

# Responses of the Lichen *Cladonia convoluta* to High CO<sub>2</sub> Level and Heavy Metal Treatment

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Despite of the downward acclimation of photosynthesis in *C. convoluta*, increased net photosynthesis and carbon balance can be anticipated in response to elevated atmospheric CO<sub>2</sub> level. CO<sub>2</sub> exchange measurement seems to be more indicative when detecting heavy metal stress than fluorescence parameters. Among these, the relative fluorescence decrease ratio (RFd690) shows damage first, suggesting that the primary attack site for heavy metal ions is CO<sub>2</sub> fixation and reaction centres are harmed last. Long-term elevated CO<sub>2</sub> ameliorates partly this damage by improving C-balance to a greater extent in the heavy-metal stressed lichens.

## Introduction

Atmospheric CO<sub>2</sub> concentration is estimated to double from pre-industrial levels by the middle of the next century. Its effects on plants are under intense investigation. Being the key substrate of photosynthesis, most of the research focuses on assimilation and primary production under the predicted scenarios. Photosynthesis can show upward (Arp and Drake, 1991) or downward (Jarvis, 1993) regulation in response to elevated CO<sub>2</sub> level. Decreases in assimilation capacity in downward regulated plants are usually due to decreases in rubisco level and/or activity (Sicher *et al.*, 1994). Studies on lichens are extremely scarce in this context. Despite of marked downward regulation of CO<sub>2</sub> fixation increased lipid storage could be found in *Parmelia sulcata* (Balaguer *et al.*, 1996). Tuba *et al.* (1998) found that short-term elevated CO<sub>2</sub> level increases carbon gain during desiccation in *Cladonia convoluta*.

Interactions of elevated CO<sub>2</sub> concentration with different abiotic stress factors have been widely investigated. It can help plants to cope with drought (Scarasciamugnozza *et al.*, 1996; Arp *et al.*, 1998) mostly by decreasing stomatal conductance, with heat stress (Wayne *et al.*, 1998), with salinity (Reuveni *et al.*, 1997) and even with air pollutants like SO<sub>2</sub> (Lee *et al.*, 1997) or O<sub>3</sub> (McKee *et al.*, 1995). There are some reports, which provide evi-

dence that increased CO<sub>2</sub> concentration may act at the antioxidant system. It decreases intrinsic oxidative stress (Schwanz *et al.*, 1996) and strengthens acclimation at the onset of stress (Polle *et al.*, 1997).

Although lichens are used to biomonitor aerial dispersion of heavy metals (Garty *et al.*, 1997) using their ability to tolerate elevated quantities of heavy metals (Nash, 1975) and the heavy metal content of *Cladonia convoluta* correlated with soil contamination as well (Chettri *et al.*, 1997), heavy metals are by no means harmless to them. These ions denature enzymes and block their functional groups as direct toxic effects. Heavy metals also decrease chlorophyll content especially at low pH (Garty *et al.*, 1992). Besides this they also induce free radical toxicity (Gadd, 1993). Although there has been an extensive research done on the effect of heavy metals on lichens, it has not been investigated under the future high CO<sub>2</sub> environment yet. For this purpose we chose two metals with different toxicity (Wells and Brown, 1995) but having a similarly high importance in environment quality.

In this work we aimed to reveal the effects of the long-term elevated CO<sub>2</sub> level on the photosynthetic responses of the highly desiccation tolerant lichen *C. convoluta*, because lichens have rarely been the targets of such investigations. We also intended to determine whether elevated CO<sub>2</sub> level

ameliorates heavy-metal induced damage of this already highly stress-tolerant lichen species.

## Materials and Methods

The lichen *Cladonia convoluta* (Lam.) P. Cout. was collected in *Festuca vaginata* grassland near Fülöpháza, Hungary (19 ° 14' E; 47 ° 30' N, some 70 km SSE of Budapest at 130 m a.s.l.).

