

5-Hydroxy-3-vinyl-2(5H)-furanone – a New Inhibitor of Human Synovial Phospholipase A₂ and Platelet Aggregation from Fermentations of a *Calyptella* Species (Basidiomycetes)

Kirsten Lorenzen^a, Timm Anke^a, Silvia Konetschny-Rapp^b and Werner Scheuer^b

^a LB Biotechnologie der Universität, D-67663 Kaiserslautern, Bundesrepublik Deutschland

^b Boehringer Mannheim, D-68305 Mannheim, Bundesrepublik Deutschland

Z. Naturforsch. **50c**, 403–409 (1995); received January 16/February 10, 1995

Basidiomycetes, *Calyptella* Species, Platelet Aggregation, Antibiotics, Phospholipase A₂

5-Hydroxy-3-vinyl-2(5H)-furanone, a potent and selective inhibitor of human synovial phospholipase A₂ was isolated from fermentations of a *Calyptella* species. Its structure as identified by spectroscopic methods is identical to PA 147, an antibiotic previously isolated from a streptomycete. 5-hydroxy-3-vinyl-2(5H)-furanone inhibits the aggregation of human and bovine platelets stimulated by different inducers and exhibits weak antimicrobial activities.

Introduction

It is generally accepted that platelets play a major role in the pathogenesis of several vascular disorders as myocardial infarction and arteriosclerosis (Holmsen, 1982). Their receptors for different agonists as well as the following pathways of signal transduction offer a number of interesting pharmacological targets e.g. phospholipases A₂ (PLA₂) and C (PLC), protein kinase C (PKC) and adenylate cyclase (ADC). PLA₂ of platelets is one central enzyme in the cascade of signals leading to complete aggregation (Kramer *et al.*, 1989).

Arachidonic acid liberated by PLA₂'s of different tissues serves as a precursor of prostaglandines, leukotrienes and thromboxanes, mediators that play a central role in diseases like arthritis or inflammation (Decker, 1991).

In the course of a screening several hundred extracts derived from submerged cultures of basidiomycetes were tested for the presence of inhibitors of collagen-induced aggregation of bovine platelets.

In the following we describe the fermentation, isolation, structure elucidation and new biological properties of 5-hydroxy-3-vinyl-2(5H)-furanone **1** from fermentations of *Calyptella* sp. 9039.

Material and Methods

General

IR and UV spectra were measured with a Bruker ISF 48 and a Perkin-Elmer Lambda 16 UV/VIS spectrometer, respectively. For analytical HPLC a Hewlett-Packard 1090 series II instrument was used.

All NMR spectra were recorded on a Bruker AMX500 spectrometer working at 500.14 MHz for ¹H and at 125.77 MHz for ¹³C. The solution of 5-hydroxy-3-vinyl-2(5H)-furanone (**1**) was 24 mM in D₂O.

High resolution mass spectra were measured with a Finnigan MAT 312 spectrometer (ion energy 70 eV, source temperature 250 °C, acc. volt 3 kV).

For LC coupled thermospray mass spectroscopy (LC-TSPMS) a Finnigan MAT TSQ45 instrument was used (source temperature 140 °C, nebulizer temperature 140 °C, repeller 130 V; MS/MS: collision gas N₂, collision energy 5.1 V; chromatographic conditions: HPLC: LiChropher RP 18, 10 µm, Merck, column 250×4 mm, water–methanol–trifluoroacetic acid (85:15:0.05), flow rate 1.2 ml/min).

Calyptella sp. strain 9039

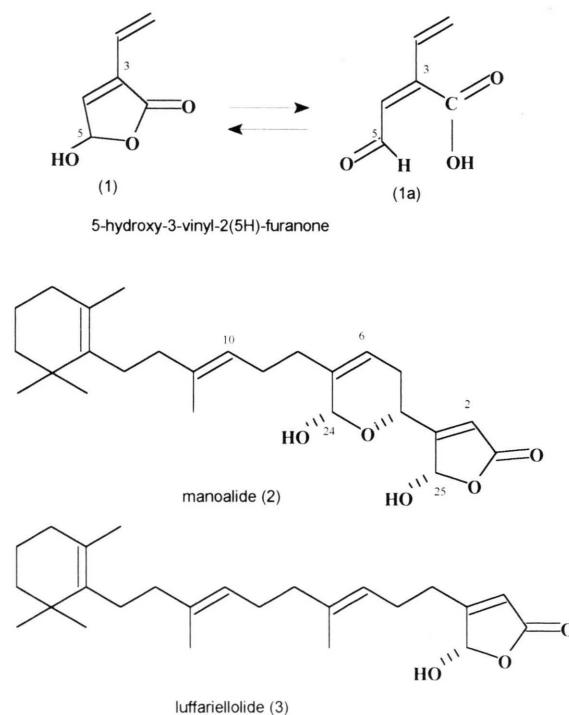
Calyptella sp. strain 9039 was isolated from the spore print of a fruiting body collected on wood in Mossman/Cairns, Australia. The specimen

