.

The rats were divided into 2 groups of 7 rats each: a control (sedentary) group and one group which performed the physical exercise. In order to obtain a minimal difference in the experimental manipulation of controls, the sedentary rats were placed for the same amount of time (5 minutes) on the switched-off treadmill.

Immediately after that the rats were anesthetized with pentobarbital and sacrificed. Also, the age-matched control animals, which were kept sedentary in their cages, were sacrificed using the same procedure. All animals were fasted for 12 h before death. After the blood was taken, it was allowed to clot and centrifuged for 15 min at 3000 rpm. Serum was then aliquoted into plastic tubes and stored at −40°C until measurement.

### 3. Biochemical estimations

#### 3.1. Determination of SOD

Superoxide dismutase (SOD) activity was measured by the percentage reaction inhibition rate of enzyme with WST-1 substrate (a water soluble tetrazolium dye) and xanthine oxidase using a SOD Assay Kit (FLUKA, 19160) according to the manufacturer's instructions. Each endpoint assay was monitored by absorbance at 450 nm (the absorbance wavelength for the colored product of WST-1 reaction with superoxide anions) after 20 minutes of reaction time at 37°C. The percent inhibition was normalized by mg protein and presented as SOD activity units.

#### 3.2. Determination of GPX

The glutathione peroxidase (GPX) activity was measured using the GPX cellular activity assay kit CGP-1 (SIGMA). This kit uses an indirect method, based on the oxidation of glutathione (GSH) to oxidized glutathione (GSSG) catalyzed by GPX, which is then coupled with recycling GSSG back to GSH utilizing glutathione reductase (GR) and NADPH. The decrease in NADPH at 340 nm during oxidation of NADPH to NADP is indicative of GPX activity.

Regarding the units of activity for the investigated enzymes we used units/ml = micromol/min/ml.

#### 3.3. Total antioxidant status

TAS was assayed with a chemiluminometric method, with luminol – horseradish peroxidase system (Berthold Lumat 9507 chemiluminometer). In this method,

constant light emission result from luminol degradation in the presence of a catalyst (horseradish peroxidase) with an enhancer (p-iodo-phenol) and is kinetically recorded. When a biological fluid is introduced into this system, the level of light emission decreases for a period of time proportional to the total antioxidant capacity. Trolox (6-hydroxy-2,5,7,8-tetramethylchroman-2-carboxylic acid), a water-soluble alpha-tocopherol analogue, was used as the standard. Calibration was made with Trolox (hydro-soluble vitamin E) and final results are related to Trolox equivalents. The prooxidant system which generates light, was calibrated to five million relative units of light (RLU) and serum samples were used at a dilution of 1/10.

#### 3.4. Determination of malondialdehyde

MDA levels were determined by thiobarbituric acid reactive substances (TBARs) assay. 200  $\mu$ L of serum were added and briefly mixed with 1 ml of trichloroacetic acid at 50%, 0.9 ml of TRIS-HCl (pH 7.4) and 1 ml of thiobarbituric acid 0.73%. After vortex mixing, samples were maintained at 100 °C for 20 minutes. Afterward, samples were centrifuged at 3000 rpm for 10 min and supernatant was read at 532 nm. The signal was read against an MDA standard curve, and the results were expressed as nmol/ml [20,21].

Total protein was measured using Bradford dyebinding method, with bovine serum albumin as the standard [22].

#### 3.5. Protein carbonyl determination

Plasma proteins carbonyl content was measured by forming labeled protein hydrazones derivates, using 2, 3-dinitrophenylhydrazine (DNPH), which were then quantified spectrophotometrically at 360 nm, using a molar absorption coefficient for hidrazones of 21000 mol/L.cm.

### 4. Data analysis

The results were statistically analyzed by using Student's t-test (two tailed, unpaired). All results are expressed as mean  $\pm$  SEM. P<0.05 was regarded as statistically significant.

#### 5. Results

We report here a significant decrease (p=0.01) for the specific activity of SOD, the first antioxidant enzyme in

the way of ROS, in the exercised group, as compared to the sedentary rats (Figure 1).

However, we did not find any significant modifications (p=0.34) between the group of rats which was subjected to the 5 minutes exercise bout, when compared to the sedentary control group, in the case of GPX specific activity (Figure 2).