Lichens with their original soil substrates were transplanted into boxes (three replicates) and put in Plexiglas chambers (one chamber for the fumigation treatment and one for the control) situated in the botanical garden in Gödöllő, some 30 km NE of Budapest. High-CO<sub>2</sub> (700 µl l<sup>-1</sup>) treatment was maintained as described in previous studies (Tuba *et al.*, 1995). Since the chambers were closed on the top, plants had to be sprayed every morning with [1] deionized water (control); [2] 350 µmol l<sup>-1</sup> Pb<sup>2+</sup>; [3] 350 µmol l<sup>-1</sup> Cd<sup>2+</sup> or [4] 175 µmol l<sup>-1</sup> Pb<sup>2+</sup> + 175 µmol l<sup>-1</sup> Cd<sup>2+</sup> (combined treatment). Metals were given as nitrate salts. Exposure lasted for one month.

The measurements were made on the upper young parts of the lichen thallus. To ensure full photosynthetic activity the samples were kept close to the optimum water contents (130–150% of DM) for 24 h at a photosynthetic photon flux density (PPFD) of 400 µmol m<sup>-2</sup> s<sup>-1</sup> at 20 °C before all gasexchange and chlorophyll-fluorescence measurements on fully hydrated material. Chlorophyll fluorescence was recorded with a modulated one-channel Hansatech fluorometer. We calculated the maximal photosystem (PS)II quantum efficiency [ $F_v/F_m = (F_m - F_o)/F_m$ ] and vitality index, or relative fluorescence decrease ratio measured at 690 nm [ $RFd = (F_m - F_s)/F_s$ ] (Lichtenhaler, 1988). Dark respiration and light-saturated net photosynthesis rates were measured using an IRGA system (Type LCA2, ADC Co. Ltd., Hoddesdon, U.K.), operated in differential mode at a PPFD of 400 µmol m<sup>-2</sup> s<sup>-1</sup> (see Tuba *et al.*, 1996). The CO<sub>2</sub> concentrations of 350 and 700 µmol mol<sup>-1</sup> for the IRGA measurements were produced by a gas diluter (GD 600, ADC Co. Ltd., Hoddesdon, U.K.) (see Tuba *et al.*, 1998). Element analysis (Al, Cd, Cr, Cu, Fe, Ni, Pb, V, Zn) was carried out with ICP-AES technique preceded by dry cleaning and high pressure and temperature digestion of the plant material with 1:1 (v/v) ratio of concentrated

HNO<sub>3</sub> and concentrated H<sub>2</sub>O<sub>2</sub> (Tuba and Csintalan, 1993). Measurements were performed in three replicates except for the element content data where we had five samples digested and analyzed.

## Results and Discussion

CO<sub>2</sub>-treatment did not cause significant differences in heavy metal content. Pb content was raised from 13.4±3.6 to 1274±49 µmol g<sup>-1</sup> dwt, Cd content from 1.13±1.01 to 581±31 µmol g<sup>-1</sup> dwt by the corresponding treatments. Simultaneous treatment with the half dose of the two metals resulted in 692±55 µmol g<sup>-1</sup> dwt Pb and 343±19 µmol g<sup>-1</sup> dwt Cd concentration. Berg *et al.* (1995) noted that the uptake efficiency for Cd is 40–65% of that for Pb in the case of mosses. *C. convoluta* fell within this range with 45.6% or 49.6% at the combined treatment. Heavy metal treatments reduced Ca-content by 18.6% ( $p = 3.78 \cdot 10^{-6}$ ) and K-content by 10.4% ( $p = 7.74 \cdot 10^{-4}$ ). K-loss is a generally used measure of heavy metal toxicity, however inhibition of photosynthesis in small poikilohydric plants can occur without membrane leakage in Cd-treatment (Brown and Wells, 1990). Decreased Ca-content can be explained by its displacement by Cd from the extracellular binding sites (Brown and Beckett, 1985). When lichens are immersed in a solution of metal ions uptake is nearly complete within an hour (Nieboer *et al.*, 1976). Since metals were given as dissolved ions and drying out lasted at least four hours in the cool autumn weather, we do not assume any errors due to significant amount of externally stored heavy metal containing particles. We can neither tell the ratio of cell wall-bound ions nor the ratio of ions taken up by the two different symbionts. However, Brown and Sidhu (1992) suggest that the cell wall does not prevent but only delays heavy metal induced impairments in mosses.

