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Isosativan: an Isoflavan Phytoalexin from Trifolium hybridum and other Trifolium Species

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(Z. Naturforsch. 31 c, 331-332 [1976]; received January 14, 1976)

Leguminosae, Trifolium, Isoflavan, Pterocarpan, Phytoalexin

An isoflavonoid phytoalexin isolated from the fungus-infected leaves of *Trifolium hybridum* has been identified as 7,4'-dimethoxy-2'-hydroxyisoflavan.

The disease resistance of many higher plants may depend on the post-infectional accumulation of antifungal compounds called phytoalexins ¹. In general, species of the Leguminosae (subfamily Lotoideae) produce isoflavonoid phytoalexins (pterocarpans and isoflavans) although one exception (the furanoacetylene, wyerone acid from *Vicia faba*) has been reported ². During a phytochemical survey of the genus *Trifolium*, it was found that in addition to known compounds, the fungus-infected leaves of alsike clover (*T. hybridum* L.) produced an isoflavonoid not previously described as a phytoalexin. From the evidence presented below, this compound has been formulated as vestitol-7-O-methyl ether (1).

Phytoalexins were isolated from the detached leaves of T. hybridum using the drop-diffusate technique as previously described 3 . Conidial suspensions of the non-pathogenic fungus, Helminthosporium carbonum Ullstrup served as the phytoalexin inducer. Diffusates from infected leaves were extracted with EtOAc and the organic fractions bulked and reduced to dryness. TLC (CHCl $_3$: MeOH, 100:2, Merck, Si-gel F_{254} , $0.25 \, \text{mm}$) of the residue afforded three major phenolic zones at R_F 0.14 (Band 1), 0.47 (Band 2) and 0.55 (Band 3); a minor zone (Band 4) was also apparent at approx. R_F 0.66. The Band 1 component was purified in n-pentane: Et $_2$ O: HOAc (PEA) (75:25:3, 3 X) to afford the known isoflavan phytoalexin, vestitol 4

1: R¹=R³=CH₃; R²=H 2: R¹=R²=H; R³=CH₃ 3: R¹=H; R²=R³=CH₃ 4: R¹=R²=R³=CH₃

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(2). Purification of Band 2 (CHCl₃, 3 X) gave sativan³ (3) together with a lower phenolic fraction which separated in PEA (75:25:3, 3 X) to give the pterocarpans medicarpin (5) (upper zone) and maackiain (6)⁵ (lower zone). Identification

5: R¹=OH; R²=R³=H; R⁴=OCH₃
6: R¹=OH; R²=H; R³=R⁴=O-CH₂-O
7: R¹=OH; R²=OCH₃; R³=R⁴=O-CH₂-O
8: R¹=R⁴=OCH₃; R²=R³=H

of compounds 2-6 was based on a UV and TLC comparison with authentic material. TLC of Band 4 (PEA, 75:25:1) gave small quantities of a compound (R_F 0.37) identified (MS and UV) as 4-methoxymaackiain (7). Control diffusates 3 contained only traces of 2,5 and 6.

Further purification of Band 3 (PEA, 75:25:1, R_F 0.63; CHCl₃, R_F 0.37) afforded a phenolic compound 1 which reacted to both diazotised p-nitroaniline 6 (yellow) and Gibbs reagent 7 (deep blue). UV maxima [nm] recorded for this compound were, 1. EtOH: 214, 227 sh, 281, 284 and 289 sh and 2. EtOH + NaOH: 219, 245 sh, 286 sh, 291 and 300 sh. The MS was typical of a simple isoflavan 8 and gave a molecular ion at m/e 286 (corresponding to $C_{17}H_{18}O_4$) and prominent fragments at m/e151, 150 (base), 149, 148, 138 and 137. The ions at m/e 150 and 137 can be obtained by fragmentation of an isoflavan with monomethoxymonohydroxy substitution of the B-ring; the ion at m/e 149 can be formulated as a monomethoxylated fragment derived from ring-A. Since naturally occurring isoflavans are oxygenated at C-7, 2' and 4', the above compound was formulated as 7,4'-dimethoxy-2'-hydroxyisoflavan (1) (vestitol-7-0-methyl ether). The phenolic hydroxyl group was located at C-2' (rather than C-4') from the positive (blue) Gibbs reaction 7. Although 1 has been extracted from the wood of Dalbergia ecastophyllum 9, it has not previously been associated with herbaceous plant tissues. Nor has I been assigned a common name; in view of its isomeric relationship to the isoflavan phytoalexin, sativan 3 (3), the trivial name Isosativan would seem appropriate.

Structure 1 for isosativan was confirmed by acetylation, methylation and synthesis, since data (UV, MS) reported for the *Dalbergia* metabolite ⁹ differ slightly from those noted above. Acetylation and TLC purification (CHCl₃, R_F 0.49) gave a monoacetate ($\lambda_{\max}^{\rm EtOH}$ nm: 215, 226, 280 sh, 283 and 289) with M⁺ 328 and fragments at m/e 286

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(M·42), 151, 150 (base), 149, 148 and 137. The ion at M·42 is indicative of an aromatic hydroxyl group ¹⁰. When methylated with diazomethane, isosativan afforded a monomethyl ether (CHCl₃, R_F 0.90) ($\lambda_{\rm max}^{\rm EtOH}$ nm: 211, 226, 280, 284 and 289 sh; M⁺ 300, m/e 165, 164 (base), 152, 151, 149 and 121) indistinguishable (UV, MS and TLC) from a sample of the trimethoxyisoflavan 4. Formation of this compound establishes unequivocally the 7,2′,4′-oxygenation pattern of isosativan.

The structure of isosativan was finally verified by synthesis from homopterocarpin (8). Crystalline 8 (5 mg), glacial HOAc (3 ml) and 10% Pd-C (5 mg) were hydrogenated at 80 °C for 25 min. After removal of catalyst and solvent, the residue was chromatographed in CHCl₃: MeOH (100:2) to afford dihydrohomopterocarpin (1). This compound was found to be identical (UV, MS and TLC) with isosativan.

When bioassayed against the mycelial growth of H. carbonum, the antifungal activity of isosativan (ED₅₀ 16 μ g/ml) was found to be comparable with that of the related isoflavans, vestitol (ED₅₀ 17 μ g/ml) and sativan (ED₅₀ 10 μ g/ml). Although leaf diffusates from T. $h\gamma bridum$ contain isosativan (10

 μ g/ml) in relatively small quantities (cf. 2, 170 μ g/ml; 3, 8 μ g/ml; 5, 70 μ g/ml; 6, 56 μ g/ml; 7, 2 μ g/ml) there seems little doubt that this compound functions as a resistance factor.

As well as T. hybridum, isosativan is produced by T. subterraneum L., T. spumosum L., T. scabrum L., T. stellatum L. and T. tomentosum. These species also accumulate vestitol from which isosativan can be derived. For T. hybridum, the terminal stages of phytoalexin biosynthesis presumably involve conversion of medicarpin (5) to vestitol (2) and methylation (at C-7 or 2') of the latter compound to give either isosativan (1) or sativan chemically 'advanced' pterocarpan maackiain (6) is apparently produced by a route which does not require the participation of medicarpin 11. From a phytochemical comparison with over 50 other Trifolium species (J. L. Ingham, unpublished data), it appears that in terms of isoflavonoid production, T. hybridum is one of the more chemically evolved members of the genus. The results of the abovementioned survey will be published elsewhere.

The author thanks R. W. Butters (Tate and Lyle Memorial Laboratory, Whiteknights, Reading) for MS analyses.

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