

Review

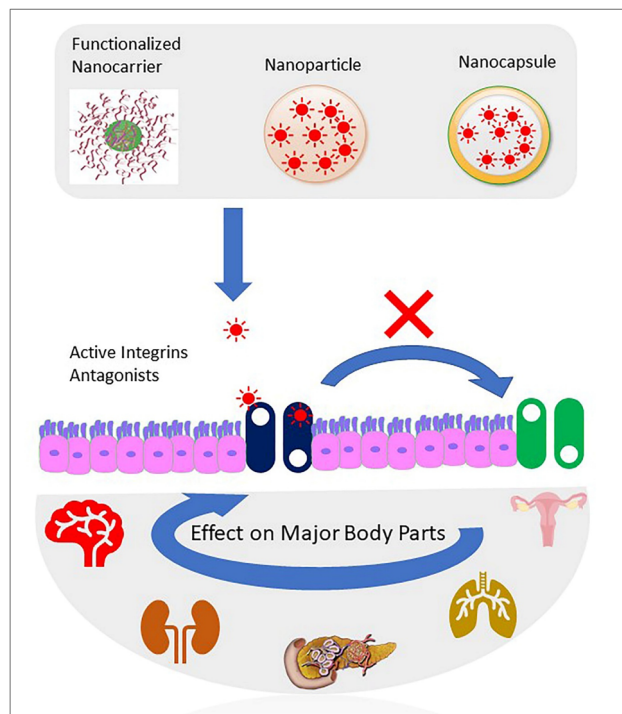
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Contemporary nano-architected drugs and leads for $\alpha\text{v}\beta 3$ integrin-based chemotherapy: Rationale and retrospect

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Abstract: The integrins belong to the cell-surface polypeptide family and are the mediating partners among the cells, and extracellular matrix (ECM). They are also involved in the biological processes of cell migration, wound healing, blood clotting, immunological response generation, tissue morphogenesis, leucocyte reticulations, and angiogenesis and are therefore very relevant in stem cell technology and are useful as biomarkers, diagnostic probes, and drug-target ligands. The $\alpha\text{v}\beta 3$ (*alpha-nu-beta3*) integrin antagonists are an excellent target example for designing and developing newer drug candidates, drug leads and templates for various diseases, and physiological malfunctioning, including cancers. The current review examines the $\alpha\text{v}\beta 3$ integrin structural features involved in the drug design and its antagonistic ligands and highlights the development of anti- $\alpha\text{v}\beta 3$ integrin-antagonists as nano-architectural design-based nanomedicine, especially for cancer chemotherapy. The perspectival review discusses the $\alpha\text{v}\beta 3$ integrin structure, mode of action, involved pathways, and the concepts utilized in nanomedicine design, and ligands related to integrins. It also covers the latest thyrointegrin approaches toward the development of anti-angiogenesis agents and entails the anti-angiogenesis approach to cancer growth inhibition through targeting by the anti-integrin ligands and related chemical entities. The current perspective on the nano-



Graphical abstract: The $\alpha\text{v}\beta 3$ integrin antagonistic ligands, *i.e.*, drugs as nano-architectural design-based nanomedicine, especially for cancer chemotherapy, are discussed.

architectural design approach for the known anti-integrin compounds is also outlined.

Keywords: integrins, $\alpha\text{v}\beta 3$, antagonists, thyrointegrins, therapeutics, nanomedicine

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1 Introduction

The functioning of multicellular organisms involves complex interactions within the cells, in the extracellular matrix (ECM), surrounding cells, and other bio-entities in the ECM arena, and integrins are the foremost bio-entities involved in these actions [1]. In multicellular

organizations, cells adherence, facilitated through the integrins, has a distinctive and diverse role for a number of biological level feedback, signaling, physico-chemical, physico-mechanical, and biomechanistics aspects of cell and organ functions, such as cell divisions, maintenance of tissue integrity, embryonic cell development during embryo development, cell migration, cell proliferation, blood cell involvement, blood coagulation, and immune system functioning [2]. Any modification in the cellular structure, functions, cellular etiology, and cellular adhesion may cause several diseases including atherosclerosis, inflammatory disorders, and different types of cancers. This has helped to identify the integrins as an appealing target for the discovery and development of new and improved therapeutic agents.

Integrins, the vital component in cells' signal transduction and biochemical functioning at physiological and biochemical levels, have been located in several organs and their tissues and are expressed as part of the cell function requirements, immunological dealings, and signal transduction activities. Several integrin types, with specificity to different receptors, *i.e.*, RGD, collagen, laminin, and leucocyte-specifics, are known (Figure 1). Among the vertebrates, including human's integrins, the $\alpha\text{v}\beta 3$ (*alpha-nu-beta3*) is the most commonly expressed integrin in proliferative endothelial cell types, vascular smooth muscle cells, macrophages, and the monocytes.

These integrins are involved in signal transduction through different signaling pathways within and across the cells, with the ECM components, and in blood cells in various tissues and organs. The integrins are also involved in angiogenesis and tumor cells of several organs in facilitating types of cancer cell proliferation and cancer metastasis [3].

2 Integrins: Structure, heterodimeric junction, and functional domains

Structurally, the integrins are a family of polypeptidic receptors and mainly function as cell adhesion receptors. The integrins are noncovalently linked, heterodimeric molecules incorporating an α and a β subunit as part of their tertiary structure and are embedded across the plasma membrane. These portions contain a high molecular weight fraction of the integrin structure outside of the cell, and the shorter ratio of the structural part lies in the cytoplasm. Vertebrates' origin integrin entities are composed of 24 α subunit types and 9 β subunits with specified structure types till date with different α - β combinations of receptors with different binding properties

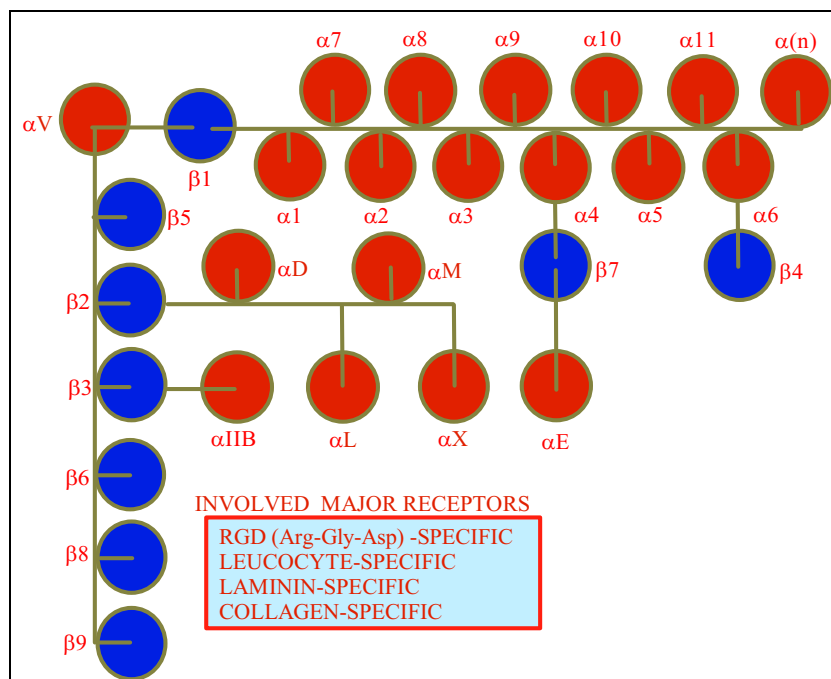


Figure 1: Integrin superfamily of vertebrates' origins including humans. With the α and β subunits combining to form heterodimeric integrins, and the ongoing discoveries, the number of α and β subunits are increasing $\{(\alpha)_n$ and $(\beta)_n\}$ to add to new integrin types.