The chlorophyll fluorescence parameters did not show any acclimation to the elevated CO<sub>2</sub> concentration at the photochemistry level (Table I). PSII activity (presented by  $F_v/F_m$ ) did not reflect any change due to the heavy metal treatments either. According to Atal *et al.* (1991) water splitting is more harmed by Cd than PSII. Hence one can obtain significant  $F_v/F_m$  values without prolonged electron supply from water.  $RFd_{690}$ , however, which corresponds to the whole thylakoid

Table I. Fv/Fm and RFd690 values of *Cladonia convoluta* grown at present (350  $\mu\text{mol mol}^{-1}$ ) and future elevated (700  $\mu\text{mol mol}^{-1}$ ) CO<sub>2</sub> concentration and different heavy metal treatments. control; Pb: 350  $\mu\text{mol l}^{-1}$  Pb(NO<sub>3</sub>)<sub>2</sub>; Cd: 350  $\mu\text{mol l}^{-1}$  Cd(NO<sub>3</sub>)<sub>2</sub>; Pb+Cd: 175  $\mu\text{mol l}^{-1}$  Pb(NO<sub>3</sub>)<sub>2</sub> + 175  $\mu\text{mol l}^{-1}$  Cd(NO<sub>3</sub>)<sub>2</sub>.

| CO <sub>2</sub> level    |         | Fv/Fm | st.dev. | RFd690 | st.dev. |
|--------------------------|---------|-------|---------|--------|---------|
| Present CO <sub>2</sub>  | Control | 1.184 | 0.096   | 0.581  | 0.058   |
|                          | Pb      | 0.995 | 0.098   | 0.580  | 0.026   |
|                          | Cd      | 1.024 | 0.202   | 0.562  | 0.035   |
|                          | Pb+Cd   | 0.903 | 0.070   | 0.569  | 0.043   |
| Elevated CO <sub>2</sub> | Control | 1.120 | 0.057   | 0.548  | 0.063   |
|                          | Pb      | 0.985 | 0.115   | 0.571  | 0.023   |
|                          | Cd      | 0.953 | 0.111   | 0.533  | 0.035   |
|                          | Pb+Cd   | 0.837 | 0.046   | 0.534  | 0.023   |

membrane activity being influenced by CO<sub>2</sub> fixation (Lichtenthaler, 1988) was slightly lowered by the heavy metal treatments. This supports Krupa and Baszynski's view (1995) that heavy metals harm Calvin cycle first. This decrease was significant only in case of the combined (Pb + Cd) treatments.

The CO<sub>2</sub> exchange rates changed in response to CO<sub>2</sub>- and heavy metal treatments. Since we aimed

to investigate the long-term effects of elevated CO<sub>2</sub> level to control and heavy metal stressed plants, they had to be measured at their own exposure CO<sub>2</sub> concentration. From this aspect, short-term effects presented by the plants grown at 350 and measured under 700 ppm CO<sub>2</sub> are of secondary interest. These can be compared to those grown and measured under high CO<sub>2</sub> in order to detect the presence or the direction of acclimation within the photosynthetic apparatus.

Lead treatment decreased net carbon assimilation by 50 and 40% at present and future CO<sub>2</sub> concentration, respectively (Fig. 1). Cadmium had an even more drastic effect with decreases of about 85 and 70%. Replacing half of the Cd by Pb reduced the damage to 75 and 60%. Although lead was taken up to a greater extent than cadmium, its toxicity was milder. This can be explained by its inactivation as extra- and intracellular lead-phosphate precipitation (Koeppel, 1981). Net CO<sub>2</sub> assimilation was enhanced by elevated CO<sub>2</sub> concentration to a greater extent in the heavy metal stressed plants (except for the Pb-treatment).

