

Review Article

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Nanozymes – A route to overcome microbial resistance: A viewpoint

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Abstract: The bactericide is one of the major objective consequences related to healthcare in the world. Natural enzymes have been broadly utilized in various applications such as biomedical areas due to their broad catalytic activities and substrate particularity. While anticipating, it has drawbacks like higher cost, low stability, and troubles in reprocessing. Additionally, artificial enzymes (nanozymes) have favors above natural enzymes, for example, the effortless yield on a big scale, low costs, and high stability in coarse surrounds. The amount of antibiotic repellent microorganisms has activated big concern in the growth of stuff with essential bactericide potentials such as metal or metal oxide nanoparticles, cationic polymeric compounds, graphene oxide, and other carbon materials that can be used as antibacterial agents by altering cell morphology. In this report, we have summarized catalytic antibacterial strategies by natural enzymes, artificial enzymes, or photocatalytic activity. Furthermore, the demands and hereafter contents about catalytic antibacterial strategies are supposed in this report.

Keywords: antibacterial activity, strategies, natural enzyme, nanozymes

1 Introduction

The developing health troubles due to bacteria is one of the major concerns causing infective diseases conducting to millions of illnesses and demises yearly [1–3]. Haunting diseases invariably relate to establishing bacterial colonies [4–6]. The bacteria in the exo-polymeric matrix were highly tolerant to formal antibiotics due to developed resistors, controlled dissemination, and deactivation of antibiotics [7–9]. To overcome this limitation, significant attempts have been devoted to colloidal materials to defend the world *versus* diseases caused by bacteria [10–16]. Due to the lack of bioavailability, they had to be more attentive in their virtual efforts. ROS stands for reactive oxygen species, including chemicals like hydrogen peroxide and superoxide, effective against antibiotic-resistant bacteria [17–20]. ROS and hydrogen peroxide has affected bacterial structure by disturbing biomolecules that lead to the death of a cell [21–23]. Unfortunately, hydrogen peroxide has limitations in applications such as lower efficiency, slow procedure, and eminent concentrations [24]. Particularly, high concentrations of hydrogen peroxide induce immunogenicity and inflammation and have perniciousness extravagantly to healthy tissues and yet delay wound healing [25,26]. Practicality surface modification of nano-stuffs supplies a flexible stage to pattern new disinfectants to fight multidrug-resistant (MDR) pathogens [27]. Similarly, nanomaterials have limitations over antibacterial activity, such as high doses of AgNPs can affect the mammalian cells. Techniques such as core-shell (Au@AgNPs) material production or surface coating of particles on or in electrospun fibers have been introduced to minimize the toxicity of nanomaterials. However, many researchers are still working to develop a superior antimicrobial infection treatment. Recently, catalytic antimicrobial materials are getting more attention; such materials have enzyme-like activities and are safer for cells. Peroxidase and oxidase in lysosomes can catalyze ROS production to fight bacterial encroachment because the living mechanism has a self-defensive arrangement that can work as a biocatalyst to inhibit bacterium or

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interrupt biofilm state. Nevertheless, natural enzymes frequently endure underlying defects. Therefore, materials having photocatalytic activity are prominent antibacterial materials due to oxygen reduction and bringing forth ROS as though O_2^- and hydrogen peroxide [28–30]. Enzyme-mimicking catalytic performances of nanomaterials have been reported by Chen *et al.*, in a recent report [31]. Despite lots of fantabulous exploits that have been described, to our know-how, no comprehensive report has been given on catalytic antibacterial strategies. Despite lots of fantabulous exploits that have been described, to our know-how, no comprehensive report has been given on catalytic antibacterial strategies. In this report, the recent developments in the area of catalytic antibacterial strategies have been presented. Briefly, catalytic antibacterial strategies including natural enzymes, nanozymes, and photocatalytic antibacterial strategies have been discussed.

2 Catalytic antibacterial strategies

Serial publications of examinations have been brought out to figure out the antibacterial mechanisms. The most recent reports on performance of nanozymes have been described by Dong *et al.*, Huang *et al.*, and Sun *et al.* [32–34]. Hence, antibacterial strategies have been well described, but it still needs a flick to make them simpler for the research world.

2.1 Enzymes effective antibacterials and antibiofilms

Enzymes are effective biocatalysts and are primarily made up of proteins, although some are catalytic RNA

molecules [35,36]. Where distinctive catalysts are frequently employed in coarse considerations like level of temperature and pressure [37,38], they are principally applied to catalyze the salvation of biological specks, and these reactions are generally brought out beneath comparatively modest circumstances [39,40]. Enzymes have been broadly employed in the area of industry, medicine, and biology [41–44]. The classification of bactericide enzymes is shown in Figure 1. However, it has been depicted that natural enzymes have intrinsic shortcomings or disadvantages. Extremely developed enzymes – lysins have a bacterial effect (Figure 2). Phage lytic enzymes are extremely effective specks that have been purified over decades of evolution. This lysin directs the incorruptibility of the cell wall and is intended to affect one of the five important bonds in the peptidoglycan. However, lysins only exploit gram-positive bacterium because they can interact with the cell wall and are intended to affect one of the five important bonds in the peptidoglycan. However, lysins only exploit gram-positive bacteria because they can interact with the cell wall carbohydrates and peptidoglycan since the gram-negative bacterium defends this contact due to the outer membrane. Furthermore, T4 lysozyme is acknowledged as a bactericide protein having antibacterial activity *versus* both types of bacteria. The bacterial property of T4 lysozyme is supposed to occupy in its muramidase action that extends to change in the state of the murein layer and reduction of the mechanical intensity of the microbial cell wall and finally leading to death by lysis. It has been detailed on construction and role states the value of amino acid components for the muramidase action of T4 lysozyme. The performance of T4 lysozyme for bacterial membrane degradation or lysis is as detected for respective antimicrobial peptides [45].