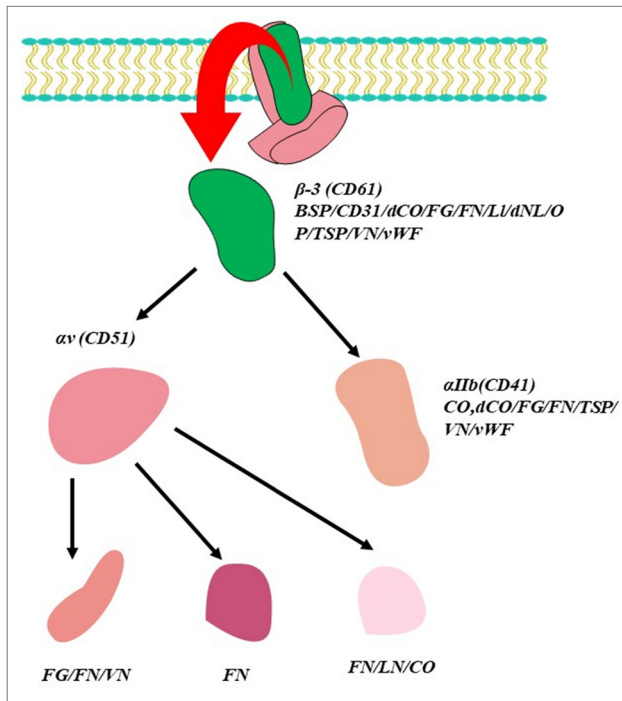


Figure 2: Integrin α - β heterodimers identified and grouped according to the broad families, and the ligands bound by the heterodimer are presented in *italics* either under α subunit, β subunit, or adjacent to the $\alpha\beta$ pair. Ligands BSP, bone sialoprotein; CD31, cluster of differentiation antigen 31; CO, collagens; dCO, denatured collagens; FG, fibrinogen; FN, fibronectin; L1, cell adhesion molecule; LN, laminins; dLN, denatured laminin; OP, osteopontin; TN, tenascin; TSP-1, thrombospondin; VN, vitronectin; vWF, von Willebrand fac. The ligands are listed in alphabetical order and are not presented as a major ligand for any of the receptors.

with various tissues distribution [4,5]. The α and β subunits form the outer cell located in ligand-binding domain, where two multidomain longer chains are termed as the “legs” with the two single-pass membrane helices and the two short cytoplasm embedded tails. These α and β subunits show nonuniformity, they do not express homology to each other, and their conserved regions are common among subtypes of both the subunit groups (Figure 2) [6,7]. Generally, an α subunit specifically associates with a particular β subunit only, e.g., $\alpha 5$ subunit only attaches to $\beta 1$, wherein the β subunits are often more random in their organizations, e.g., the $\beta 2$ subunit attaches with αM , αL , αX , and αD , except the subunit $\alpha 4$, which is associated with the $\beta 1$ and $\beta 7$, and αV binds to $\beta 1$, $\beta 3$, $\beta 5$, $\beta 6$, and $\beta 8$ subunits. Each of these subunits is composed of large extracellularly located amino acid (AA) sequence, together with ~740–780 AAs as part of the β kinds of subunits. There is also a singular domain of the structure located across the membrane consisting of nearly 20 AAs together

with a short structure part located in the cytoplasm having a chain length of 40–50 AAs (Figure 2) [8,9]. However, an exceptionally high chain length of 1,000 AAs is found for the $\beta 4$ subunits [10]. There is a sevenfold repeating unit of about 60 AAs of the α subunits in the *N*-terminal half of the subunit, and these repeating units get folded into a single compact part, wherein they are arranged around a pseudo-symmetry axis to form the structural domain, called β -propeller.

The ligand-binding substructure, the loops in $\alpha 4\beta 1$ were identified by Irie *et al.* [11]. These loops are critical for ligand binding and are available as repeats of 2 and 4 loops on the upper-face side of the β -propeller (Figure 3). For the residues in the $\alpha 5\beta 1$, they are in close proximity with the ligand-binding site of this integrin [12], which was found in close proximity to the propeller near the putative loop. Binding to integrins is achievable through divalent cation dependency. However, the role of the cation needs to be probed in detail. Certain repeats of the α subunit possess multiple cation-binding sites, and these sites were thought to be at the lower surface of the propeller [13]. Around $\frac{1}{5}$ of the integrin α subunits possess an I-domain of nearly 200 AA, homologous to von Willebrand factor (vWF), and are located between the blades 2,3 of the β -propeller [14]. These I-domains mimic the ligand-binding characteristics of the intact integrin, and thus, the I-domain, whose structures are well understood, are involved in ligand binding [15–18]. The crystal structures of these domains have been determined for the αM , αL , and $\alpha 2$ subunits. These I-domains form Rossman dinucleotide-type fold with a central β -sheet, consisting of five parallels and one antiparallel strand encircled by eight α -helices in association with divalent cation that is coordinated through the loops at the top to form metal ion-dependent site for adhesion. The I-domain had no other major structural motifs in the presence of either Mg^{++} or Mn^{++} or in the absence of cations, i.e., Zn^{++} (Figure 3a and b) [19].

Integrin, $\alpha v\beta 3$, possess a cluster of differentiation (CD), the antigen 31 for αV and 61 for $\beta 3$. The X-ray diffraction (XRD) studies devised crystal structure showed αM I-domain in complex formation with a cation that is unusually bound to glutamic acid (Glu) AA residue rather than to the water molecule. The observation has led to speculation that Glu binding is mimicked by the aspartic acid (Asp) AA residue of the RGD (Arg-Gly-Asp) tripeptide, and the aspartic acid residue of another tripeptide LDV (Leu-Asp-Val) of the recognition sequence of the integrin also led to the design and preparation of several drugs and drug candidates as potential $\alpha v\beta 3$ integrin inhibitors [20].

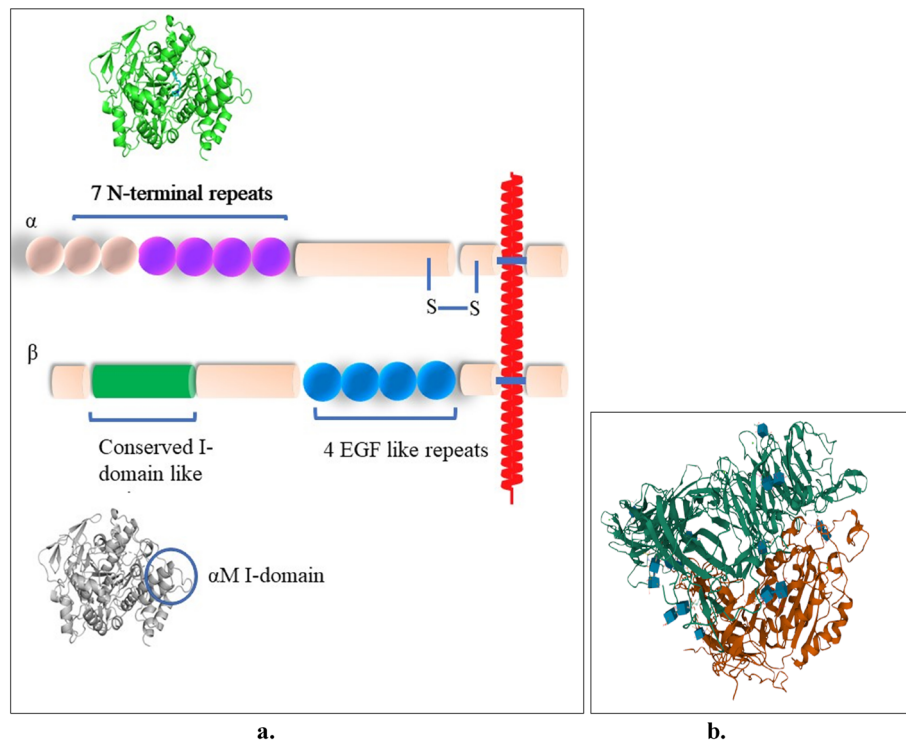


Figure 3: (a) Schematic diagram of an integrin α - β heterodimeric structure. Seven α subunits *N*-terminal repeats with the last four containing EF-hand-like divalent cation-binding sites. The seven repeats fold to form the β propeller part (β -sheets labeled as W1–W7). The putative I-domain-like structure of the integrin β subunit is also shown. The αM I-domains illustrate Rossmann folds as adopted by β I-domains. Down from the putative β I-domain-like motif are located four cysteine-rich epidermal growth factor-like repeats, inspired and with permission from Humphries [7]; (b) the crystal structure of the extracellular segment of the integrin $\alpha\text{v}\beta 3$ with no bound RGD peptide. Source: Protein Data Bank, PDB code 1JV2) [14,21].