Reprint requests to Prof. T. Anke.
Telefax: (0631) 2052999.

showed all characteristics of the genus *Calyptella* (Singer, 1986) (Agaricales, Tricholomataceae, Collybieae), the species, however, could not be identified. Herbarium specimen and mycelial cultures are deposited in the culture collection of the LB Biotechnologie, University of Kaiserslautern. For maintenance on agar slants the fungus was grown on YMG medium (yeast extract 0.4 %, malt extract 1.0 %, glucose 0.4 %, pH 5.5).

Fermentation and isolation of 5-hydroxy-3-vinyl-2(5H)-furanone (1)

Fermentations were carried out in a Biostat U fermentor equipped with a MFCS system (B. Braun Biotech) containing 100 l of YMG medium at 135 rpm, 20 liters air/min and 22 °C. The content of oxygen in the medium and CO₂ and oxygen in the exhaust were determined on-line. During fermentation and in fractions during chromatography the content of **1** was measured by analytical HPLC (LiChrosorb RP-18, 5 µm, Merck, column 4×125 mm; 1.5 ml/min, 40 °C; H₂O : MeOH 0 → 100% in 20 min; retention time 4 min) and by platelet aggregation assay.



After 70 hours the oxygen content of the medium was limited by reducing the stirrer speed to 90 rpm and the aeration to 4 litres air/min. After 116 hours of fermentation the mycelia were separated from the culture fluid by filtration. **1** was removed from the culture fluid (95 l) by adsorption to HP21 resin (Mitsubishi) and eluted with acetone. The crude extract (12.86 g) was subjected to gel permeation chromatography (LiChrogel PS1, 7 µm, Merck, column 250×25 mm, elution with 2-propanol) resulting in 2.14 g of enriched product containing approximately 25 % **1**. Pure **1** was obtained by isocratic HPLC (LiChropher RP 18, 10 µm, Bischoff, column 250×20 mm) using water–methanol–trifluoroacetic acid (85:15:0.05); retention time 26 min.

Biological assays

The platelet aggregation assay and the test for inhibitory effects on the synthesis of macromolecules was carried out as reported previously (Lorenzen *et al.*, 1994). Further tests as for cytotoxic effects, hemolytic activity and the antimicrobial activity in the serial dilution assays are described elsewhere (Zapf *et al.*, 1995). The inhibition of phospholipases A₂ from snake venom was tested as described by Nieuwenhuizen *et al.* (1974). The tests for inhibitory effects of **1** on human synovial phospholipase A₂, human pancreatic phospholipase A₂ and human cytosolic phospholipase A₂ were performed according to Scheuer *et al.* (1989) and to Rodewald *et al.* (1994).

Results and Discussion

Fermentation and isolation

Fig. 1 shows a typical fermentation of *Calyptella* sp. 9039 in 100 liters of YMG-medium. After 186 hours the content of 5-hydroxy-3-vinyl-2(5H)-furanone **1**, as detected by analytical HPLC, reached a maximum of 6.5 mg/liter and **1** was isolated as described in the experimental section.

5-Hydroxy-3-vinyl-2(5H)-furanone (1)

The highly unstable 5-hydroxy-3-vinyl-2(5H)-furanone **1** was obtained as slightly yellowish oil soluble in methanol, acetone or ethyl acetate, and moderately soluble in water. At room temperature **1** undergoes rapid polymerisation forming a com-