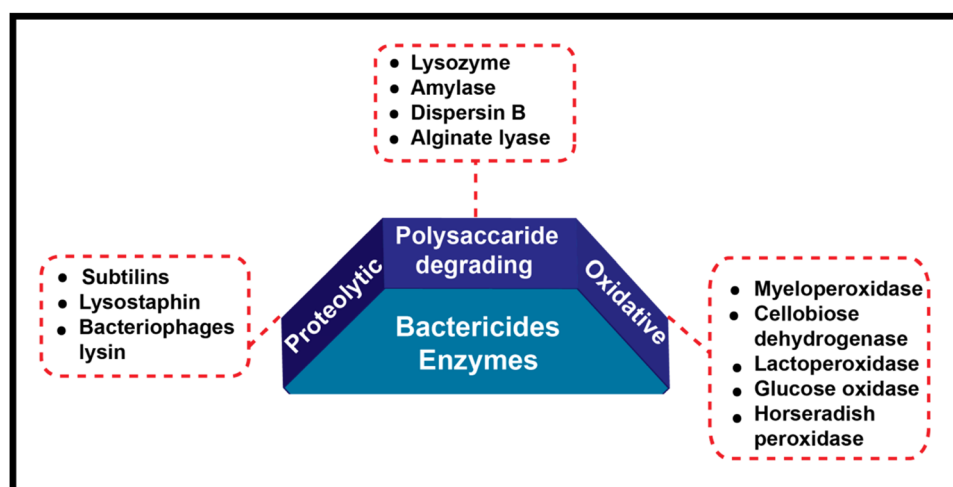


Figure 1: Antimicrobial enzymes.



Figure 2: Lysine-affected bacillus displaying attribution of the cytoplasmic membrane [49].

Aforementioned, one enzyme catalytically should be enough to properly break bonds to inhibit bacteria. Regarding the efficacy of lysin, it only kills the organisms that produced it. Particularly, it is *C. streptococcal* phage that destroys C, A, and E *streptococci* set, also *Streptococcus uberis* and the horse-infective bacteria *S. equi* can be resisted by a bovine infectious agent. Moreover, *streptococci* did not found effective in the oral cavity. Recently, lysin CF-301 phage has been introduced for the disruption of *Staphylococcus aureus* biofilm [46]. Figure 3 shows the morphology of control and

lysin CF-301 phage biofilm using scanning electron microscopic images. Before and after treatment of lysin CF-301 phage has different morphologies, which means it has an effect on lysin. Bacteriophage as a bicomponent molecule, such as holin and endolysin, allow novel produced phage offspring to depart dead bacterium and occupy early sensitive host bacterium. In the inner membrane of the bacterium kiln, a molecule of permeable enzyme can create pores, thereby inhibiting bacterial growth. Another component of phage disturbs the exo-membrane of cells that demolish the peptidoglycan layer [47]. The clinical testing CF-301 against methicillin-resistant *Staphylococcus aureus* (MRSA) is currently complete. Human trials are currently underway to cure patients in hospitals with MRSA germs or inflammation of the endocardium and heart valves [48].

2.2 Nanozymes – effective antibacterials

The name “artificial enzyme” was created verbally by scientists to explain mimic enzyme models [50]. It is a material with sizes of 1–100 nm [51,52]. Artificial enzymes are productively employed as chemical catalysts. Newly published literature is significantly lined up on consideration of nano-size materials that possess the biocatalytic ability. Artificial enzymes have benefits over natural

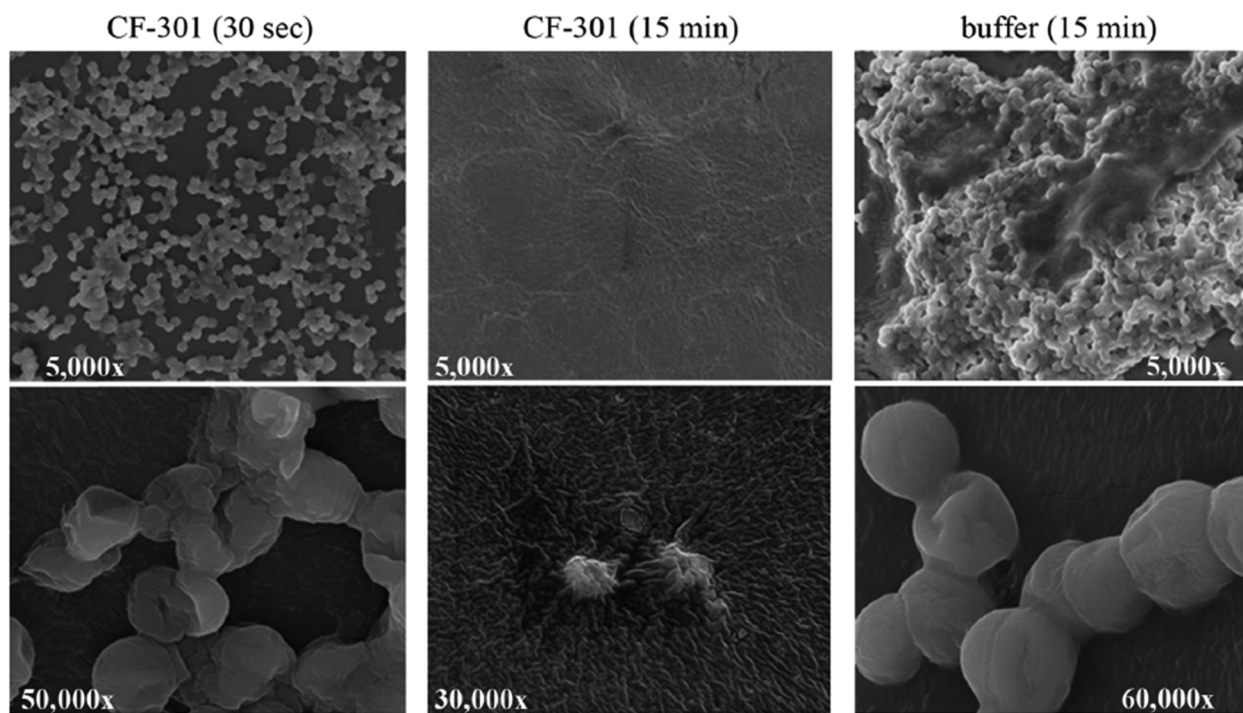


Figure 3: MRSA strain ATCC BAA-42 biofilms developed for 3 days in the catheter lumen against CF-301 [46].