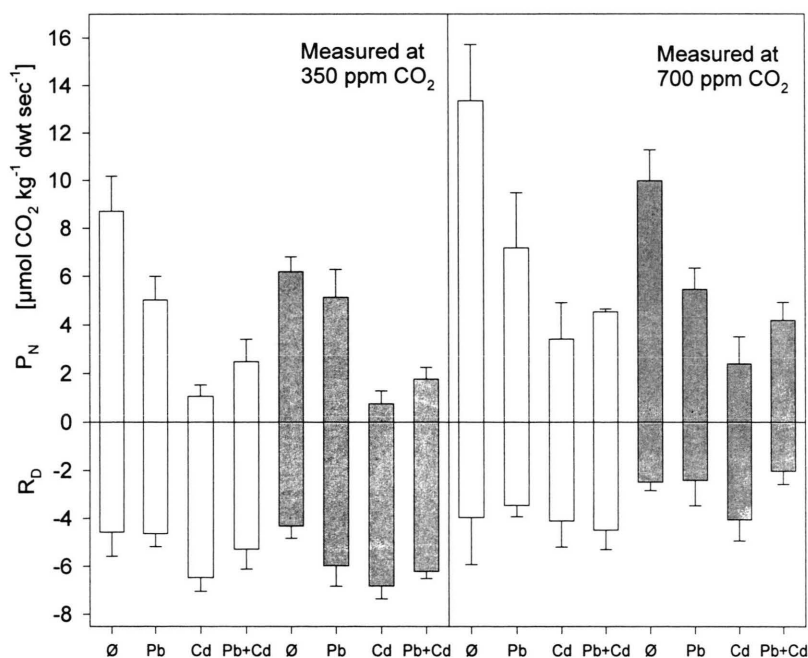


Fig. 1. Net CO<sub>2</sub> assimilation (P<sub>N</sub>) and dark respiration (R<sub>D</sub>) values of *Cladonia convoluta* grown at 350 (open bars) and 700 (hatched bars) in  $\mu\text{mol mol}^{-1}$  CO<sub>2</sub> and different heavy metal treatments. Ø: "control"; Pb: 350  $\mu\text{mol l}^{-1}$  Pb(NO<sub>3</sub>)<sub>2</sub>; Cd: 350  $\mu\text{mol l}^{-1}$  Cd(NO<sub>3</sub>)<sub>2</sub>; Pb+Cd: 175  $\mu\text{mol l}^{-1}$  Pb(NO<sub>3</sub>)<sub>2</sub> + 175  $\mu\text{mol l}^{-1}$  Cd(NO<sub>3</sub>)<sub>2</sub>. Error bars represent standard deviation of three replicates.

When comparing net assimilation values measured at the same CO<sub>2</sub> concentration, those grown at high CO<sub>2</sub> have a lower rate indicating downward acclimation of photosynthesis. However, this acclimation appears only in the control plants. We suggest that heavy metal treatment caused a greater variance in the net assimilation values than CO<sub>2</sub> exposure and hid its effects.

When comparing the lichens measured at their own CO<sub>2</sub> exposure level, we can find a significant decrease in dark respiration in control and all metal treatments due to the high CO<sub>2</sub> (Fig. 1). Heavy metals can increase respiration rate in plants (Poskuta *et al.*, 1996). In this case plants receiving high concentration of Cd indicate this. This phenomenon was attributed to the uncoupling of the mitochondrial membrane (Brown and Wells, 1990). According to Ryan (1991), elevated CO<sub>2</sub> suppresses dark respiration because high intercellular CO<sub>2</sub> might affect indirectly respiratory enzymes and intercellular pH. This view supports our data, which represent rather a prompt effect of high CO<sub>2</sub> on dark respiration.

P<sub>N</sub>/R<sub>D</sub> (net assimilation divided by dark respiration) values representing overall carbon balance

(Meenks *et al.*, 1991) doubled in the control and Pb-treated samples while they increased 3.6-fold at Cd-treated lichen and 4.3-fold at the combined treatment due to the long-term elevated CO<sub>2</sub> concentration.

These results affirm earlier observations (Tuba *et al.*, 1998) that future high CO<sub>2</sub> level might be beneficial for desiccation tolerant cryptogamic plants by improving their carbon balance. This may be even more the case in the presence of a stress factor, namely heavy metal pollution. We would like to point out that elevated CO<sub>2</sub> can ameliorate partly the deleterious effects of heavy metal stress.