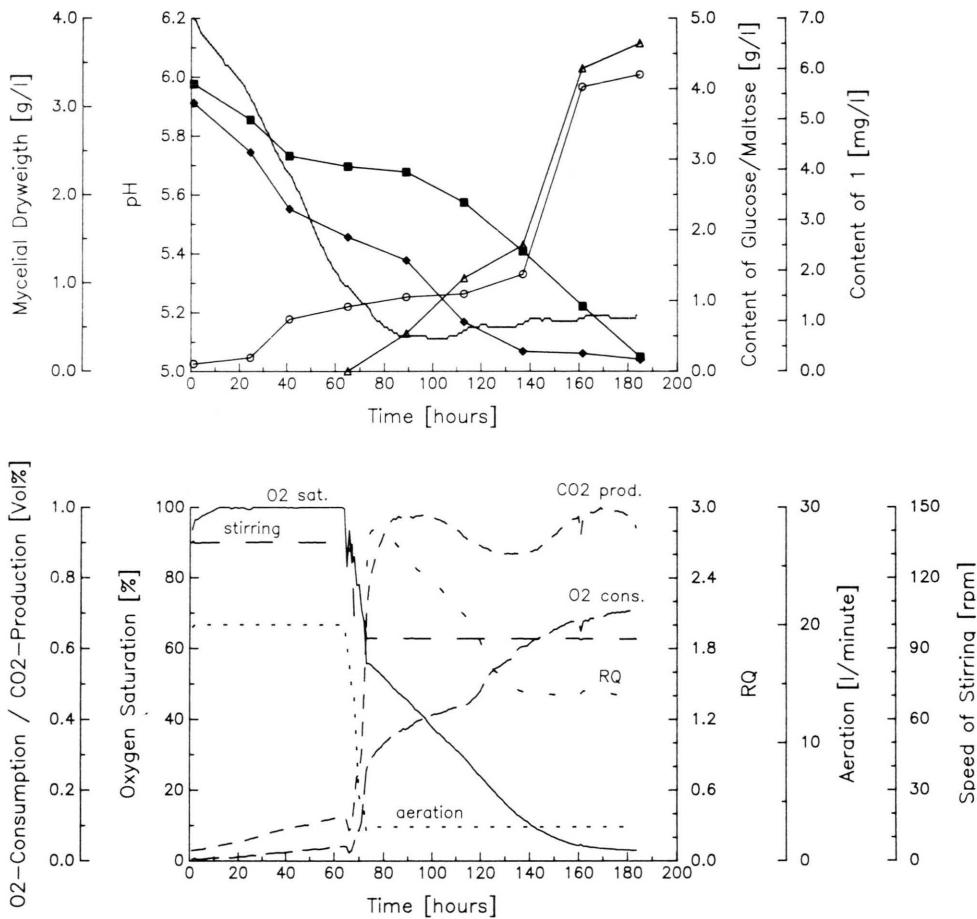


Fig. 1. Fermentation of *Calyptella* sp. TA 9039 in 100 l of YMG-medium. ◆, Content of glucose; ■, content of maltose; —, pH; ○, mycelial dry weight; △, content of **1**.

pletely insoluble polymer. Depending on the pH the 5-hydroxy-3-vinyl-2(5H)-furanone ring of **1** opens to *1a*, the predominant species in neutral methanol. The physico-chemical data of **1** (Table I) prove that the compound isolated from *Calyptella* sp. 9039 is identical to 5-hydroxy-3-vinyl-2(5H)-furanone, first isolated by Els *et al.* (1958) from a *Streptomyces* strain and published as antibiotic PA 147 (Els *et al.*, 1958). **1** is the first metabolite isolated from a *Calyptella* species.

From other authors a synthesis of **1** (Black *et al.*, 1973) starting from prop-2-ynyl vinyl ether has been described. It is interesting that the hydroxy furanone moiety of *1* also occurs in the structure of the well known PLA₂-inhibitors manoalide (**2**) and luffariellolide (**3**) which have

been isolated from several marine sponges (Silva *et al.*, 1980; Potts and Faulkner, 1992).