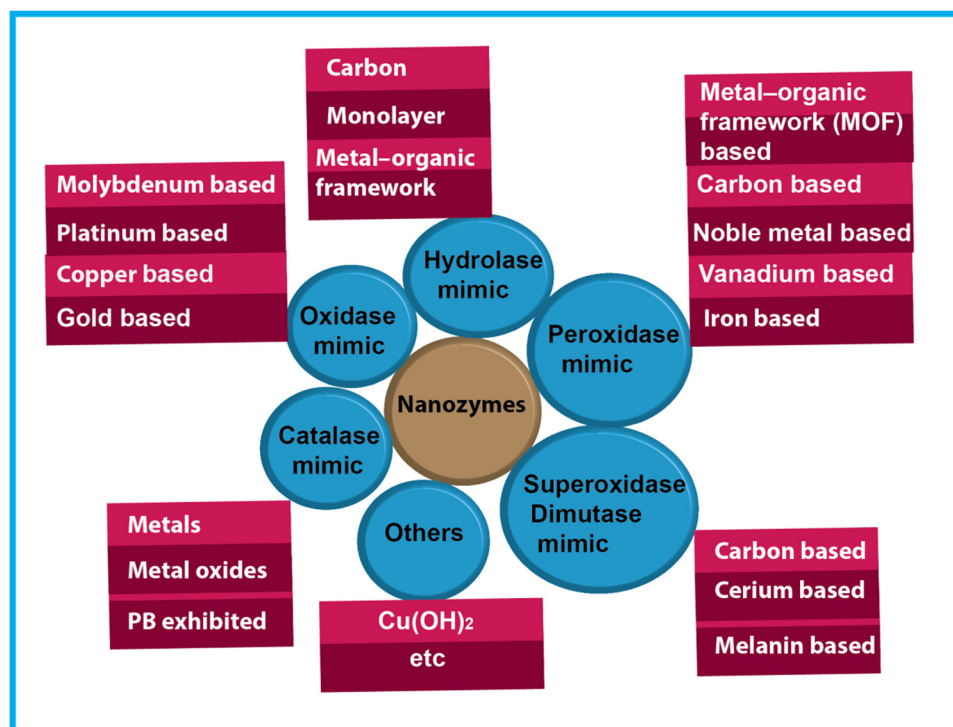


Figure 4: Types of nanozymes.

enzymes like low cost, high stability, and so on [29,53–55]. Artificial enzymes have contributed to the area of medical science. Subsequently, a number of nanozymes have been found, and from those, some are shown in Figure 4 with types of nanozymes. New scientific findings for artificial enzymes are as follows:

- 1) Sensing, ion determination, molecule determination, nucleic acid determination, protein determination, and cancer cell determination.
- 2) Environmental intervention and degradation of contamination in the environment.
- 3) Nanozymes for cancer treatment and the most important for this report, antibacterial, and antibiofilm. The basic of nanozymes antimicrobial properties was necessary before the antibacterial flick performance of nanozymes.

2.2.1 Antibacterial performance of nanozymes

A series of nanozymes have been discovered for antibacterial activities and biofilms such as single atom nanozyme [56], *o*-carbon nanotubes [13], copper oxide nanorods [57], GQDs [58], gold nanoclusters [59], platinum hollow nanodendrites [58], hybrid GQD AgNPs [60], mesoporous silica/AuNPs or porous Pt/AuNPs [31], PEG-MoS₂ nanoflowers [61], CaO₂/H-G@alginate [62], IOPs [63], DMAE [64], V₂O₅

nanowires [31], CeO_{2-x} nanorod [65], PAN/FeNPs nanofibers [108], *etc.* In brief, the antibacterial mechanism of nanozymes is based on the release of $\cdot\text{OH}$ and $\cdot\text{O}_2^-$. Figure 5

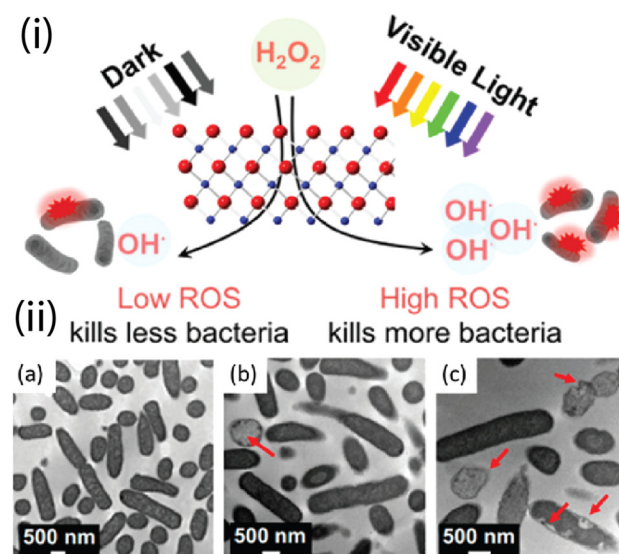


Figure 5: (i) Illustration of antibacterial mechanism of nanozyme; (ii) transmission electron microscopic images of copper oxide nanorods against *E. coli* at various treatments: (a) control; (b) copper oxide nanorods with hydrogen peroxide in the absence of light; and (c) copper oxide nanorods with hydrogen peroxide in the presence of light [57].

demonstrates oxidase- and peroxidase-like activity and death of bacteria due to $\cdot\text{OH}$ release; however, in the absence of light, the dark copper oxide nanorods release lower ROS than visible light and also affect the death rate of bacteria. The bactericide effect of artificial enzymes initiates from their capability to regulate the phase of ROS like $-\text{OH}$ and $\cdot\text{O}_2^-$ [66]. Due to higher oxidation potentiality, ROS may affect bacteria and biofilms; indiscriminating disturbs the function of biomolecules inside or in the cell resulting in damaging of the cytoplasmic membrane and the death of bacteria. Enhancing the ROS and the death of bacteria faster destroys bacterial cell membrane and makes bacterial resistance rate lower toward nanozyme [30,61,67]. Moreover, it is well known that only hydrogen peroxide can work as an antimicrobial agent, but it may have an effect on tissues as well; therefore, the presence of nanozymes having an activity like peroxidase has fortune in the biomedical era [61,67–69].