A number of ligands of small molecular weights (SMWs) and macromolecular templates of synthetic, recombinant, and biological origins bind to the integrin $\alpha\text{v}\beta 3$ for regulating the cancer cell proliferation and metastasis; however, the cross-talks between the different integrin types and other entities of several types involved in signal transduction pathways affect the modulation of cancer proliferation and cancer metastasis by integrin interference. Also, the carcinoembryonic antigen cell adhesion molecule 6 (CEACAM6) is involved in activating the integrin focal adhesion kinase, a nonreceptor tyrosine kinase type toward stimulating many biological activities, including cancer proliferation and metastasis, and hence is of concern in cancer biology. The signaling inhibition of the $\alpha\text{v}\beta 3$ integrin serves as an important hit point for developing effective therapies for different cancers of several organs and tissues. The role of Tetrac (3,3',5,5'-tetraiodothyroacetic acid), which provides best receptor binding to the integrin receptor-binding domain, categorically subdues the cancer cell proliferation, and integrin works as a coactivator [3]. Several cancer conditions have been implicated to arise from different factors of structural and functional aberrations,

and the $\alpha\text{v}\beta 3$ integrin molecule, their ligands, and their heightened interactions promote several disease states. The hepatic stellate cells upon activation, liver sinusoidal endothelial cells on capillarization and the angiogenic endothelium, profusely express the $\alpha\text{v}\beta 3$ integrin and are involved in liver disorders, including intrahepatic angiogenesis, liver fibrosis, and chronic liver diseases. Rationally designed protein ligands have also been developed to successfully reverse liver fibrosis and reduce the activated and capillarized liver cells [22]. Recently, a new class of integrin antibodies has been discovered for fibrosis [23]. The $\alpha\text{v}\beta 3$ integrin ligand is also known to regulate the quantitative increase in smooth muscles' hyperplasia to restrict Crohn's disease [24]. The severely upregulated $\alpha\text{v}\beta 3$ integrins in lung fibrosis, where the SSTR2 (somatostatin receptors subtype 2) helped to confirm the fibrosis after 2 weeks of induction, were used for imaging purposes [25]. The cancers of breasts, lungs, and prostate tend to metastasize in the bone and the bone marrow, owing to modulation in $\alpha\text{v}\beta 3$ integrins' expressions, and the processes of involved signaling, wherein the $\alpha\text{v}\beta 3$ integrin was overexpressed and followed downstream signaling

pathways to induce osteolysis, have been observed [26]. Integrin dysregulation is the key in tumor-induced bone destruction through metastatic participation of cancers, especially solid tumors [26]. Also, the $\alpha\beta3$ integrin dysregulation together with the CD47 (cluster of differentiation 47, immunoglobulin, cell surface located, integrin associated) signaling is reported to promote inflammation of the joints, breakdown of cartilage, and osteoarthritic progression [27]. The $\alpha\beta3$ and $\alpha\beta5$ integrin receptor involvements in animal models of atherosclerosis and modulation of macrophagic functions through omentin-1 to reverse the plaque vulnerability have been reported recently [28]. Moreover, the $\alpha\beta3$ integrin in podocyte-mediated kidney disease of chronic nature together with suPAR (soluble urokinase plasminogen activator receptor) and APO1 (apolipoprotein L1) risk variants as part of a tripartite complex has also been reported [29]. An important understanding related to the structural motif changes in the integrin molecules led to conclude that the aberrant glycosylation of $\alpha\beta3$ integrin is among one of the causes of progression of melanoma, which has been confirmed through the WM1205Lu cell lines, the highly metastatic variant of the WM793 primary melanoma cell line [30]. Cytomegalovirus, an opportunistic pathogen causing birth defects in neonatal and that altered physiological conditions in immuno-compromised individuals, has a co-receptor in $\alpha\beta3$, together with epidermal growth factor receptor [31]. Moreover, all Parkinson-diseased (PD) and related syndromes showed higher $\alpha\beta3$ levels in *locus ceruleus* and *substantia nigra pars compacta* in live subjects having incidental Lewy body disease with confirmed Lewy bodies, but the PD and progressive supranuclear palsy conditions exhibited higher $\alpha\beta3$ levels in post-mortem brain tissues [32]. The $\alpha\beta3$ integrin has also been proposed as a drug target for rheumatic disorders and rheumatoid arthritis [33]. Activation of $\alpha\beta3$ integrin-based signaling promotes fibrotic changes in glaucoma and glucocorticoid-induced glaucoma [34]. The inflamed muscles, especially in hypoxia and ischemia, also promote higher expressions of $\alpha\beta3$, an angiogenic factor [35]. The prominent role of $\alpha\beta3$ integrin in tumor angiogenesis has led to strategies to counter them for further angiogenesis, tumor growth inhibition, and metastasis of cancer [36].

The thyroid-regulated disorders and involvement of the $\alpha\beta3$ integrin are well known [37–42]. The angiogenic signaling in multipotent stem cells through the $\alpha\beta3$ integrin is also influenced by the thyroid hormones. The proven role of Tetrac, an ingredient of thyroid hormones, in the antiangiogenic role in the tumor microenvironment has also been established [43]. The thyroid hormonal role in chronic lymphocytic leukemia [44] is

well known. The role of thyroid hormone–integrin $\alpha\beta3$ and the therapeutic strategies for colorectal cancer has been reported recently [37]. The crosstalk between the $\alpha\beta3$ integrin and the estrogen receptor is implicated in the proliferation of ovarian cancer [38] and human lung carcinoma [45].

Thyroid hormones, *e.g.*, T3 (3,5,3'-triiodo-L-thyronine) and T4 (L-thyroxine), produce nongenomic effects and are well known to control several cellular and subcellular functions, thereby affecting multiple organs. The effects are primarily maintained through $\alpha\beta3$ integrin together with other receptors, not to mention the TR α and TR β . The cancer cells, capable of reprogramming their metabolism, adopt aerobic glycolysis instead of oxidative phosphorylation, thereby dysregulating the PKM2 (pyruvate kinase isoform M2, cellular energetics mediator), the rate-limiting enzyme of glycolysis, and due to several biochemical processes and signal transduction activities, the cancer cells produce more reactive oxygen species (ROS) to survive and propagate. Thyroid hormones are also associated with oxidative injury in hyperthyroidism where increased production of ROS and reactive nitrogen species is observed. The inhibition of $\alpha\beta3$ signaling, through ligand binding, has a prominent role in mediating the dysregulation and finally the cancer progression by stopping the increased angiogenesis. The involvement of thyroid constituents, T3 and T4, in liver cancer has been described [46]. The implications of the thyroid hormones and the integrin over expression are well debated and have led to the focus primarily on the development of novel drugs, of which both traditional and contemporary nano-architectural originating drugs for anti-integrin therapies in conjunction with the thyroid hormones are designed and produced. The thyroid hormones, T3 and T4, including other thyroid hormone agonist analogs [47] are pro-angiogenic in nature [48]. The pro-angiogenic activity removal of thyroid hormone at cell surface receptor is thought to regress the tumor xenografts by ~50% [49–51] and hence can shrink the tumor significantly. As the thyroid hormones are the key regulators of several cellular processes of cell proliferation, differentiation, apoptosis, and metabolism, their association with the cancer mass has been suggested very earlier [52]. The clinical observations have supported the notion that hormonal deficiency and hypothyroidism inhibit tumor growth [24,53,54]. These actions are facilitated through several nongenomic pathways, which include activation of integrin $\alpha\beta3$. The $\alpha\beta3$ integrin, containing binding sites, S1 and S2 [55], binds to the thyroid components, T3 and T4, with a lower affinity for T4 than T3. The physiologically present T3 specifically binds to S1 and activates the phosphoinositide 3-kinase pathway, while the T4 binds at the S2 receptor site and

activates the MAPK3 and MAPK1, or ERK1 and ERK2 (44 and 42 kDa Ser/Thr kinases) pathways involving the Ras-Raf-MEK-ERK cascade of signal transduction, which participates in the regulation of adhesion, progression, migration, survival, differentiation, proliferation, transcription, and metabolism [56]. Thus, the $\alpha\text{v}\beta 3$ integrin enables the proliferative action of thyroid hormones on cancer cells and participates in angiogenesis involving the blood vessel cells [35]. In the nutshell, the dysregulation of physiologically bioavailable thyroid hormones affects cancer development and progression and escalates the risk of solid tumors of different organs and tissues through higher availability of T3, together with the T4 thyroid hormones, thereby increasing the integrin binding, which has been mainly supported by clinical and subclinical studies [57]. The reduction in the biological presence of thyroid hormones or lesser bioavailability, their inhibition, and their restricted and reduced binding to the integrin receptor through antagonists have led to newer anticancer agents. In this context, the unraveling of the 3D structure of the $\alpha\text{v}\beta 3$ integrin, its regulation of the ligand binding, and binding affinity manipulated through antagonistic molecules of low and high molecular weight with the capability to stabilize certain specific conformations of the integrin receptor and the signaling to the cell; the molecules that may antagonistically inhibit certain intercellular adhesion functions have been proposed as the rationale for integrin-based drug design. The heterodimeric structure, being capable of recognizing several different types of structural motifs, together with regulating the binding affinity, directs the conformational changes at the receptor site and establish communication between the extracellular and intracellular domains of the cells. Thus, the ligand binding influences the allosteric changes in the integrin conformation, and the integrin's information feeds up about the external cellular environment to the cell. Therefore, the integrins that serve as sensors of the extracellular matrix, and their surroundings at the molecular level, together with working as an effector system for certain cytoskeletal forces, including the blood vessels, effectively make the sensor-effector system based on its heterodimeric structure and its amino acid sequence employed in different functions of the cells [58]. Moreover, as the thyroid hormones support the proliferation of cancer cells [59–62], any change in the tumor size suggested several plausible mechanisms with the hormonal activity being reduced at the tumor site [57]. Importantly, the pro-angiogenic activity of T3 and T4 hormones originate at the surface of the cellular receptor, integrin $\alpha\text{v}\beta 3$, partly found extracellularly on the cell membrane whose mechanism has been understood on the chick chorioallantoic membrane (CAM) model-based assays [62], and the integrins have been shown