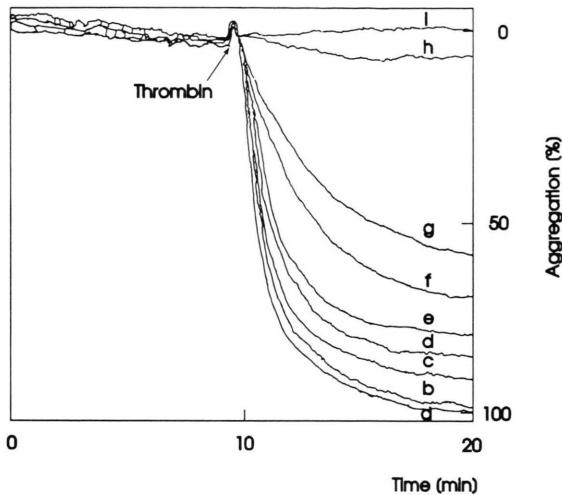
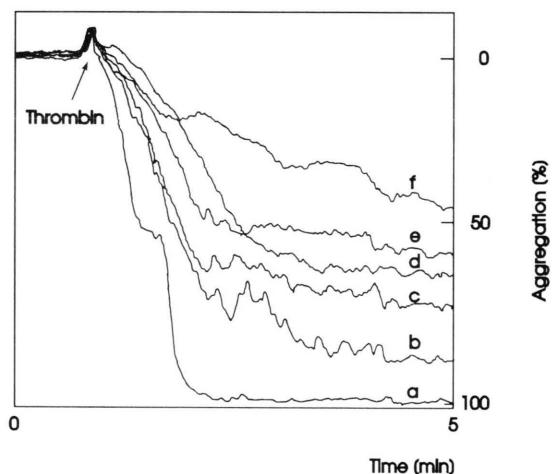
Biological activities

The inhibitory effect of **1** on the thrombin-induced aggregation of bovine platelets is shown in Fig. 2. A complete inhibition of aggregation was obtained by 50 µg/ml (396 µM) of 5-hydroxy-3-vinyl-2(5H)-furanone, the IC₅₀ value (30 % inhibition) of **1** was determined to 9 µg/ml (71 µM). The first, reversible phase, was hardly affected, the second irreversible phase of aggregation was inhibited by concentrations above 10 µg/ml (79 µM).

Figure 3 shows the influence of **1** on the thrombin-induced aggregation of human platelets. The

Table I. Physicochemical properties of 5-hydroxy-3-vinyl-2(5H)-furanone (**1**).

Appearance	yellowish oil
Molecular formula	C ₆ H ₆ O ₃
IR ν_{max} (KBr) cm ⁻¹	3400 (OH), 1750 (C=O), 1640 (C=C)
UV $\lambda_{\text{max}}^{0.1 \text{ n HCl}}$ nm (ϵ)	240 (10,900)
UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (ϵ)	272 (17,800) \Rightarrow 1a
LC-TSPMS (<i>m/e</i>)	127 (17%) MH ⁺ , 159 (100%) [MH + MeOH] ⁺ , 173 (15%) [MH + 2 MeOH - H ₂ O] ⁺ , 191 (15%) [MH + 2 MeOH] ⁺ 191 \rightarrow 159, 127; 159 \rightarrow 127, 109 [MH - H ₂ O] ⁺ ; 127 \rightarrow 109 \Rightarrow 126 Da
MS/MS (<i>m/e</i>) ^a	468 (rel. int. 1%) M ⁺ (tri-TMS derivative of the dimer); 378.1305 (rel. int. 8%) \rightarrow C ₁₈ H ₂₆ O ₅ Si ₂ (D 3.7 ppm) [M - HOTMS] ⁺ molecular formula of dimer: C ₁₂ H ₁₂ O ₆ (252 Da)
¹ H NMR (500 MHz, D ₂ O)	δ 7.28 (s, H-4), 6.48 (dd, H-6, <i>J</i> = 11.4 Hz, 17.8 Hz), 6.24 (s, H-5), 6.16 (d, H-7 (Z), <i>J</i> = 17.8 Hz), 5.59 (d, H-7 (E), <i>J</i> = 11.4 Hz)
¹³ C NMR (125 MHz, D ₂ O)	δ 174.3 (C-2), 147.1, 133.79, 126.61, 124.63, 99.11 (C-5)
TLC <i>R_f</i> value (silica gel, Merck, toluene - acetone (70:30))	0.43
Cyclohexane - ethyl acetate (50:50)	0.30

^a Parent ion \rightarrow daughter ions.^b After TMS derivatization.Fig. 2. Effect of **1** to the thrombin-induced aggregation of bovine platelets (thrombin 0.1 U/ml). (a) Control, (b) 7.9 μ M, (c) 39 μ M, (d) 48.5 μ M, (e) 71 μ M, (f) 79 μ M, (g) 158 μ M, (h) 276 μ M, (i) 396 μ M.Fig. 3. Effect of **1** to the thrombin-induced aggregation of human platelets (thrombin 0.1 U/ml). (a) Control, (b) 39 μ M, (c) 79 μ M, (d) 158 μ M, (e) 396 μ M, (f) 793 μ M.