The catalytic activity of nanozymes can be tuned by altering pH, temperature, ionic changes, light and morphology [70–80]. It is well described in a previous report by Huang *et al.* [33]. Many factors have been described well but still, nanozyme is a hot research topic and it is gaining importance day by day. Considering these challenges, Gao and co-workers have introduced the impact of metal states and their antimicrobial effects that are predominately little known [81]. Further investigation has shown that copper/carbon is a mimic enzyme material and is described as copper/carbon state-dependent nanozymes; Cu^0 has shown better enzyme-like activity than Cu^{2+} . Bactericide performances of copper/carbon hybrid nanozyme showed that ROS and transfer of ions from Cu^{2+} are responsible for the inhibition of bacteria. However, Cu^{2+} release can inhibit only gram-negative bacteria, and ROS generation can kill both gram-positive and gram-negative bacteria; therefore, controlling the state of metal can be beneficial for selective bacteria destruction. In addition, injury of the intestine caused by *S. typhimurium* can be treated by copper/carbon artificial enzyme [81]. Furthermore, iron oxide has been used against the influenza virus, which had a great breakthrough in the nanozyme area [82]. An excellent impact on the viral lipid envelope was demonstrated to deactivate it and give shelter from the contagion of the virus. The mechanism of antiviral nanozymes can be understood through the oxidative breakdown of lipids presented in Figure 6. $\text{Ag@Fe}_3\text{O}_4$ core@shell has been developed for antibiofilm activity [83], due to the well-known magnetic property of Fe_3O_4 antimicrobial activity, which can be enhanced by external magnetic force. Hence,

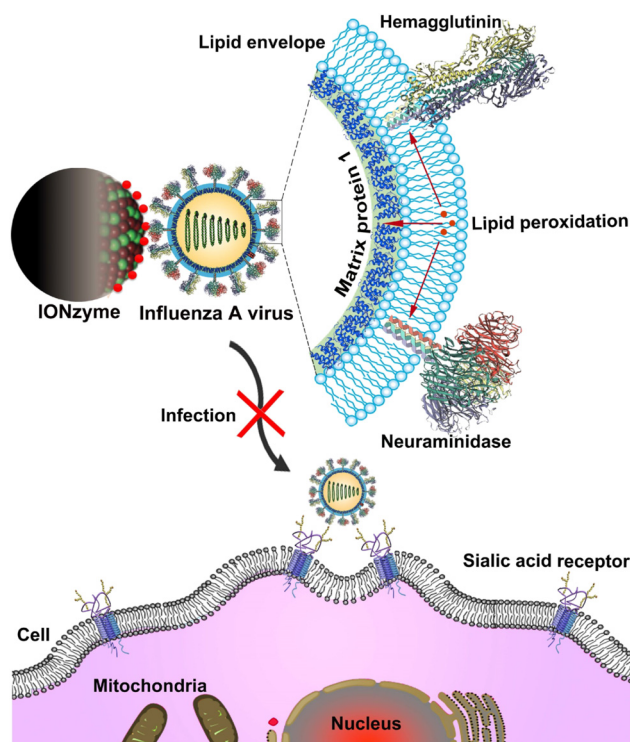


Figure 6: Illustration of viral oxidative breakdown of lipids by iron oxide nanozyme [82].

the magnetic property of Fe_3O_4 and silver ions released from silver are responsible for the killing of microbes [84].

2.3 Photocatalysis as an antibacterial agent

Photocatalysis plays an important role in microbial degradation, but it is well known for the degradation of pollutants [85,86]. Herein, we have briefly described the best photocatalytic antimicrobial agent. Ferrite nanoparticles (NPs) have been examined as antibacterial and antibiofilm [87] agent that is an excellent catalyst for many reactions [88]. The $\text{CeO}_2\text{--TiO}_2$ has shown excellent antibacterial effect against gram-positive and gram-negative bacteria in the presence of light [89]. However, ferrite NPs and $\text{CeO}_2\text{--TiO}_2$ mechanism has not been described. The possibility of bacterial disruption is due to photocatalytic activity of relevant stuff such as TiO_2 . TiO_2 possesses low antibacterial activity due to a band gap [90], but it can be improved by the addition of substitutes or increasing visible light [91]. Wang *et al.* have designed photocatalytic CT/poly(vinyl alcohol) (PVA) hydrogel as an antimicrobial agent and wound dressing material [92]. The antibacterial effect of CT/PVA hydrogel

is due to the generation of $-OH$ and ROS. Zinc oxide is widely utilized as a photocatalytic and antimicrobial agent due to its unique properties such as being cheaper, atoxic, *etc.* [93–101]. The toxic effect of ZnO is restricted, and bacterial death is usually due to the generation of ROS and disrupting bacteria. Moreover, nano-ZnO is atoxic to mammalian cells and more effective toward bacteria [102–104]. Commonly employed organic compounds such as pigments, dyes, and catalysts in organic synthesis are known as Schiff bases. It has been demonstrated with a wide range of antimicrobial activities [105,106]. Recently, it has been studied against MDR bacteria (MDRB), while the mechanism is not well defined [107]. Furthermore, the attachment of groups, such as aldehyde or amino with cell wall of bacteria, could be the reason for bacterial inhibition.

3 Conclusion

In this report, recent development in the area of catalytic antibacterial strategies has been presented. Briefly, catalytic antibacterial strategies including natural enzymes, nanozymes, and photocatalytic antibacterial strategies have been discussed. As a result, ROS, $-OH$, aldehyde or amino can inhibit microbes and Cu^{2+} can only inhibit gram-negative bacteria. Iron oxide possesses antiviral activity, and carbon-based material has been reported as a great enzyme mimic material. Yet researchers have many challenges to produce novel antibiotics such as for nCovid-2019.