to express multiple functions that connect to the ECM proteins [63], cell surface, and within the cells located growth factor receptors [64], as well as in the specific gene transcriptions [65]. Also, the iodothyronine receptors on the integrin have been found to regulate the cancer-relevant angiogenesis and are considered as the key for cancer cells' survival through the involved pathways. The iodothyronine receptor can be covered with certain drugs and their nanoscale covalent conjugate [66,67] to provide antitumor effects through regressed or terminated angiogenesis.

3 Anti- $\alpha\text{v}\beta 3$ integrin-based drugs for cancer chemotherapy

The tumor cells repenetrate the vessels or walls and continue to multiply forming another clinically detectable tumor over a period, which is characterized as metastatic tumors [68]. Alterations in the adhesive nature of the tumor cells can bring significant changes in the metastasis of these cancers. Thus, limiting the role of the molecules involved in various pathological activities, such as tumor's neo-vascularization and metastasis, can bring significant therapeutic effects. An encouraging target for cancer treatment, integrin $\alpha\text{v}\beta 3$, is capable of binding to several ECM components, *i.e.*, fibrinogen, fibronectin, and so on to provide the desired effects on cell proliferation, cell adhesion, and control on the further development of the cells colony. The $\alpha\text{v}\beta 3$ is expressed on the malignant tumor cells that have made them an attractive target for developing anti- $\alpha\text{v}\beta 3$ antagonists' molecular templates and biologically active new chemical entities. The antagonists of $\alpha\text{v}\beta 3$ receptor protein by inducing the apoptosis of the new blood vessel can block tumor-associated angiogenesis and thereby leaves the malignant cells dysfunctional. Figure 4 shows selected $\alpha\text{v}\beta 3$ antagonists developed by various groups and pharmaceutical concerns.

On the molecular level, the thyroid hormone analog molecules, T3 and T4, also termed thyrointegrins, work and selectively activate the extracellularly available integrin receptors. The recognition site, Arg-Gly-Asp (RGD), is in close approach within the receptor [69], and the receptors, thyroid-hormone, and $\alpha\text{v}\beta 3$, structurally and functionally are neither analogous nor related to each other, but triggering of the thyrointegrin receptors located on the cell surface results in nuclear mediation to elicit pro-angiogenic activity. The radio-ligand binding studies have confirmed the preferences of the purified $\alpha\text{v}\beta 3$ receptor, which is higher to T4 than T3 [70]. Sources also demonstrated that the activation and nuclear translocation of mitogen-activated

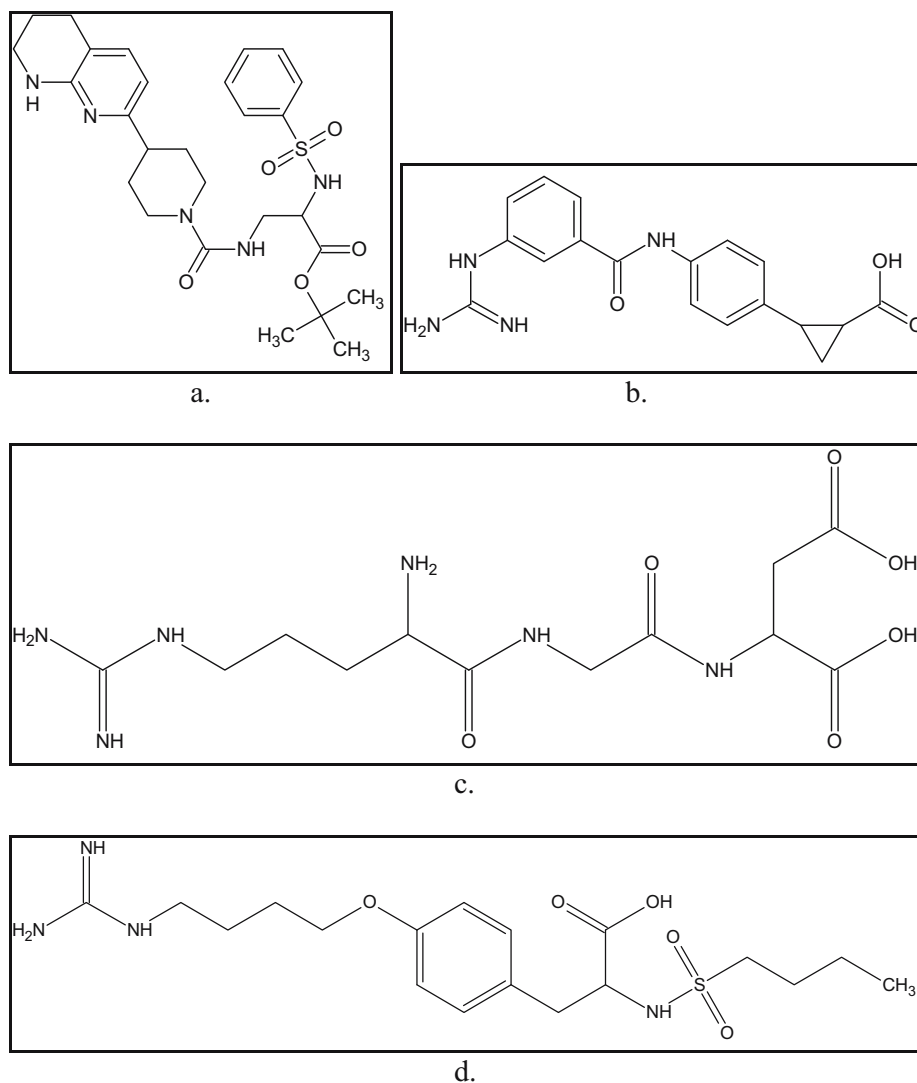


Figure 4: Major $\alpha v \beta 3$ antagonists; (a) Merck & Co, patent WO 9818461-A1: 4-(δ -6,7,8-tetrahydro-[1,8]naphthyridin-2-yl)piperidin-1-yl-carbonyl-2-(*S*)-phenylsulfonylamino- β -alanine-*t*-butylester; (b) searle, patent WO 9736858-A1: 2-[3-[[[3-[(amino-imino methyl)-amino] phenyl] carbonyl]-amino] methyl] phenyl]-cyclopropane carboxylic acid (trifluoro acetate salt); (c) Merck KGaA, patent DE 19548709-A: 2(*S*)-2-[[2-[[[(2*S*)-2-amino-5-(diamino methylidene amino)]amino]acetyl]amino] butanedioic acid; (d) (*S*)-2-butylsulfonamido-3-[4-(3-amino-propoxy) phenyl]-propionic acid.

protein kinases (MAPKs) and hormone-induced angiogenesis is comparatively higher because of the thyroid hormone T4 compared to the T3. The role of thyrointegrin is imperative in these senses. The binding of the T4 to the integrin receptor is inhibited by the integrin $\alpha v \beta 3$ antagonists, which thereby prevents the activation of MAPK signals, which were conclusively based on the spotting of the binding site for iodothyronine on the integrin receptor, and well relates to the functional aspects of the MAPK signal cascade activation by the thyroid hormone. The thyroid hormone derivative, 3-iodothyronamine, is also reported to conjugate with a trace amine receptor (TAR-I), but operationally, the entity is opposite to thyroid hormones, T3 and T4 [71]. The binding

domains of the integrin $\alpha v \beta 3$ constitute two binding domains for the thyroid hormone [72]. Domain 1 is an RGD tripeptide-sensitive site, and it has no relation to cellular proliferation. However, the RGD-recognition site, which contributes to the interaction of different proteins, connects the integrin systemically and mechanistically with the ECM proteins. However, domain 2, is an insensitive RGD tripeptide site and contributes to cellular proliferation as well as angiogenesis, thereby making both the sites structurally distinct and functionally altogether different. The thyroidal receptor and the RGD tripeptide recognition sites are nearly overlapping in their situational domains. The site of the thyroid hormone also functions in a unique way in the process of gene

expression during the tumor cell proliferation and contributes its role effectively [73,74]. Moreover, these receptors (thyroid hormone receptors) present on the cell membrane are G protein sensitive [75].