Asignificant decrease (p=0.02) was found for the total antioxidant status in the serum of rats which performed the physical exercise, as compared to the sedentary control rats (Figure 3).

Additionally, we observed a significant increase in the lipid peroxidation processes, as demonstrated by the increased levels of MDA (p=0.03) from the serum of exercised rats, when compared to the sedentary controls (Figure 4).

Moreover, we showed a significant increase (p=0.03) in the serum protein oxidation markers (as assessed by the levels of protein carbonyls) in the exercised rats, when compared to the sedentary control group (Figure 5).

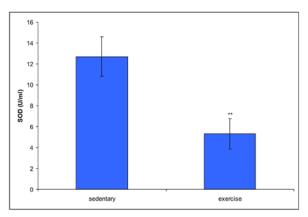


Figure 1. The effects of five minutes treadmill exercise on SOD specific activity from the rat serum. The values are mean ± SEM (n=7 animals per group). \*\*p=0.01 vs. sedentary-control group.

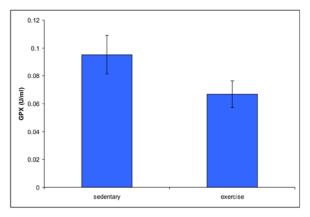
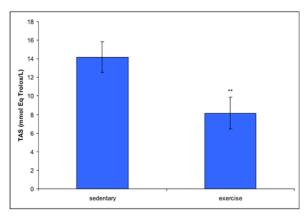


Figure 2. The effects of five minutes treadmill exercise on GPX specific activity from the rat serum. The values are mean ± SEM (n=7 animals per group).



**Figure 3.** The effects of five minutes treadmill exercise on total antioxidant status (TAS) from the rat serum. The values are mean ± SEM (n=7 animals per group). \*\*p=0.02 vs. sedentary-control group.

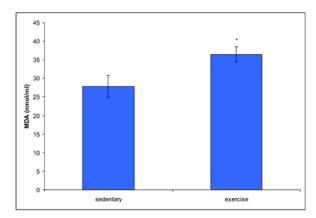


Figure 4. The effects of five minutes treadmill exercise on malondyaldehide (MDA) levels from the rat serum. The values are mean ± SEM (n=7 animals per group). \*p=0.03 vs. sedentary-control group.

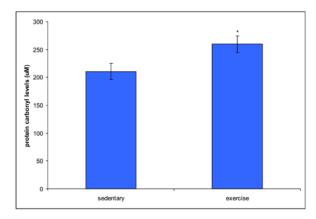


Figure 5. The effects of five minutes treadmill exercise on the levels of protein carbonyls from the rat serum. The values are mean ± SEM (n=7 animals per group). \*p=0.03 vs. sedentary-control group.

### 6. Discussion

The results we presented here provide additional evidence regarding the increase in the oxidative stress status as a result of short-time physical exercise and expressed through a decrease in the SOD specific activity and the total antioxidant status. There was also an increase of the lipid peroxidation and protein oxidation processes, when compared to the sedentary control group. As mentioned, according to the best of our knowledge, this is the first time when these markers are determined after only one bout of 5 minutes exercise.

Similar results regarding the possible increase of the oxidative stress processes as a result of exercise have been reported as early as 1978 [23]. These results were confirmed by additional studies performed after that, such as the one of Barclay et al. that showed that free radicals which result after the contraction will generate muscular fatigue [5] or the one of Kondo that proposed the theory regarding the very important role of ROS in the degenerative processes resulting in muscular mass loss [24].

However, as we previously mentioned, there are on the other side. reports describing the possibility that the organism may generate increased concentrations of antioxidants, as a result of physical exercise, in order to cope with the new generated oxidative stress [7,25].

In this way, as we initially mentioned, in the case of SOD a significant increase of its specific activity was reported after performing some various physical exercises [26-28]. However, when other research groups performed similar testing the results could not be easily reproduced [19,25]. Moreover, in the case of CAT, another antioxidant enzyme, very different results were reported after physical training, ranging from increased specific enzymatic activity [29,30], to no modifications at all [7] or even decreased activity [19]. Similar aspects were also reported in the case of GPX [18,31].