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- Arp W. J. and Drake B. G. (1991), Increased photosynthetic capacity of *Scirpus alnei* after four years of exposure to elevated CO<sub>2</sub>. *Plant Cell Environ.* **14**, 1003–1006.
- Arp W. J., Van Mierlo J. E. M., Berendse F. and Snijders W. (1998), Interactions between elevated CO<sub>2</sub> concentration, nitrogen and water: effects on growth and water use of six perennial plant species. *Plant Cell Environ.* **21**, 1–11.
- Atal N., Saradhi P. P. and Mohanty P. (1991), Inhibition of the chloroplast photochemical reactions by treatment of wheat seedlings with low concentrations of cadmium: Analysis of electron transport activities and changes in fluorescence yield. *Plant Cell Physiol.* **32**, 943–951.
- Balaguer L., Valladares F., Ascaso C., Barnes J. D., Delos Rios A., Manrique E. and Smith E. C. (1996), Potential effects of rising tropospheric concentrations of CO<sub>2</sub> and O<sub>3</sub> on green-algal lichens. *New Phytol.* **132**, 641–652.
- Brown D. H. and Sidhu M. (1992), Heavy metal uptake, cellular location, and inhibition of moss growth. *Crypt. Bot.* **3**, 82–85.
- Berg T., Røyset O. and Steinnes E. (1995), Moss (*Hylocomium splendens*) used as biomonitor of atmospheric trace element deposition: Estimation of uptake efficiencies. *Atmospheric Environ.* **29**, 353–360.
- Brown D. H. and Beckett R. P. (1985), Intracellular and extracellular uptake of cadmium by *Rhytidiadelphus squarrosus*. *Ann. Bot.* **55**, 179–188.
- Brown D. H. and Wells J. M. (1990), Physiological effects of heavy metals on the moss *Rhytidiadelphus squarrosus*. *Ann. Bot.* **66**, 641–647.
- Chettri M. K., Sawidis T. and Karataglis S. (1997), Lichens as a tool for biogeochemical prospecting. *Eco-tox. Environ. Safety.* **38**, 322–335.
- Gadd G. M. (1993), Interactions of fungi with toxic metals. *Tansley Review No. 47. New Phytol.* **124**, 25–60.
- Garty J., Karary Y. and Harel J. (1992), Effect of low pH, heavy-metals and anions on chlorophyll degradation in the lichen *Ramalina duriae* (De Not) Bagl. *Environ. Exp. Bot.* **32**, 229–234.
- Garty J., Kloog N., Wolfson R., Cohen Y., Karnieli A. and Avni A. (1997), The influence of air pollution on the concentration of mineral elements, on the spectral reflectance response and on the production of stress-ethylene in the lichen *Ramalina duriae*. *New Phytol.* **137**, 587–597.
- Jarvis P. G. (1993), Global change and plant water relations. In: *Water Transport in Plants under Climatic Stress* (M. Borghetti, J. Grace and A. Raschi, eds.), chap. 1. Cambridge University Press.