The Tetrac blocked the effects of T4 and T3. Thyroid hormone analogs, when applied locally, promoted the desirable neovascularization, *e.g.*, in healing the wounds or to prevent the undesirable angiogenesis that supports the growth of tumors and can be inhibited by using Tetrac. The polymer conjugates and nanoparticle formulations of thyroid hormones and their analogs were also prepared to test the inhibition of angiogenesis [76]. Other anti-angiogenic agents and their derivatives with the polymeric entities, liposomes, nano, and microstructured materials have been prepared and studied. mAb LM609 Tetrac, Triac, and XT 199 are a few illustrations of such anti-angiogenic thyroid hormone antagonists (Figure 5). The nanoparticles were attached through a hydroxyl group. Among the well-known thyroid agonists, the diiodo thyro propionic acid and the GC-I, GC-1 agonist's linking was also achieved. The ether bond, as found in T3, T4, Triac, and Tetrac, does not seem to be essential [77] for the activity exhibition. However, the intervening carbon is required rather than the ethereal bond. The 3' iodine of the outer ring is also not necessary according to the structure–activity relationship (SAR) requirements. Similarly, position 3' need not be modified [78] for any elicitation of the biological activity. Through the amide bond embedded in the nanoparticles, the linker was linked to the nanoparticle and thus contributed to the development of novel nano-formulations containing $\alpha\beta 3$ antagonists. The synthesis of guanidine, urea, methylamine, and propylamine derivatives of the Tetrac has also shown anti-angiogenic bioactivity in the CAM model from 65% to 73% inhibitions at the dose of 0.25–2.0 $\mu\text{g/mL}$ [79].

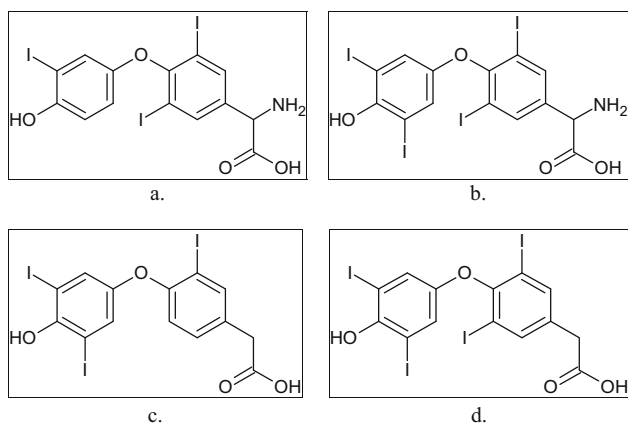


Figure 5: Thyroid hormone compounds, its analogs, and thyrointegrin antagonist; (a) T3, (b) T4, (c) Triac, and (d) Tetrac.

4 Contemporary nano-architected drugs

Nanoparticulate formulations of thyroid hormones and analogs were synthesized with additional polymeric conjugations. To locally deliver the thyroid hormone and its analogs, the nanoparticles delivery modalities together with polymeric conjugates were used as a carrier and delivery/transportation matrices. The chronologically defined delivery at the targeted tissue and cell sites with the nano-sized entities was achieved, wherein the prepared nano-enabled antagonist drugs were between ~150 and 250 nm in size [80]. The nanoparticles or the thyroid analog-conjugated polymers can also target cancers in various body sites, including the skin. In addition, the thyroid hormone analogs and their antagonist can also be employed for hematopoietic, as well as stem cell-related malfunctions and disorders.

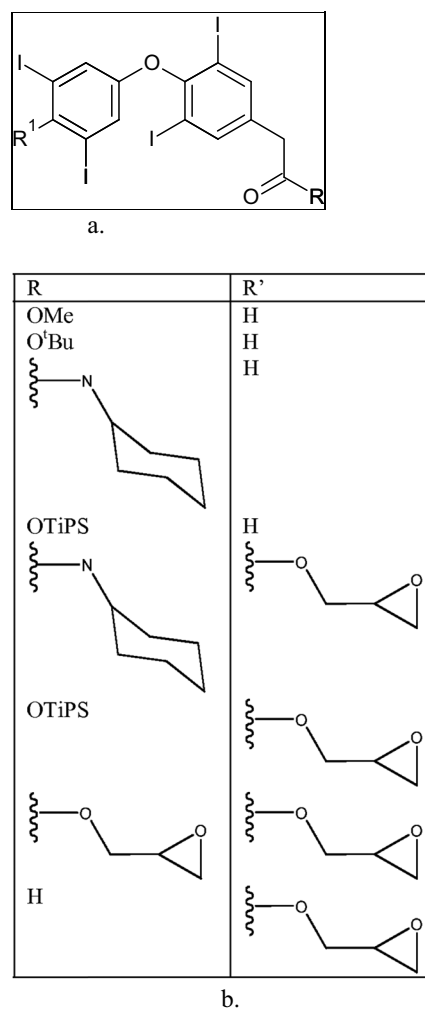


Figure 6: Tetrac and related products: (a) Tetrac ($R^1 = R = \text{OH}$) and (b) various semisynthetic products.

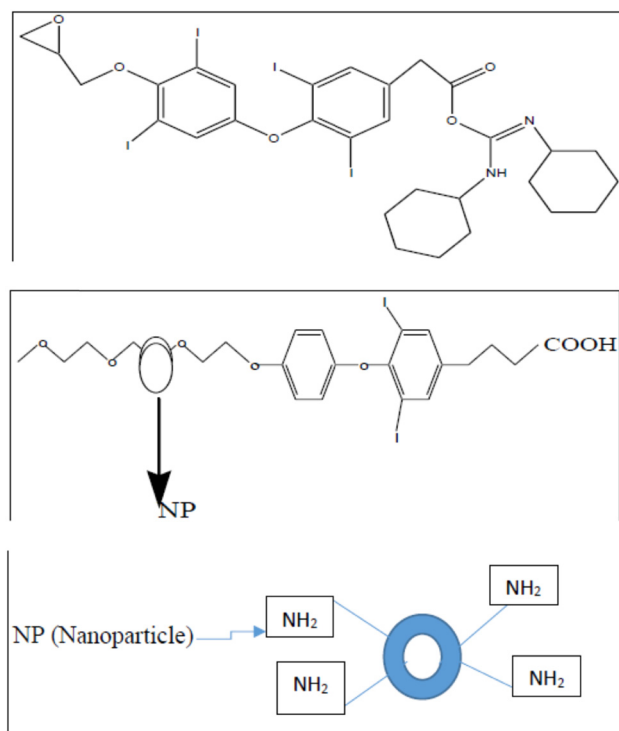


Figure 7: Nano-Tetrac drug model.

These formulations have been recommended for delivery for faster cell reproduction at the time of bone marrow transplant. The semisynthetic precursors of Tetrac (Figure 6a, and b) were also researched to find and institute the best product for angiogenesis [81,82].

In addition, with the help of various RNA microarray techniques, the genomic activities within thyroid hormones, specifically at $\alpha\beta 3$ receptors, were studied in

various cancer cell lines obtained from human samples [37,83–86]. Previous studies have also established the anticancer ability of the Tetrac, which antagonizes the proliferation of cancer cells. The thyroid hormone is anti-apoptotic [87], whereas the Tetrac has pro-apoptotic characteristics [39]. The Tetrac analog-treated tumors have been investigated in detail for apoptosis-related and differently regulated gene contributions. The unchanged moieties of Tetrac and Triac molecules were covalently conjugated with nanoparticles that inhibited entrance into the cells and effectively destructed the MDA-MB-231 cells (estrogen receptor-negative human breast cancer cells). The genes, *BCL2L14* and *CASP2*, were also accelerated, but the Tetrac formulation diminished the expressions of anti-apoptotic *MCL1* and *XIAP* genes [88,89].