These facts could be explained through the differences in the antioxidant enzymes isovariations, different methods in assaying the enzymatic activity (e.g spectrophotometric vs. chemiluminometric), and in some cases the different types of exercise used and the muscles that are involved in the specific types of efforts [7]. Additionally, it is important to mention that the way exercise influences the oxidative stress status, depends also on the level of training, as was demonstrated by Senturk et al. in 2001, which showed that exercise-induced oxidative stress affects erythrocytes only in sedentary rats but not exercise-trained rats [32].

Moreover, it was stated that the antioxidant enzymes could have very different patterns of the aforementioned

adaptation to physical exercises [7]. Also, it is important to keep in mind that low levels of ROS are indeed required for normal force production in the skeletal muscles [4].

Regarding the time of the physical training in our experiment we decided to use a short-time of exercise performing (5 minutes), considering that to the best of our knowledge, this is the first study using only a 5 minute set of exercise. Also, we did not find in the literature any previous use of the TAS as a parameter of oxidative stress status, after performing physical exercise.

Additionally, considering the previous described variety of results for the oxidative stress markers and their different adaptations to the performance of exercise, we decided to use TAS as a parameter, together with the specific activity of SOD and GPX, also considering that it covers all aspects of antioxidant defending capacity (e.g. not a single enzymatic/non- enzymatic antioxidant). In this way, we obtained a significant decrease of SOD specific activity and also a decrease in the Total Antioxidant Status. Still, no significant modifications of GPX were observed. This could be perhaps explained by the fact that SOD is most sensitive to and the primary defense against ROS damage [1], exhibiting prompt compensation processes when encountering heavy oxidative injury. Therefore it is possible for the compensatory modifications of GPX to be less sensible, as compared to SOD.

Also, we observed an increase in protein carbonylation processes which are the most frequent type of protein modification in response to oxidative stress and is thought to be irreversible and destined only to induce protein degradation in a nonspecific manner [1].

Of course, all of these aspects regarding the importance of the oxidative stress status in the modifications that appear after the physical training, led to the idea of using various antioxidants as a possible solution for longer physical training or shorter recovery periods.

In this way, probably the most tested antioxidant in this area of research was represented by vitamin E, which was showed to reduce and prevent some specific tissue-damage during exercise [33]. I It also seems that the lack of vitamin E increases the risk of contraction-induced membrane damage in the muscle [34]. Also, it was demonstrated that vitamin E could delay the onset of muscular fatigue [35]. Additionally N-acetylcysteine (NAC), which is also a very well recognized antioxidant, was also reported to prevent muscle fatigue in some experimental muscle preparations [36].

However, most of the studies regarding the positive effects of the antioxidants in this area of research are controversial, since some authors showed for example that both vitamin C and E did not affect in any way the performance of the exercise [4,37]. Different results could be perhaps explained by the maximal/submaximal intensity of the physical exercise, the dosage of antioxidants, as well as the time of administration.

In fact, similar contradictory aspects were also described in terms of the effects for various antioxidants on the recovery time after the effort, with reports about some positive effects [38] in this matter, as well as with authors stating no effects at all [4,39].

In this way, future studies will need to carefully determine which antioxidants and in what dosages do they have the maximal therapeutic benefit.

There are also several limitations to our study that should be discussed. First of all, the usage of pentobarbital as an anesthetic before the killing of the animals, considering that this reagent was previously suggested to influence the levels of some oxidative stress markers [40,41]. However, pentobarbital was injected in both our study groups (the sedentary and the exercised one).

Another aspect could be represented by the relatively small number of animals we used.

### 7. Conclusions

The results presented here provide additional evidence regarding the increase in the oxidative stress status as a result of a 5 minute bout of treadmill exercising and expressed through a decrease in the SOD specific activity and the total antioxidant status and also an increase of the lipid peroxidation and protein oxidation processes.

## **Acknowledgments**

The authors would also like to show their gratitude to the reviewers of this paper which significantly improved the value of the presented data by adding very important insights, comments, and suggestions. Florin Trofin is supported by a POSDRU/159/1.5/S/133675 grant.

### **Conflict of interest statement**

Authors state no conflict of interest.