IC_{30} was determined to 10 μ g/ml (79 μ M), a complete inhibition could not be observed at concentrations up to 100 μ g/ml (793 μ M).

The effects of **1** on the aggregation of human and bovine platelets stimulated with different inducers are compared in Table II. **1** inhibited preferentially the aggregation induced by thrombin and arachidonic acid (AA). In the latter case AA can act both as stimulator of PLA₂ and substrate

Table II. IC_{30} values of **1** for the inhibition of aggregation of human and bovine platelets stimulated by different inducers.

Inducer		IC_{30} value ^a [μ M]	
	Bovine platelets	Human platelets	
Collagen (0.3 mg/ml)	142	198	
ADP (2.5 μ M)	396	198	
Thrombin (0.1 U/ml)	71	79	
Ristocetin (0.4 mg/ml)	—	198	
Arachidonic acid (0.6 μ g/ml)	—	59.5	
U 46619 (0.45 μ M) ^b	—	158	

^a 30% inhibition of aggregation.^b Thromboxane A₂ analogue (UpJohn).

— Inducer not suitable.

for cyclooxygenase. The IC₅₀ values of **1** for the other inducers are significantly higher. A specific interference of **1** with one of the receptors for the agonists seems unlikely.

The receptor for thrombin couples directly to PLA₂ (Lapetina *et al.*, 1990). Therefore an inhibition of membrane bound PLA₂ of platelets seemed possible. If this were true, **1** would interfere with the stimulatory activity of AA under the chosen conditions.

With this in view, the inhibitory activity of **1** was tested against PLA₂'s from *Naja mosambica* (type I), *Crotalus atrox* (type II), U937 cells (type II), human pancreas (type I) and human synovial fluid (type II). The IC₅₀ values presented in Table III show a preferential inhibition of human synovial PLA₂ at nanomolar concentrations, a value which is in good accordance with the inhibition of PLA₂'s (bee venom, cobra venoms, human synovial fluid) observed with manoalide (**2**) and luffariellolide **3** (Potts and Faulkner, 1992).

1, **2**, and **3** differ from each other by the length and position of the hydroxy furanone side chain. **2** and **3** are potent inhibitors of several phospholipases A₂ by forming a Schiff base (imine) between the aldehyde group of the open hydroxybutenolide ring and lysine residues not located in the active centre of the enzymes (Potts *et al.*, 1992).

Because of the described blockage of Ca²⁺ channels by **2** and **3**, the effect of **1** on the secretion of Ca²⁺ by stimulated platelets was tested with the fluorescence indicator Quin-2-acetoxymethyl ester (Tsien *et al.*, 1982; Hallam *et al.*, 1984). For **1** no decrease of intracellular Ca²⁺ after stimulation of platelets was observed.

Other biological activities

Els *et al.* had reported antibacterial (*Staphylococcus aureus*, *Xanthomonas oryzae*) and cytotoxic (HeLa) effects for PA 147 **1** (Els *et al.*, 1958). In mice bearing Ehrlich carcinoma a pronounced antitumor activity was observed.

The cytotoxic activities of **1** against BHK-, HeLa S3-, L 1210-, HL 60- and U 937-cell cultures were tested as described previously (Zapf *et al.*, 1995). A lytic action of **1** on BHK- and HL 60-cells could be observed at concentrations higher than 20 µg/ml (158 µM), while HeLa S3 cells were affected by concentrations starting from 50 µg/ml (396 µM) (Table IV). With L 1210- and U 937 cells a weak inhibition of cell growth at concentrations of 100 µg/ml (793 µM) was observed.

The incorporation of ¹⁴C-labeled thymidine, uridine and leucine into DNA, RNA and proteins of HL 60 cells was tested as described previously (Lorenzen *et al.*, 1994). Figure 4 shows a preferen-

Table IV. Cytotoxic activities of **1** on different cells.

Cells		IC ₁₀₀ ^a [µM]
BHK 21	ATCC CCL 10	158
HeLa S3	ATCC CCL 22	396
L 1210	ATCC CCL 219	>793 ^b
HL 60	ATCC CCL 240	158
U 937	ATCC CRL 1593	>793 ^b

^a Complete lysis of cells.

^b Inhibition of growth at this concentration detectable.