Angiogenesis in cancers involves downregulation of thrombospondin-1 (TSP-1), which is an endogenous inhibitor of angiogenesis. The transformation of genetic lesions through the expression of mutant RAS oncogene, which is intrinsic to the cancer cells, induces the TSP-1 downregulation. However, the effect of the mutant RAS gene on tumor neo-vascularization is confined not only to angiogenic modulation in cancer cells but also to the RAS with different signaling mechanisms and also to elicit proangiogenic effects [90]. The Rab18 and Rab1B were among various RAS oncogenes, which were considered to be highly affected by the nanoparticles of Tetrac [90]. The Rab18 was downregulated, whereas Rab1B was upregulated. The Rab1B expression is considered responsible for the differentiation of malignant cells, *e.g.*, the monocytes in promyelocyte leukemia [91]. The agonist thyroid hormone-supported proangiogenic activity is blocked by Tetrac, an anti-angiogenic product. The microarray results

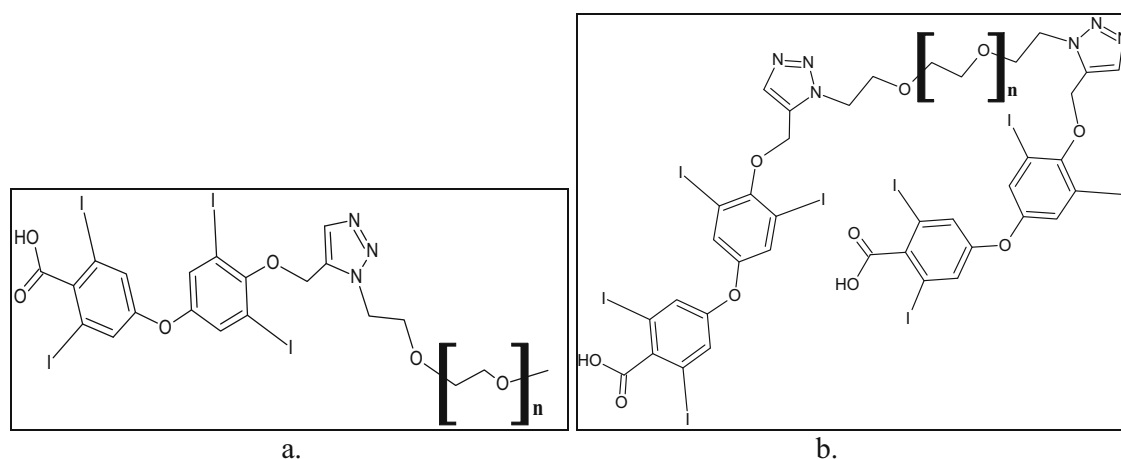
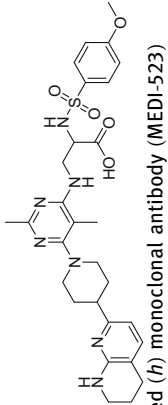
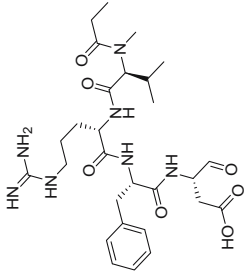
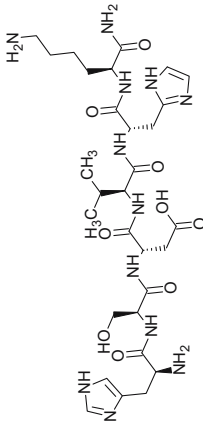


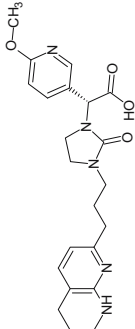
Figure 8: Chemical structures of PEG (polyethylene glycol) conjugated Tetrac: (a) PEG-conjugated one Tetrac unit conjoined through single triazole unit (PEG-triazole-Tetrac, P-mono-TAT); (b) two Tetrac units conjoined through distantly placed two triazole structures (Tetrac-triazole-PEG-triazole-Tetrac [P-bi-TAT]); n means the average number of repeating oxy ethylene units (90) of the polymer PEG, PEG A_v MW 4,000 amu.

Table 1: Drug candidates based on integrin antagonists under development

Category	Compound/code	Target and structure	Application/description	Ref.
Small molecule	GLPG 0187	Targets αvβ1, αvβ3, αvβ5, αvβ6, αvβ8, and α5β1 	Selective 1.3 nM IC ₅₀ for αvβ1, RGD binding, clinical trial for liver cancer	[108]
Monoclonal antibodies	<i>h</i> -Vitaxin (<i>h</i> humanized)	αvβ3, humanized (<i>h</i>) monoclonal antibody (MEDI-523)	Clinical trials for metastatic melanoma and prostate cancers, phase I and II trials	[109]
	Etaracizumab	αvβ3, humanized monoclonal antibody (MEDI-522)	Clinical trials for melanoma, prostate, and ovarian cancers, phase II trials	[110]
	Abituzumab (ITGAV)	αvβ1, αvβ3, αvβ5, αvβ6, αvβ8, humanized IgG2 monoclonal antibody	Clinical trials for colorectal, multiple sclerosis, interstitial lung, and prostate cancers, phase I and II trials	[111]
Peptidic	Cilengitide (EMD 121974)	Selective for αvβ3, αvβ5 	Clinical trials for multiple cancers, phase I, II, and III trials	[112]
	HSDVHK-NH ₂	αvβ3, antagonist of integrin αvβ3-vitronectin interaction 	Site-specific RGD recognition, antagonist against αvβ3-GRGDSP, integrin αvβ3-vitronectin interaction, with an IC ₅₀ of 1.74 pg/mL	[113,114]
	Echistatin and its TFA (trifluoro acetic acid) salt	αvβ3, αvβ5, and α5β1, smallest active RGD protein, disintegrin type – derived from viper snake venom; sequence structure with disulfide bridges at Cys ² -Cys ¹¹ ; Cys ⁷ -Cys ³² ; Cys ⁸ -Cys ³⁷ ; Cys ²⁰ -Cys ³⁹ :AA (amino acid) sequence: ECESGPCCRNCKFLKEGTICKRARGDDMDYCNCKTKDCPRNPHKGPAT	An integrin antagonist developed to inhibit osteoclastic bone resorption	[115]

(Continued)

Table 1: Continued

Category	Compound/code	Target and structure	Application/description	Ref.
Nonpeptide	MK-0429 (L-000845704)	Selective antagonist, orally active, targets $\alpha\beta 1$, $\alpha\beta 3$, $\alpha\beta 5$, $\alpha\beta 6$, $\alpha\beta 8$, and $\alpha 5\beta 1$ 	Pan-integrin antagonist	[116]

have proposed another mechanism for Tetrac, wherein it is supposed to inhibit angiogenesis through upregulation of *THBSI* gene expression [92]. The prohibition of tumor cells growth is supported by the pure, un-modified Tetrac and its different nano-preparations (Figure 7) through upregulation of *THBSI* expression. The expressions of *CBYI*, *XIAP*, and *THBSI* genes are affected by Tetrac nanoparticles. In addition, the nanoparticle formulations of Tetrac helped in showing MDA-MB-231 cell gene expressions, which are part of the cell survival mechanism [89]. This suggested that the pure, unchanged Tetrac and its nanoformulations can fit in the integrin-binding site, which is slightly different. The thyroid hormone receptor domain contains closely related receptor sites, and either of these two acts through specific transduction. Therefore, the fact of distinction between nanoparticulate and unmodified Tetrac by receptor domains is not surprising. The other possibility is that the structurally unchanged Tetrac might have triggered actions that offset to initiate the integrin $\alpha\beta 3$ receptor site activation and binding. Interestingly, in the absence of T3 and T4, the coherent behaviors of the antibasic fibroblast growth factor and the anti-vascular endothelial growth factor, with Tetrac involvement, have been found in the endothelial cells [93,94]. Therefore, the nanoparticle-loaded Tetrac's action at triple-negative breast cancer, the most malignant tumor, is not surprising.

These novels Tetrac nanodrugs also inhibit the human cancers' major oncogenic Wnt signaling pathway. It also happens to be conjoined with downregulation of *CTNNA1* and *CTNNA2* expressions, the catenin genes [95], along with the concurrent up-regulation of the *CBYI* gene, the nuclear antagonist for the catenin activity. Also, there are 13 differentially regulated RAS oncogenes reported [96], of which 8 are downregulated after the Tetrac and its nano-sized formulated drugs through interfering with the oncogenic-signaling pathways, and this is a clear indication of the signaling pathways and the integrin involvement in anticancer activities. The cross-talks add to the complex signaling and can lead to cells' unpredictable fate and therapeutic outcomes [97].