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References

- [1] Hook AL, Chang CY, Yang J, Atkinson S, Langer R, Anderson DG, et al. Discovery of novel materials with broad resistance to bacterial attachment using combinatorial polymer microarrays. *Adv Mater.* 2013;25(18):2542–7.
- [2] Pham VTH, Truong VK, Quinn MDJ, Notley SM, Guo Y, Baulin VA, et al. Graphene induces formation of pores that kill spherical and rod-shaped bacteria. *ACS Nano.* 2015;9(8):8458–67.
- [3] Zhao Y, Chen Z, Chen Y, Xu J, Li J, Jiang X. Synergy of non-antibiotic drugs and pyrimidinethiol on gold nanoparticles against superbugs. *J Am Chem Soc.* 2013;135(35):12940–3.
- [4] Barraud N, Kardak BG, Yepuri NR, Howlin RP, Webb JS, Faust SN, et al. Cephalosporin-3'-diazoniumdiolates: targeted NO-donor prodrugs for dispersing bacterial biofilms. *Angew Chem Int Ed.* 2012;51(36):9057–60.
- [5] Durmus NG, Taylor EN, Kummer KM, Webster TJ. Enhanced efficacy of superparamagnetic iron oxide nanoparticles against antibiotic-resistant biofilms in the presence of metabolites. *Adv Mater.* 2013;25(40):5706–13.
- [6] Li Y, Fukushima K, Coady DJ, Engler AC, Liu S, Huang Y, et al. Broad-spectrum antimicrobial and biofilm-disrupting hydrogels: stereocomplex-driven supramolecular assemblies. *Angew Chem Int Ed.* 2013;52(2):674–8.
- [7] Chen WY, Chang HY, Lu JK, Huang YC, Harroun SG, Tseng YT, et al. Self-assembly of antimicrobial peptides on gold nanodots: against multidrug-resistant bacteria and wound-healing application. *Adv Funct Mater.* 2015;25(46):7189–99.
- [8] Liu R, Chen X, Falk SP, Masters KS, Weisblum B, Gellman SH. Nylon-3 polymers active against drug-resistant *Candida albicans* biofilms. *J Am Chem Soc.* 2015;137(6):2183–6.
- [9] Zhao Y, Tian Y, Cui Y, Liu W, Ma W, Jiang X. Small molecule-capped gold nanoparticles as potent antibacterial agents that target gram-negative bacteria. *J Am Chem Soc.* 2010;132(35):12349–56.
- [10] Fasciani C, Jazmin Silvero M, Anghel MA, Arguello GA, Becerra MC, Scaiano JC. Aspartame-stabilized gold–silver bimetallic biocompatible nanostructures with plasmonic photothermal properties, antibacterial activity, and long-term stability. *J Am Chem Soc.* 2014;136(50):17394–7.
- [11] Leidinger P, Treptow J, Hagens K, Eich J, Zehethofer N, Schwudke D, et al. Isoniazid@ Fe₂O₃ nanocontainers and their antibacterial effect on tuberculosis mycobacteria. *Angew Chem Int Ed.* 2015;54(43):12597–601.
- [12] Xiong MH, Li YJ, Bao Y, Yang XZ, Hu B, Wang J. Bacteria-responsive multifunctional nanogel for targeted antibiotic delivery. *Adv Mater.* 2012;24(46):6175–80.
- [13] Wang H, Li P, Yu D, Zhang Y, Wang Z, Liu C, et al. Unraveling the enzymatic activity of oxygenated carbon nanotubes and their application in the treatment of bacterial infections. *Nano Lett.* 2018;18(6):3344–51.
- [14] Ivanova EP, Hasan J, Webb HK, Gervinskas G, Juodkazis S, Truong VK, et al. Bactericidal activity of black silicon. *Nat Commun.* 2013;4(1):1–7.
- [15] Richter AP, Brown JS, Bharti B, Wang A, Gangwal S, Houck K, et al. An environmentally benign antimicrobial nanoparticle based on a silver-infused lignin core. *Nat Nanotechnol.* 2015;10(9):817–23.
- [16] Zhao Y, Ye C, Liu W, Chen R, Jiang X. Tuning the composition of AuPt bimetallic nanoparticles for antibacterial application. *Angew Chem Int Ed.* 2014;53(31):8127–31.
- [17] Lucky S, Swarnalatha KC, Soo, Zhang Y. Nanoparticles in photodynamic therapy. *Chem Rev.* 2015;115(4):1990–2042.
- [18] Strassert CA, Otter M, Albuquerque RQ, Hoene A, Vida Y, Maier B, et al. Photoactive hybrid nanomaterial for targeting,