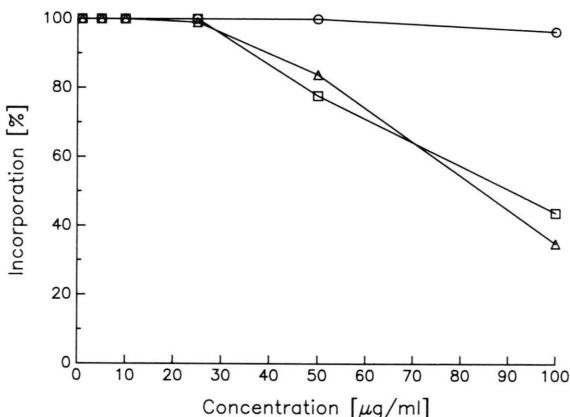


Fig. 4. Effect of **1** to the incorporation of ¹⁴C-labeled precursors in macromolecules of HL 60 cells. ○, ¹⁴C-labeled uridine, control 6354 cpm; □, ¹⁴C-labeled thymidine, control 935 cpm; △, ¹⁴C-labeled leucine, control 3596 cpm.

Table III. Inhibition of phospholipases A₂ from different sources by **1**.

PL A ₂	IC ₅₀ value [µM]
Human synovial PL A ₂ , type II	0.3
Human pancreatic PLA ₂ , type I	32
Human cytosolic PL A ₂ from U 937 cells, type II	<793
PL A ₂ from <i>Naja mosambica</i> , type I	24
PL A ₂ from <i>Crotalus atrox</i> , type II	39

tial inhibition of protein and DNA syntheses by 5-hydroxy-3-vinyl-2(5H)-furanone at concentrations starting from 50 µg/ml (396 µM). The incorporation of ¹⁴C-labeled uridine into RNA was not effected. The complete lack of hemolytic action of **1** on bovine erythrocytes at concentrations up to 100 µg/ml (793 µM) renders a direct action on the cytoplasmic membrane unlikely.

The antifungal and antibacterial effects of 5-hydroxy-3-vinyl-2(5H)-furanone were determined in the serial dilution assay (Table V). The organisms most sensitive to **1** were the yeast *N. coryli* and the bacteria *M. luteus* and *S. thyphimurium*, whereas other bacteria and fungi were less sensitive.

In the "pour-plate-test" for mutagenicity (without microsomes) with four strains of *S. thyphimurium* (TA 97, TA 98, TA 100 and TA 102) (Ames *et al.*, 1975) no increase of the number of revertants

could be observed at concentrations from 1–20 µg/ml (8–158 µM) of 5-hydroxy-3-vinyl-2(5H)-furanone.

In conclusion **1** appears to be a highly potent and selective inhibitor of human synovial phospholipase A₂. The selectivity towards this enzyme is much higher as compared to manoalide (**2**) and luffariellolide (**3**).

Acknowledgements

Part of the work described here was supported by a grant from the Bundesministerium für Forschung und Technologie. We thank Drs Stegmeier and Dörge (Boehringer, Mannheim) and their staff for providing facilities to perform the aggregation assays with human blood and Drs Anders and Hindermayr (Boehringer, Mannheim) for NMR- and mass-spectroscopy.

Table V. Minimal inhibitory concentration (MIC) of **1** in the serial dilution assay.

Organism		MIC [µM]
Bacteria		
<i>Acinetobacter calcoaceticus</i>	DSM 30005	>793
<i>Escherichia coli</i> K12	(BT KL)	>793
<i>Salmonella thyphimurium</i> TA 98	(Dr.N.B. Ames)	79 z
<i>Athrobacter citreus</i>	ATCC 11624	>793
<i>Bacillus brevis</i>	ATCC 9999	793 s
<i>Bacillus subtilis</i>	ATCC 6633	>793
<i>Corynebacterium insidiosum</i>	ATCC 10253	>793
<i>Micrococcus luteus</i>	ATCC 381	7.9 z
<i>Mycobacterium phlei</i> (Lehmann & Neumann, KL)		793 s
<i>Streptomyces</i> spec.	ATCC 23836	>793
Fungi		
<i>Nadsonia fulvescens</i>	ATCC 10645	>793
<i>Nematospora coryli</i>	ATCC 10647	79 z
<i>Saccharomyces cerevisiae</i> is 1		>793
<i>S. cerevisiae</i> S 288 c (Prof. Lacroute, Straßburg)		>793
<i>Fusarium oxysporum</i>	CBS 149.25	>793
<i>Paecilomyces variotii</i>	ETH 114646	>793
<i>Penicillium notatum</i>	(BT KL)	793 z
<i>Mucor miehei</i>	TÜ 284	>793
<i>Rhodotorula glutinis</i>	ATCC 26086	>793
<i>Ustilago nuda</i>	CBS 118.19	>

s. Bacterio-/fungistatic effects.

z. Bacterio-/fungizide effects.