Among other products mimicking the Tetrac in completely binding to the $\alpha\beta 3$ integrin, and with the thyroid hormone receptor, to restrict cancer proliferation, the analog of (*E*)-stilbene, resveratrol, is worth mentioning. Nanotechnical advances in preparing the RGD (Arg-Gly-Asp) tripeptide conjugate with gadolinium-molybdenum dioxide (RGD-Gd-MoO₂) for magnetic resonance imaging (MRI) and cancer therapy, as an ideal theranostic agent, the derivative of Tetrac, the 150–200 nm sized PLGA-encapsulated N-DAT (Nano-Diamino-Tetrac-PLGA), and nano-resveratrol (N-RES) for targeting the integrin $\alpha\beta 3$

for cancer controls are reported [98]. The diamino-Tetrac-PLGA conjugates (DAT-PLGA) and the PLGA-encapsulated nano-DAT (N-DAT-PLGA, or N-DAT) effects on different cancer cell lines, *i.e.*, pancreatic, breast, lungs (non-small cell), colorectal, hepatocellular, glioblastoma, bladder, and gastric, namely, SUIT-2, MPanc96, MDA-MB-231, MCF-7, H1299, HepG2, U87, 253JBV, and AGS in the *in vitro* and *in vivo* xenograft models, were of downregulating nature for the $\alpha\beta 3$ expressions. Reduction and viability were exhibited in N-DAT-treated glioblastoma xenografts, and N-DAT was found to be safe at higher doses [99]. The N-DAT and Tetrac together are known to induce antiproliferation activity through $\alpha\beta 3$ intermediacy in different K-RAS (K-RAS-mutant HCT116 cells and K-RAS-wild type HT-29 cells) colorectal cancer [100]. In addition, the cyclic RGD-based pentapeptide derivative {*cyclo*-(Arg-Gly-Asp-D-Phe-Lys)}, the c-(RGDfK)₂, conjugates of poly-L-glutamic acid (PGA) and polyethylene glycol (PEG) with an anticancer agent, paclitaxel (PTX), as PGA-PTX-E-[c(RGDfK)2] and PTX-PEG-E-[c(RGDfK)2] conjugates exhibited boosted anticancer activity against MDA-MB-231 tumor cells [101]. The PLGA-PEG-NPs {poly-(D,L-lactic-co-glycolic acid)-*block*-polyethylene glycol nanoparticles} in conjunction with the c-(RGDfK) motifs were utilized to nano-formulate another anticancer agent, cisplatin, to target integrin $\alpha\beta 3$ at RGD-binding domain in cancer cells [102]. The interesting observation of shape effects of the constructs, including nano, micellar, cyclic, and linear formulation products, was remarkable and noteworthy. The solubilization capacity, and the stability in kinetic terms of lower energy status, provided the cyclic RGD derivatives improved targeting, while the linear RGD constructs were less pronounced in their targeting efficiency [103]. The nanoparticulate shape provided better-enhanced cytotoxicity in comparison to the non-nano formulations for the cisplatin in prostate and breast cancer cell lines [104]. The nanoparticle-shaped molecular entities also exhibited selective efficiency and potent activation through enhancements in their permeability retentions [105].

Recently (12/2021), PEG-conjugated two Tetrac units (Tetrac-triazole-PEG-triazole-Tetrac) conjoined through distantly placed two triazole entities, together with PEG-conjugated one Tetrac unit conjoined through a single triazole unit (PEG-triazole-Tetrac) (Figure 8) [106], were reported for their pharmacokinetics and biodistribution studies in animal models' serum through utilizing a concurrently developed bioanalytical method for the purpose. The product, P-bi-TAT, was found to be involved in down-regulating several signaling pathways, including the NF- κ B, and was suggested for acute myeloid leukemia and other malignancies owing to their high thyrointegrin $\alpha\beta 3$ affinity [107].

A list (Table 1) of drug candidates' structural classes types under evaluations at different phases of clinical trials (phase I–III) also discusses the applications and integrin receptor specificity along with their molecular structures developed as integrins, especially $\alpha\beta 3$ inhibitors.

5 Summary and future prospects

Significant knowledge about the role of integrins in various pathological and physiological conditions led to the invention of various therapeutic agents. The nano-sized formulated $\alpha\beta 3$ antagonist drugs certainly play an important role in several biological activities in therapeutically relevant pathways, which needs further and deeper studies, to pinpoint $\alpha\beta 3$ inhibition targets. The Tetrac model is an initiating point in developing $\alpha\beta 3$ antagonist drugs as nano modalities that might form part of the combination therapy as an anticancer agent. Opportunities for other cross-reactive and closely related thyrointegrin antagonists may also be searched to lead the design of newer and specifically targeted new chemical entities at specific locations in the receptor. Efforts to design and develop new chemical entities fitting the receptor binding site, and pharmacophore-based model products exists which could be transformed in future into nanostructural motifs, and conjugates of a plethora of biodegradable and biocompatible polymers of natural, synthetic, and semi-synthetic origins.

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References

- [1] Mangale SS, Modi DN, Reddy KVR. Identification of genes regulated by an interaction between $\alpha\beta 3$ integrin and vitronectin in murine decidua. *Rep Fer Dev.* 2008;20:311–19.
- [2] Desgrosellier JS, Cheresh DA. Integrins in cancer: biological implications and therapeutic opportunities. *Cancer: Nature Reviews.* 2010;10:9–22.