- labeling, and killing antibiotic-resistant bacteria. *Angew Chem Int Ed.* 2009;48(42):7928–31.
- [19] Su HL, Chou C-C, Hung DJ, Lin S-H, Pao I-C, Lin J-H, et al. The Disruption of bacterial membrane integrity through ROS generation induced by nanohybrids of Silver and Clay2. *Biomaterials.* 2009;30:5979.
- [20] Zhu C, Yang Q, Liu L, Lv F, Li S, Yang G, et al. Multifunctional cationic poly (p-phenylene vinylene) polyelectrolytes for selective recognition, imaging, and killing of bacteria over mammalian cells. *Adv Mater.* 2011;23(41):4805–10.
- [21] Becker MH, Harris RN. Cutaneous bacteria of the redback salamander prevent morbidity associated with a lethal disease. *PLoS One.* 2010;5(6):e10957.
- [22] Mishra S, Imlay J. Why do bacteria use so many enzymes to scavenge hydrogen peroxide? *Arch Biochem Biophys.* 2012;525(2):145–60.
- [23] Zhang C, Zhao K, Bu W, Ni D, Liu Y, Feng J, et al. Marriage of scintillator and semiconductor for synchronous radiotherapy and deep photodynamic therapy with diminished oxygen dependence. *Angew Chem Int Ed.* 2015;54(6):1770–4.
- [24] Gao L, Giglio KM, Nelson JL, Sondermann H, Travis AJ. Ferromagnetic nanoparticles with peroxidase-like activity enhance the cleavage of biological macromolecules for bio-film elimination. *Nanoscale.* 2014;6(5):2588–93.
- [25] Dickinson BC, Chang CJ. Chemistry and biology of reactive oxygen species in signaling or stress responses. *Nat Chem Biol.* 2011;7(8):504–11.
- [26] Loo AEK, Wong YT, Ho R, Wasser M, Du T, Ng WT, et al. Effects of hydrogen peroxide on wound healing in mice in relation to oxidative damage. *PLoS One.* 2012;7(11):e49215.
- [27] Gupta A, Mumtaz S, Li CH, Hussain I, Rotello VM. Combatting antibiotic-resistant bacteria using nanomaterials. *Chem Soc Rev.* 2019;48(2):415–27.
- [28] Li P, Li J, Feng X, Li J, Hao Y, Zhang J, et al. Metal-organic frameworks with photocatalytic bactericidal activity for integrated air cleaning. *Nat Commun.* 2019;10(1):1–10, 2177.
- [29] Lin Y, Ren J, Qu X. Catalytically active nanomaterials: a promising candidate for artificial enzymes. *Acc Chem Res.* 2014;47(4):1097–105.
- [30] Liu C, Kong D, Hsu PC, Yuan H, Lee HW, Liu Y, et al. Rapid water disinfection using vertically aligned MoS₂ nanofilms and visible light. *Nat Nanotechnol.* 2016;11(12):1098–104.
- [31] Chen Z, Wang Z, Ren J, Qu X. Enzyme mimicry for combating bacteria and biofilms. *Acc Chem Res.* 2018;51(3):789–99.
- [32] Dong H, Fan Y, Zhang W, Gu N, Zhang Y. Catalytic mechanisms of nanozymes and their applications in biomedicine. *Bioconjugate Chem.* 2019;30(5):1273–96.
- [33] Huang Y, Ren J, Qu X. Nanozymes: classification, catalytic mechanisms, activity regulation, and applications. *Chem Rev.* 2019;119(6):4357–412.
- [34] Sun H, Zhou Y, Ren J, Qu X. Carbon nanozymes: enzymatic properties, catalytic mechanism, and applications. *Angew Chem Int Ed.* 2018;57(30):9224–37.
- [35] Bornscheuer UT, Huisman GW, Kazlauskas RJ, Lutz S, Moore JC, Robins K. Engineering the third wave of biocatalysis. *Nature.* 2012;485(7397):185–94.
- [36] Breaker RR. DNA enzymes. *Nat Biotechnol.* 1997;15(5):427–31.
- [37] Behrens M, Studt F, Kasatkin I, Kühl S, Hävecker M, Abild-Pedersen F, et al. The active site of methanol synthesis over Cu/ZnO/Al₂O₃ industrial catalysts. *Science.* 2012;336(6083):893–7.
- [38] Genet JP. Asymmetric catalytic hydrogenation. Design of new Ru catalysts and chiral ligands: from laboratory to industrial applications. *Acc Chem Res.* 2003;36(12):908–18.
- [39] Kirby AJ. Efficiency of proton transfer catalysis in models and enzymes. *Acc Chem Res.* 1997;30(7):290–6.
- [40] Meunier B, De Visser SP, Shaik S. Mechanism of oxidation reactions catalyzed by cytochrome P450 enzymes. *Chem Rev.* 2004;104(9):3947–80.
- [41] Abuchowski A, Kazo GM, Verhoest Jr CR, Van Es T, Kafkewitz D, Nucci ML, et al. Cancer therapy with chemically modified enzymes. I. Antitumor properties of polyethylene glycol-asparaginase conjugates. *Cancer Biochem Biophys.* 1984;7(2):175–86.
- [42] Choct M. Enzymes for the feed industry: past, present and future. *World Poult Sci J.* 2006;62(1):5–16.
- [43] Gurung N, Ray S, Bose S, Rai V. A broader view: microbial enzymes and their relevance in industries, medicine, and beyond. *BioMed Res Int.* 2013;2013:329121.
- [44] Posorske LH. Industrial-scale application of enzymes to the fats and oil industry. *J Am Oil Chem Soc.* 1984;61(11):1758–60.
- [45] Düring K, Petra P, Mahn A, Brinkmann O, Gieffers W. The non-enzymatic microbicidal activity of lysozymes. *FEBS Lett.* 1999;449(2–3):93–100.
- [46] Schuch R, Khan BK, Raz A, Rotolo JA, Wittekind M. Bacteriophage lysin CF-301, a potent antistaphylococcal biofilm agent. *Antimicrob Agents Chemother.* 2017;61(7):e02666-16.
- [47] Schmelcher M, Loessner MJ. Bacteriophage endolysins: applications for food safety. *Curr Opin Biotechnol.* 2016;37:76–87.
- [48] Fischetti VA. Development of phage lysins as novel therapeutics: a historical perspective. *Viruses.* 2018;10(6):310.
- [49] Fischetti VA. Bacteriophage lysins as effective antibacterials. *Curr Opin Microbiol.* 2008;11(5):393–400.
- [50] Breslow R, Overman LE. “Artificial enzyme” combining a metal catalytic group and a hydrophobic binding cavity. *J Am Chem Soc.* 1970;92(4):1075–7.
- [51] Bleeker EA, de Jong WH, Geertsma RE, Groenewold M, Heugens EH, Koers-Jacquemijns M, et al. Considerations on the EU definition of a nanomaterial: science to support policy making. *Regul Toxicol Pharmacol.* 2013;65(1):119–25.
- [52] Maynard AD. Don't define nanomaterials. *Nature.* 2011;475(7354):31.
- [53] Wei H, Wang E. Nanomaterials with enzyme-like characteristics (nanozymes): next-generation artificial enzymes. *Chem Soc Rev.* 2013;42(14):6060–93.
- [54] Wu J, Wang X, Wang Q, Lou Z, Li S, Zhu Y, et al. Nanomaterials with enzyme-like characteristics (nanozymes): next-generation artificial enzymes (II). *Chem Soc Rev.* 2019;48(4):1004–76.
- [55] Zhou Y, Liu B, Yang R, Liu J. Filling in the gaps between nanozymes and enzymes: challenges and opportunities. *Bioconjugate Chem.* 2017;28(12):2903–9.
- [56] Xu B, Wang H, Wang W, Gao L, Li S, Pan X, et al. A single-atom nanozyme for wound disinfection applications. *Angew Chem.* 2019;131(15):4965–70.
- [57] KarimMdN, Singh M, Weerathunge P, Bian P, Zheng R, Dekiwadia C, et al. Visible-light-triggered reactive-oxygen-