- Koeppel D. E. (1981), Lead: Understanding the minimal toxicity of lead in plants. In: *Effect of Heavy Metal Pollution on Plants* (N. W. Lepp, ed.), chap. 2. Applied Science Publishers, London, New Jersey.
- Krupa Z. and Baszynski T. (1995), Some aspects of heavy metal toxicity towards photosynthetic apparatus – direct and indirect effects on light and dark reactions. *Acta Physiol. Plant.* **17**, 177–190.
- Lee E. H., Pausch R. C., Rowland R. A., Mulchi C. L. and Rudorff B. F. T. (1997), Responses of field-grown soybean (cv. Essex) to elevated SO<sub>2</sub> under two atmospheric CO<sub>2</sub> concentrations. *Environ. Exp. Bot.* **37**, 2–3.
- Lichtenthaler H. K. (1988), *In vivo* chlorophyll fluorescence as a tool for stress detection in plants. In: *Application of Chlorophyll Fluorescence* (H. K. Lichtenthaler, ed.), Kluwer Acad. Publ., Dordrecht, pp. 129–142.
- McKee I. F., Farage P. K. and Long S. P. (1995), The interactive effects of elevated CO<sub>2</sub> and O<sub>3</sub> concentration on photosynthesis in spring wheat. *Photosynth. Res.* **45**, 111–119.
- Meenks J. L. D., Tuba Z. and Csintalan Zs. (1991), Ecophysiological responses of *Tortula ruralis* upon transplantation around a power plant in west Hungary. *Journ. Hattori Bot. Lab.* **69**, 21–35.
- Nash T. H. (1975), Influence of effluents from a zinc factory on lichens. *Ecol. Monogr.* **45**, 183–198.
- Nieboer E., Puckett K. J. and Grace B. (1976), The uptake of nickel by *Umbilicaria muhlenbergii*: a physicochemical process. *Can. J. Bot.* **54**, 724–733.
- Polle A., Eiblmeier M., Sheppard L. and Murray M. (1997), Responses of antioxidative enzymes to elevated CO<sub>2</sub> in leaves of beech (*Fagus sylvatica* L.) seedlings grown under a range of nutrient regimes. *Plant Cell Environ.* **20**, 1317–1321.
- Poskuta J. W., Parys E. and Romanowska E. (1996), Toxicity of lead to photosynthesis, accumulation of chlorophyll, respiration and growth of *Chlorella pyrenoidosa* – protective role of dark respiration. *Acta Physiol. Plant.* **18**, 165–171.
- Reuveni J., Gale J. and Zeroni M. (1997), Differentiating day from night effects of high ambient CO<sub>2</sub> on the gas exchange and growth of *Xanthium strumarium* L. exposed to salinity stress. *Ann. Bot.* **79**, 191–196.
- Ryan M. G. (1991), Effects of climate change on plant respiration. *Ecol. Appl.* **1**, 157–167.
- Scarasciamugno G., Deangelis P., Matteucci G. and Valentini R. (1996), Long-term exposure to elevated CO<sub>2</sub> in a natural *Quercus ilex* L. community – net photosynthesis and photochemical efficiency of PSII at different levels of water-stress. *Plant Cell Environ.* **19**, 643–654.
- Schwanz P., Haberle K. H. and Polle A. (1996), Interactive effects of elevated CO<sub>2</sub>, ozone and drought stress on the activities of antioxidative enzymes in needles of norway spruce trees (*Picea abies*, [L.] Karsten) grown with luxurious N-supply. *J. Plant Physiol.* **148**, 351–355.
- Sicher R. C., Kremer D. F. and Rodermeil S. R. (1994), Photosynthetic acclimation to elevated CO<sub>2</sub> occurs in transformed tobacco with decreased ribulose-1,5-bisphosphate carboxylase/oxygenase content. *Plant Physiol.* **104**, 409–415.
- Tuba Z. and Csintalan Zs. (1993), Bioindication of road motor traffic caused heavy metal pollution by lichen transplants. In: *Plants as Biomonitors. Indicators for Heavy Metals in the Terrestrial Environment* (B. Markert, ed.), chap. 12. VCH, Weinheim.
- Tuba Z., Csintalan Zs. and Proctor M. C. F. (1996), Photosynthetic responses of a moss, *Tortula ruralis*, ssp. *ruralis*, and the lichens *Cladonia convoluta* and *C. furcata* to water deficit and short periods of desiccation, and their ecophysiological significance: a baseline study at present-day CO<sub>2</sub> concentration. *New Phytol.* **133**, 353–361.
- Tuba Z., Csintalan Zs., Szente K., Nagy Z. and Grace J. (1998), Carbon gains by desiccation-tolerant plants at elevated CO<sub>2</sub>. *Funct. Ecol.* **12**, 39–44.
- Tuba Z., Szente K., Nagy Z. and Koch J. (1995), Responses of CO<sub>2</sub> assimilation, transpiration and water use efficiency to long-term elevated CO<sub>2</sub> in perennial C<sub>3</sub> xeric loess steppe species. *J. Plant Physiol.* **148**, 356–361.
- Wayne P. M., Reekie E. G. and Bazzaz F. A. (1998), Elevated CO<sub>2</sub> ameliorates birch response to high temperature and frost stress: Implications for modelling climate-induced geographic range shifts. *Oecologia* **114**, 335–342.
- Wells J. M. and Brown D. H. (1995), Cadmium tolerance in a metal-contaminated population of the grassland moss *Rhytidiadelphus squarrosus*. *Ann. Bot.* **75**, 21–29.