#### References

- Sies H. Oxidative stress: oxidants and antioxidants. Experimental Physiology 1997; 82: 291–295
- [2] Reid MB, Khawli FA, Moody MR. Reactive oxygen in skeletal muscle. III. Contractility of unfatigued muscle. J Appl Physiol 1993; 75: 1081–1087
- [3] Reid MB. Redox modulation of skeletal muscle contraction: what we know and what we don't. J Appl Physiol 2001; 90: 724–731
- [4] Powers SK, Jackson MJ. Exercise-induced oxidative stress: cellular mechanisms and impact on muscle force production. Physiol Rev 2008; 88: 1243–1276
- [5] Barclay JK, Hansel M. Free radicals may contribute to oxidative skeletal muscle fatigue. Can J Physiol Pharmacol 1991; 69: 279–284
- [6] Reid MB, Shoji T, Moody MR, Entman ML. Reactive oxygen in skeletal muscle. II. Extracellular release of free radicals. J Appl Physiol 1992; 73: 1805–1809
- [7] Ji LL. Antioxidants and Oxidative Stress in Exercise. Exp Biol Med 1999; 222: 283–292
- [8] Leichtweis S, Leeuwenburgh C, Fiebig R, Parmelee D, Yu XX, Ji LL. Rigorous swim training deteriorates mitochondrial function in rat heart. Acta Physiol Scand 1997; 160: 139–148
- [9] Evelo CT, Palmen NG, Artur Y, Janssen GM. Changes in blood glutathione concentrations, and in erythrocyte glutathione reductase and glutathione S-transferase activity after running training and after participation in contests. Eur J Appl Physiol 1992; 64: 354–358
- [10] Robertson JD, Maughan RJ, Duthie GG, Morrice PC. Increased blood antioxidant systems of runners in response to training. Clin Sci 1991; 80: 611–618
- [11] Marin E, Kretzschmar M, Arokoski J, Hanninen O, Klinger W. Enzymes of glutathione synthesis in dog skeletal muscle and their response to training. Acta Physiol Scand 1993; 147: 369–373
- [12] Ohno H, Suzuki K, Fujii J, Yamashita H, Kizaki T, Oh-ishi S, Taniguchi N. Superoxide dismutases in exercise and disease. Exercise and Oxygen Toxicity 1994; 1: 127–161
- [13] Ji LL, Dillon D, Wu E. Alteration of antioxidant enzymes with aging in rat skeletal muscle and liver. Am J Physiol 1990; 258: 918–923
- [14] Powers SK, Criswell D, Lawler J, Ji LL, Martin D, Herb R, Dudley G. Influence of exercise intensity and duration on antioxidant enzyme activity in skeletal muscle differing in fiber type. Am J Physiol 1994; 266: 375–380

- [15] Tiidus PM, Pushkarenko J, Houston ME. Lack of antioxidant adaptation to short-term aerobic training in human muscle. Am J Physiol 1996; 271: 832–836
- [16] Ji LL. Antioxidant enzyme response to exercise and aging. Med Sci Sports Exerc 1993; 25: 225–231
- [17] Leeuwenburgh C, Ji LL. Alteration of glutathione and antioxidant status with exercise in unfed and refed rats. J Nutr 1996; 126: 1833–1843
- [18] Leeuwenburgh C, Ji LL. Glutathione depletion in rested and exercised mice: Biochemical consequence and adaptation. Arch Biochem Biophys 1995; 316: 941–949
- [19] Laughlin MH, Simpson T, Sexton WL, Brown OR, Smith JK, Korthuis RJ. Skeletal muscle oxidative capacity, antioxidant enzymes, and exercise training. J Appl Physiol 1990; 68: 2337–2343
- [20] Ciobica A, Olteanu Z, Padurariu M, Hritcu L. The effects of pergolide on memory and oxidative stress in a rat model of Parkinson's disease. J Physiol Biochem 2012; 68: 59-69
- [21] Ciobica A, Hritcu L, Nastasa V, Padurariu M, Bild W. Inhibition of central angiotensin converting enzyme exerts anxiolytic effects by decreasing brain oxidative stress. Journal of Medical Biochemistry 2011; 30: 109–114
- [22] Gurzu C, Artenie V, Hritcu L, Ciobica A. Prenatal testosterone improves the spatial learning and memory by protein synthesis in different lobes of the brain in the male and female rat. Cent. Eur. J. Biol 2008; 3: 39–47
- [23] Dillard CJ, Litov RE, Savin WM, Dumelin EE, Tappel AL. Effects of exercise, vitamin E, ozone on pulmonary function and lipid peroxidation. J Appl Physiol 1978; 45: 927–932
- [24] Kondo H, Miura M, Itokawa Y. Antioxidant enzyme systems in skeletal muscle atrophied by immobilization. Pflugers Arch 1993; 422: 404–406
- [25] Alessio HM, Goldfarb AH. Lipid peroxidation and scavenger enzymes during exercise: Adaptive response to training. J Appl Physiol 1988; 64: 1333–1336
- [26] Higuchi M, Cartier LJ, Chen M, Holloszy JO. Superoxide dismutase and catalase in skeletal muscle: Adaptive response to exercise. J Gerontol 1985; 40: 281–286
- [27] Powers SK, Criswell D, Lawler J, Ji LL, Martin D, Herb R, Dudley G. Influence of exercise intensity and duration on antioxidant enzyme activity in skeletal muscle differing in fiber type. Am J Physiol 1994; 266: 375–380