- species-mediated antibacterial activity of peroxidase-mimic CuO nanorods. *ACS Appl Nano Mater.* 2018;1(4):1694–704.
- [58] Ge C, Wu R, Chong Y, Fang G, Jiang X, Pan Y, et al. Synthesis of Pt hollow nanodendrites with enhanced peroxidase-like activity against bacterial infections: implication for wound healing. *Adv Funct Mater.* 2018;28(28):1801484.
- [59] Zheng Y, Liu W, Qin Z, Chen Y, Jiang H, Wang X. Mercaptopuridine-conjugated gold nanoclusters as nanoantibiotics for combating multidrug-resistant superbugs. *Bioconjugate Chem.* 2018;29(9):3094–103.
- [60] Chen S, Quan Y, Yu YL, Wang JH. Graphene quantum dot/silver nanoparticle hybrids with oxidase activities for antibacterial application. *ACS Biomater Sci Eng.* 2017;3(3):313–21.
- [61] Yin W, Yu J, Lv F, Yan L, Zheng LR, Gu Z, et al. Functionalized nano-MoS₂ with peroxidase catalytic and near-infrared photothermal activities for safe and synergetic wound antibacterial applications. *ACS Nano.* 2016;10(12):11000–11.
- [62] Yan Z, Bing W, Ding C, Dong K, Ren J, Qu X. A H₂O₂-free depot for treating bacterial infection: localized cascade reactions to eradicate biofilms in vivo. *Nanoscale.* 2018;10(37):17656–62.
- [63] Geilich BM, Gelfat I, Sridhar S, van de Ven AL, Webster TJ. Superparamagnetic iron oxide-encapsulating polymersome nanocarriers for biofilm eradication. *Biomaterials.* 2017;119:78–85.
- [64] Günther M, Juchum M, Kelter G, Fiebig H, Laufer S. Lung cancer: EGFR inhibitors with low nanomolar activity against a therapy-resistant L858R/T790M/C797S mutant. *Angew Chem Int Ed.* 2016;55(36):10890–4.
- [65] Jiang H, Chen Z, Cao H, Huang Y. Peroxidase-like activity of chitosan stabilized silver nanoparticles for visual and colorimetric detection of glucose. *Analyst.* 2012;137(23):5560–4.
- [66] Bing W, Sun H, Yan Z, Ren J, Qu X. Programmed bacteria death induced by carbon dots with different surface charge. *Small.* 2016;12(34):4713–8.
- [67] Gehring J, Trepka B, Klinkenberg N, Bronner H, Schleheck D, Polarz S. Sunlight-triggered nanoparticle synergy: teamwork of reactive oxygen species and nitric oxide released from mesoporous organosilica with advanced antibacterial activity. *J Am Chem Soc.* 2016;138(9):3076–84.
- [68] Yao J, Cheng Y, Zhou M, Zhao S, Lin S, Wang X, et al. ROS scavenging Mn₃O₄ nanozymes for in vivo anti-inflammation. *Chem Sci.* 2018;9(11):2927–33.
- [69] Ji H, Dong K, Yan Z, Ding C, Chen Z, Ren J, et al. Bacterial hyaluronidase self-triggered prodrug release for chemophotothermal synergistic treatment of bacterial infection. *Small.* 2016;12(45):6200–6.
- [70] Chen Z, Yin JJ, Zhou YT, Zhang Y, Song L, Song M, et al. Dual enzyme-like activities of iron oxide nanoparticles and their implication for diminishing cytotoxicity. *Acs Nano.* 2012;6(5):4001–12.
- [71] Asati A, Kaftanis C, Santra S, Manuel Perez J. pH-tunable oxidase-like activity of cerium oxide nanoparticles achieving sensitive fluorogenic detection of cancer biomarkers at neutral pH. *Anal Chem.* 2011;83(7):2547–53.
- [72] Fan K, Wang H, Xi J, Liu Q, Meng X, Duan D, et al. Optimization of Fe₃O₄ nanozyme activity via single amino acid modification mimicking an enzyme active site. *Chem Commun.* 2017;53(2):424–7.
- [73] Li W, Chen B, Zhang H, Sun Y, Wang J, Zhang J, et al. BSA-stabilized Pt nanozyme for peroxidase mimetics and its application on colorimetric detection of mercury (II) ions. *Biosens Bioelectron.* 2015;66:251–8.
- [74] Liu Y, Zheng Y, Ding D, Guo R. Switching peroxidase-mimic activity of protein stabilized platinum nanozymes by sulfide ions: substrate dependence, mechanism, and detection. *Langmuir.* 2017;33(48):13811–20.
- [75] Liu Y, Xiang Y, Zhen Y, Guo R. Halide ion-induced switching of gold nanozyme activity based on Au–X interactions. *Langmuir.* 2017;33(25):6372–81.
- [76] Puvvada N, Kumar Panigrahi P, Mandal D, Pathak A. Shape dependent peroxidase mimetic activity towards oxidation of pyrogallol by H₂O₂. *RSC Adv.* 2012;2(8):3270–3.
- [77] Luo W, Zhu C, Su S, Li D, He Y, Huang Q, et al. Self-catalyzed, self-limiting growth of glucose oxidase-mimicking gold nanoparticles. *Acs Nano.* 2010;4(12):7451–8.
- [78] Tao Y, Ju E, Ren J, Qu X. Polypyrrole nanoparticles as promising enzyme mimics for sensitive hydrogen peroxide detection. *Chem Commun.* 2014;50(23):3030–2.
- [79] Ge C, Fang G, Shen X, Chong Y, Wamer WG, Gao X, et al. Facet energy *versus* enzyme-like activities: the unexpected protection of palladium nanocrystals against oxidative damage. *Acs Nano.* 2016;10(11):10436–45.
- [80] Gao L, Zhuang J, Nie L, Zhang J, Zhang Y, Gu N, et al. Intrinsic peroxidase-like activity of ferromagnetic nanoparticles. *Nat Nanotechnol.* 2007;2(9):577–83.
- [81] Xi J, Wei G, An L, Xu Z, Xu Z, Fan L, et al. Copper/carbon hybrid nanozyme: tuning catalytic activity by the copper state for antibacterial therapy. *Nano Lett.* 2019;19(11):7645–54.
- [82] Qin T, Ma R, Yin Y, Miao X, Chen S, Fan K, et al. Catalytic inactivation of influenza virus by iron oxide nanozyme. *Theranostics.* 2019;9(23):6920.
- [83] Zhang C, Du C, Liao JY, Gu Y, Gong Y, Pei J, et al. Synthesis of magnetite hybrid nanocomplexes to eliminate bacteria and enhance biofilm disruption. *Biomater Sci.* 2019;7(7):2833–40.
- [84] Chernousova S, Epple M. Silver as antibacterial agent: ion, nanoparticle, and metal. *Angew Chem Int Ed.* 2013;52(6):1636–53.
- [85] Chacko JT, Subramaniam K. Enzymatic degradation of azo dyes—a review. *Int J Environ Sci.* 2011;1(6):1250.
- [86] Jauhar S, Dhiman M, Bansal S, Singhal S. Mn³⁺ ion in perovskite lattice: a potential Fenton's reagent exhibiting remarkably enhanced degradation of cationic and anionic dyes. *J Sol–Gel Sci Technol.* 2015;75(1):124–33.
- [87] Maksoud MIAA, El-Sayyad GS, Ashour AH, El-Batal AI, Elsayed MA, Gobara M, et al. Antibacterial, antibiofilm, and photocatalytic activities of metals-substituted spinel cobalt ferrite nanoparticles. *Microb Pathogen.* 2019;127:144–58.
- [88] Naghikhani R, Nabyouni G, Ghanbari D. Simple and green synthesis of CuFe₂O₄–CuO nanocomposite using some natural extracts: photo-degradation and magnetic study of nanoparticles. *J Mater Sci Mater Electron.* 2018;29(6):4689–703.
- [89] Kasinathan K, Kennedy J, Elayaperumal M, Henini M, Malik M. Photodegradation of organic pollutants RhB dye using UV simulated sunlight on ceria based TiO₂ nanomaterials for antibacterial applications. *Sci Rep.* 2016;6(1):1–12.