Ames B. N., McCann J. and Yamasaki E. (1975), Methods for detecting carcinogens and mutagens with the salmonella/mammalian-microsome mutagenicity test. *Mut. Res.* **31**, 347–364.

Black D. K., Fomum Z. T., Landor P. D. and Landor S. R. (1973), Allenes. Part XXVIII. Synthesis of antibiotic lactols and allenic ester by Claisen rearrangement of prop-2-ynyl vinyl ester. *J. Chem. Soc., Perkin Trans. I*, 1349–1352.

Decker K. (1991), Basic mechanism of the inflammatory response. In: *Molecular Aspects of Inflammation* (H. Sies, L. Flohé and G. Zimmer, eds). Springer Verlag, Heidelberg, pp. 1–32.

Els H., Sabin B. A. and Celmer W. D. (1958), PA-147 (3-carboxy-2,4-pentadienyl lactol) – a new antibiotic. *J. Am. Chem. Soc.* **80**, 878–880.

Hallam T. J., Thomson N. T., Scrutton M. G. and Rink T. J. (1984), The role of cytoplasmic free calcium in the response of quin-2-loaded platelets to vasopressin. *Biochem. J.* **221**, 897–901.

Holmsen H. (1982), Platelet secretion. In: *Hemostasis and Thrombosis* (R. W. Colman, ed.). J. B. Lippincott Company, Philadelphia, pp. 390–403.

Kramer R. D., Hession C., Johansen B., Hayes G., McGraw P., Chow E. P., Tizard R. and Pepinsky R. B. (1989), Structure and properties of a human non-pancreatic phospholipases A₂. *J. Biol.* **264**, 5768–5775.

Lapetina E. G. (1990), The signal transduction induced by thrombin in human platelets. *FEBS Lett.* **268**, 400–404.

Lorenzen K., Anke T., Anders U. and Hansske F. (1994), Two inhibitors of platelet aggregation from a *Panus* species (Basidiomycetes). *Z. Naturforsch.* **49c**, 132–138.

Nieuwenhuizen W., Kunze H. and de Haas G. H. (1974), Phospholipases A₂ from porcine pancreas. In: *Methods in Enzymology*, Vol. **32** (S. Fleischer and L. Packer, eds.). Academic Press, New York, pp. 147–154.

Potts B. C. M. and Faulkner D. J. (1992), Phospholipase A₂ inhibitors from marine organism. *J. Nat. Prod.* **55**, 1701–1717.

Potts B. C. M., Faulkner D. J., deCarvalho M. S. and Jacobs R. S. (1992), Chemical mechanism of inactivation of bee venom phospholipase A₂ by the marine natural products manoolide, luffariellolide, and scalardial. *J. Am. Chem. Soc.* **114**, 5093–5100.

Rodewald E., Tibes U., Maass G. and Scheuer W. (1994), Induction of cytosolic phospholipase A₂ in human leucocytes by lipopolysaccharide. *Eur. J. Biochem. FEBS* **223**, 743–749.

Scheuer W. (1989), Phospholipase A₂-regulation and inhibition. *Klin. Wochenschr.* **67**, 153–159.

Silva E. D. and Scheuer P. J. (1980), Manoolide, an antibiotic sesterterpenoid from the marine sponge *Luffariella variabilis* (Polejaeff). *Tetrahedron Lett.* **21**, 1611–1614.

Singer R. (1986), The Agaricales in Modern Taxonomy. Koeltz Scientific Books, Koenigstein, pp. 179–182.

Tsien R. Y., Pozzan T. and Rink T. J. (1982), Calcium homoeostasis in intact lymphocytes: cytoplasmic free calcium monitored with a new, intracellularly trapped fluorescent indicator. *J. Cell Biol.* **94**, 325–334.

Zapf S., Hoßfeld M., Anke H., Velten R. and Steglich W. (1995), Darlucin A and B, new isocyanide antibiotics from *Sphaerellopsis filum* (*Darluca filum*). *J. Antibiot.* **48**, 36–41.