- [28] Leeuwenburgh C, Fiebig R, Chandwaney R, Ji LL. Aging and exercise training in skeletal muscle: Response of glutathione and antioxidant enzyme systems. Am J Physiol 1994; 267: 439–445
- [29] Hollander J, Fiebig R, Gore M, Bejma J, Ohno H, Ji LL. Superoxide dismutase gene expression: Fiberspecific adaptation to endurance training. Am J Physiol 1999; 277: 856–862
- [30] Oh-Ishi S, Kizaki T, Nagaswa J, Izawa T, Komabayashi T, Nagata N, Suzuki K, Taniguchi N, Ohno H. Effects of endurance training on superoxide dismutase activity, content, and mRNA expression in rat muscle. Clin Exp Pharmacol Physiol 1997; 24: 326–332
- [31] Ji LL, Fu RG. Responses of glutathione system and antioxidant enzymes to exhaustive exercise and hydroperoxide. J Appl Physiol 1992; 72: 549–554
- [32] Sentürk UK, Gündüz F, Kuru O, Aktekin MR, Kipmen D, Yalçin O et al. Exercise-induced oxidative stress affects erythrocytes in sedentary rats but not exercise-trained rats. J Appl Physiol 2001; 91: 1999–2004
- [33] Jackson MJ, Khassaf M, Vasilaki A, McArdle F, McArdle A. Vitamin E and the oxidative stress of exercise. Ann NY Acad Sci 2004; 1031: 158–168
- [34] Jackson MJ, Pye D, Palomero J. The production of reactive oxygen and nitrogen species by skeletal muscle. J Appl Physiol 2007; 102: 1664–1670

- [35] Novelli GP, Bracciotti G, Falsini S. Spin-trappers and vitamin E prolong endurance to muscle fatigue in mice. Free Radic Biol Med 1990; 8: 9–13
- [36] Shindoh C, DiMarco A, Thomas A, Manubay P, Supinski G. Effect of N-acetylcysteine on diaphragm fatigue. J Appl Physiol 1990; 68: 2107–2113
- [37] Powers SK, DeRuisseau KC, Quindry J, Hamilton KL. Dietary antioxidants and exercise. J Sports Sci 2004; 22: 81–94
- [38] Diaz PT, Costanza MJ, Wright VP, Julian MW, Diaz JA, Clanton TL. Dithiothreitol improves recovery from in vitro diaphragm fatigue. Med Sci Sports Exerc 1998; 30: 421–426
- [39] Khawli FA, Reid MB. N-acetylcysteine depresses contractile function and inhibits fatigue of diaphragm in vitro. J Appl Physiol 1994; 77: 317–324
- [40] Yesilkaya A, Ertug Z, Yegin A, Melikoglu M, Baskurt OK. Deformability and oxidant stress in red blood cells under the influence of halothane and isoflurane anesthesia. Gen Pharmacol. 1998; 31: 33–36
- [41] Kotzampassi K, Kolios G, Manousou P, Kazamias P, Paramythiotis D, Papavramidis TS et al. Oxidative stress due to anesthesia and surgical trauma: importance of early enteral nutrition. Mol Nutr Food Res. 2009; 53: 770–779