- [90] Ananpattarachai J, Kajitvichyanukul P, Seraphin S. Visible light absorption ability and photocatalytic oxidation activity of various interstitial N-doped TiO₂ prepared from different nitrogen dopants. *J Hazard Mater.* 2009;168(1):253–61.
- [91] Ananpattarachai J, Boonto Y, Kajitvichyanukul P. Visible light photocatalytic antibacterial activity of Ni-doped and N-doped TiO₂ on *Staphylococcus aureus* and *Escherichia coli* bacteria. *Environ Sci Pollut Res.* 2016;23(5):4111–9.
- [92] Wang Z, Wang Y, Peng X, He Y, Wei L, Su W, et al. Photocatalytic antibacterial agent incorporated double-network hydrogel for wound healing. *Colloids Surf B Biointerfaces.* 2019;180:237–44.
- [93] Lee KM, Lai CW, Ngai KS, Juan JC. Recent developments of zinc oxide based photocatalyst in water treatment technology: a review. *Water Res.* 2016;88:428–48.
- [94] Cun T, Dong C, Huang Q. Ionothermal precipitation of highly dispersive ZnO nanoparticles with improved photocatalytic performance. *Appl Surf Sci.* 2016;384:73–82.
- [95] Abbas KN, Bidin N. Morphological driven photocatalytic activity of ZnO nanostructures. *Appl Surf Sci.* 2017;394:498–508.
- [96] Khademalrasool M, Farbod M. Preparation of ZnO nanoparticles/Ag nanowires nanocomposites as plasmonic photocatalysts and investigation of the effect of concentration and diameter size of Ag nanowires on their photocatalytic performance. *J Alloy Compd.* 2016;664:707–14.
- [97] Gancheva M, Markova-Velichkova M, Atanasova G, Kovacheva D, Uzunov I, Cukeva R. Design and photocatalytic activity of nanosized zinc oxides. *Appl Surf Sci.* 2016;368:258–66.
- [98] Ambika S, Sundrarajan M. Antibacterial behaviour of Vitex negundo extract assisted ZnO nanoparticles against pathogenic bacteria. *J Photochem Photobiol B Biol.* 2015;146:52–7.
- [99] Panigrahi J, Behera D, Mohanty I, Subudhi U, Nayak BB, Acharya BS. Radio frequency plasma enhanced chemical vapor based ZnO thin film deposition on glass substrate: a novel approach towards antibacterial agent. *Appl Surf Sci.* 2011;258(1):304–11.
- [100] Zhu P, Weng Z, Li X, Liu X, Wu S, Yeung KWK, et al. Biomedical applications of functionalized ZnO nanomaterials: from biosensors to bioimaging. *Adv Mater Interfaces.* 2016;3(1):1500494.
- [101] Svetlichnyi V, Shabalina A, Lapin I, Goncharova D, Nemoykina A. ZnO nanoparticles obtained by pulsed laser ablation and their composite with cotton fabric: Preparation and study of antibacterial activity. *Appl Surf Sci.* 2016;372:20–9.
- [102] George S, Pokhrel S, Xia T, Gilbert B, Ji Z, Schowalter M, et al. Use of a rapid cytotoxicity screening approach to engineer a safer zinc oxide nanoparticle through iron doping. *ACS Nano.* 2010;4(1):15–29.
- [103] Kononenko V, Repar N, Marušič N, Drašler B, Romih T, Hočvar S, et al. Comparative in vitro genotoxicity study of ZnO nanoparticles, ZnO macroparticles and ZnCl₂ to MDCK kidney cells: Size matters. *Toxicol Vitro.* 2017;40:256–63.
- [104] Hackenberg S, Scherzed A, Technau A, Kessler M, Froelich K, Ginzkey C, et al. Cytotoxic, genotoxic and pro-inflammatory effects of zinc oxide nanoparticles in human nasal mucosa cells in vitro. *Toxicol Vitro.* 2011;25(3):657–63.
- [105] Dhar DN, Taploo CL. Schiff-bases and their applications. *J Sci Ind Res.* 1982;41(8):501–6.
- [106] Przybylski P, Huczyński AW, Pyta KK, Brzezinski B, Bartl F. Biological properties of Schiff bases and azo derivatives of phenols. *Curr Org Chem.* 2009;13:124–48.
- [107] Anaconda JR, Ruiz K, Loroño M, Celis F. Antibacterial activity of transition metal complexes containing a tridentate NNO phenoxymethylpenicillin-based Schiff base. An anti-MRSA iron (II) complex. *Appl Organomet Chem.* 2019;33(4):e4744.
- [108] Kalwar K, Juqun JX, Dandan L, Gao L. Fabrication of PAN/FeNPs electrospun nanofibers: Nanozyme and an efficient antimicrobial agent. *Mater Today Commun.* 2021;26:102168.