- [3] Ley K, Rivera-Nieves J, Sandborn WJ, Shattil S. Integrin-based therapeutics: biological basis, clinical use and new drugs. *Nat Rev Drug Discov.* 2016;15(3):173–83.
- [4] Alberts B, Johnson A, Lewis J, Morgan D, Raff M, Roberts K, et al. *Molecular biology of the cell.* 4th edition. New York, USA: Garland Science; 2002. Available from: <https://www.ncbi.nlm.nih.gov/books/NBK21054/>.
- [5] Takada Y, Ye X, Simon S. The integrins. *Genome Biol.* 2007;8:215.
- [6] Ingber DE. Mechano-sensation through integrins: cells act locally but think globally. *Proc Natl Acad Sci USA.* 2003;100:1472–74.
- [7] Humphries MJ. Integrin structure. *Biochem Soc Trans.* 2000;28:311–40.
- [8] Humphries MJ, Travis MA, Clark K, Mould AP. Mechanisms of integration of cells and extracellular matrices by integrins. *Biochem Soc Trans.* 2004;32:822–25.
- [9] Ridley AJ. Pulling back to move forward. *Cell.* 2004;116:357–58.
- [10] Suzuki S, Naitoh Y. Amino acid sequence of a novel integrin beta-4 subunit and primary expression of the messenger RNA in epithelial cells. *EMBO J.* 1990;9:757–63.
- [11] Irie A, Kamata T, Takada Y. Multiple loop structures critical for ligand binding of the integrin alpha 4 subunit in the upper face of the beta-propeller mode 1. *Proc Natl Acad Sci USA.* 1997;94:7198–3.
- [12] Mould AP, Burrows L, Humphries MJ. Identification of amino acid residues that form part of the ligand-binding pocket of integrin alpha-5beta-1. *J Biol Chem.* 1998;273:25664–72.
- [13] Springer TA. Folding of the N-terminal, ligand-binding region of integrin alpha-subunits into a beta-propeller domain. *Proc Natl Acad Sci USA.* 1997;94:65–72.
- [14] Bienkowska J, Cruz M, Atiemo A, Handin R, Liddington R. The von Willebrand factor A3 domain does not contain a metal ion-dependent adhesion site motif. *J Biol Chem.* 1997;272:25162–67.
- [15] Emsley J, Knight CG, Farndale RW, Barnes MJ, Liddington RC. Structural basis of collagen recognition by integrin alpha2-beta1. *Cell.* 2000;101:47–56.
- [16] Shimaoka M, Xiao T, Liu JH, Yang Y, Dong Y. Structures of the alpha-L I domain and its complex with ICAM-1 reveal a shape-shifting pathway for integrin regulation. *Cell.* 2003;112:99–111.
- [17] Xiong J-P, Stehle T, Diefenbach B, Zhang R, Dunker R, Scott DL, et al. Crystal structure of the extracellular segment of integrin $\alpha\beta 3$. *Science.* 2001;294:339–45.
- [18] Calderwood DA, Tuckwell DS, Eble J, Kuhn K, Humphries MJ. The integrin 1 A-domain is a ligand-binding site for collagen and laminin. *J Biol Chem.* 1997;272:12311–17.
- [19] Dennis KP, Ke RM, Lo PC. Synthesis and in vitro activities of integrin targeting cRGD conjugated Zinc II phthalocyanine. *Chem Asian J.* 2014;92:554–61.
- [20] Weitz-Schmidt G, Welzenbach K, Dawson J, Kallen J. Improved lymphocyte function-associated antigen-1 (LFA-1) inhibition by statin derivatives: molecular basis determined by x-ray analysis and monitoring of LFA-1 conformational changes in vitro and ex vivo. *J Biol Chem.* 2004;279:46764–71.
- [21] Huhtala M, Heino J, Casciari D, de Luise A, Johnson MS. Integrin evolution: insights from ascidian and teleost fish genomes. *Matrix Biol.* 2005;24:83–95.
- [22] Turaga RC, Satyanarayana G, Sharma M, Yang JJ, Wang S, Liu C, et al. Targeting integrin $\alpha\beta 3$ by a rationally designed protein for chronic liver disease treatment. *Comm Biol.* 2021;4(1):1087.
- [23] Zhang J, Wang T, Saigal A. Discovery of a new class of integrin antibodies for fibrosis. *Sci Rep.* 2021;11:2118.
- [24] Flynn RS, Murthy KS, Grider JR, Kellum JM, Kuemmerle JF. Endogenous IGF-I and alphaVbeta3 integrin ligands regulate increased smooth muscle hyperplasia in stricturing Crohn's disease. *Gastroenterol.* 2010;38(1):285–93.
- [25] Schniering J, Benešová M, Brunner M, Haller S, Cohrs S, Frauenfelder T, et al. Visualisation of interstitial lung disease by molecular imaging of integrin $\alpha\beta 3$ and somatostatin receptor 2. *Ann Rheum Dis.* 2019;78(2):218–27.
- [26] Kwakwa KA, Sterling JA. Integrin $\alpha\beta 3$ Signaling in Tumor-Induced Bone Disease. *Cancers.* 2017;9(7):84.
- [27] Wang Q, Onuma K, Liu C, Wong H, Bloom MS, Elliott EE, et al. Dysregulated integrin and CD47 signaling promote joint inflammation, cartilage breakdown, and progression of osteoarthritis. *JCI Insight.* 2019;4(18):e128616.
- [28] Xuze L, Yan S, Shiwei Y, Mengyue Y, Liu P, Jie Y, et al. Omentin-1 modulates macrophage function via integrin receptors $\alpha\beta 3$ and $\alpha\beta 5$ and reverses plaque vulnerability in animal models of atherosclerosis. *Front Cardiovas Med.* 2021;8:1470.
- [29] Hayek S, Koh K, Grams M. A tripartite complex of suPAR, APOL1 risk variants and $\alpha\beta 3$ integrin on podocytes mediates chronic kidney disease. *Nat Med.* 2017;23:945–53.
- [30] Pocheč E, Bubka M, Rydlewska M, Janik M, Pokrywka M, Lityńska A. Aberrant glycosylation of $\alpha\beta 3$ integrin is associated with melanoma progression. *Anticancer Res.* 2015;35(4):2093–103.
- [31] Wang X, Huang DY, Huang SM, Huang ES. Integrin $\alpha\beta 3$ is a co-receptor for human cytomegalovirus. *Nat Med.* 2005;11:515–21.
- [32] Desai Bradaric B, Patel A, Schneider JA, Carvey PM, Hendey B. Evidence for angiogenesis in Parkinson's disease, incidental Lewy body disease, and progressive supranuclear palsy. *J Neural Transm.* 2012;119:59–71.
- [33] Wilder RL. Integrin $\alpha\beta 3$ as a target for treatment of rheumatoid arthritis and related rheumatic diseases. *Annals of the Rheumatic Diseases.* 2002;61(Sup 2):ii96–99.
- [34] Filla MS, Meyer KK, Faralli JA, Peters DM. Overexpression and activation of $\alpha\beta 3$ integrin differentially affects $\text{tgf}\beta 2$ signaling in human trabecular meshwork cells. *Cells.* 2021;10(8):1923.
- [35] Kontinen YT, Mackiewicz Z, Povilenaite D, Sukura A, Hukkanen M, Virtanen I. Disease-associated increased HIF-1, $\alpha\beta 3$ integrin, and Flt-1 do not suffice to compensate the damage-inducing loss of blood vessels in inflammatory myopathies. *Rheumatol Int.* 2004;24:333–9.
- [36] Liu Z, Wang F, Chen X. Integrin $\alpha\beta 3$ -targeted cancer therapy. *Drug Dev Res.* 2008;69(6):329–39.
- [37] Yang Y, Ko PJ, Pan YS, Lin HY, Whang-Peng J, Davis PJ, et al. Role of thyroid hormone-integrin $\alpha\beta 3$ -signal and therapeutic strategies in colorectal cancers. *J Biomed Sci.* 2021;28(1):24.
- [38] Hsieh MT, Wang LM, Changou CA, Chin YT, Yang YCSH, Lai HY, et al. Crosstalk between integrin $\alpha\beta 3$ and ER α contributes to thyroid hormone-induced proliferation of ovarian cancer cells. *Oncotarget.* 2017;8(15):24237–49.

- [39] Li J, Luo M, Ou H, Liu X, Kang X, Yin W. Integrin $\beta 4$ promotes invasion and anoikis resistance of papillary thyroid carcinoma and is consistently overexpressed in lymphovascular tumor thrombus. *J Cancer*. 2019;10(26):6635–48.
- [40] Liu S, Liang B, Gao H, Zhang F, Wang B, Dong X, et al. Integrin $\alpha\beta 6$ as a novel marker for diagnosis and metastatic potential of thyroid carcinoma. *Head Neck Oncol*. 2013;5(1):7.
- [41] Davis PJ, Mousa SA, Schechter GP. Platelet ATP, thyroid hormone receptor on integrin $\alpha\beta 3$ and cancer metastasis. *Horm Canc*. 2020;11:13–6.
- [42] Schmohl KA, Han Y, Tutter M, Schwenk N, Sarker R, Steiger K, et al. Integrin $\alpha\beta 3$ -dependent thyroid hormone effects on tumour proliferation and vascularisation. *Endocrine-related Cancer*. 2020;27(12):685–97.
- [43] Schmohl KA, Mueller AM, Dohmann M, Spellerberg R, Urnauer S, Schwenk N, et al. Integrin $\alpha\beta 3$ -mediated effects of thyroid hormones on mesenchymal stem cells in tumor angiogenesis. *Thyroid*. 2019;29(12):1843–57.
- [44] Abadi U, Weisz A, Kidron D, Katzav A, Hercbergs A, Davis PJ, et al. $\alpha\beta 3$ integrin expression and mitogenic effects by thyroid hormones in chronic lymphocytic leukemia. *J Clinical Med*. 2021;10(8):1766.
- [45] Meng R, Tang H-Y, Westfall J, London D, Cao JH, Mousa SA, et al. Crosstalk between integrin $\alpha\beta 3$ and estrogen receptor- α is involved in thyroid hormone induced proliferation in human lung carcinoma cells. *PLoS One*. 2011;6(11):e27547.
- [46] Fabio G, De Vito P, Valentina P, Hung-Yun L, Davis PJ, Pedersen JZ, et al. The role of thyroid hormones in hepatocyte proliferation and liver cancer. *Front Endocrinol*. 2019;10:532.
- [47] Davis FB, Mousa SA, O'Connor L, Mohamed S, Lin HY. Proangiogenic action of thyroid hormone is fibroblast growth factor-dependent and is initiated at the cell surface. *Circul Res*. 2004;94:1500–6.
- [48] Mousa SA, Davis FB, Mohamed S, Davis PJ, Feng X. Pro-angiogenesis action of thyroid hormone and analogs in a three-dimensional in vitro microvascular endothelial sprouting model. *Int Angiol*. 2006;25:407–13.
- [49] Yalcin M, Bharali DJ, Lansing L, Dyskin E, Mousa SA. Tetra iodo thyroacetic acid (tetrac) and tetrac nanoparticles inhibit growth of human renal cell carcinoma xenografts. *Anticancer Res*. 2009;29:3825–31.
- [50] Yalcin M, Dyskin E, Lansing L, Bharali DJ, Mousa SA. Tetra iodo thyroacetic acid (tetrac) and nanoparticulate tetrac arrest growth of medullary carcinoma of the thyroid. *J Clin Endocrinol Metab*. 2010;95:1972–80.
- [51] Yalcin M, Lin HY, Sudha T, Bharali DJ, Meng R. Response of human pancreatic cancer cell xenografts to tetra iodo thyroacetic acid nanoparticles. *Horm Cancer*. 2013;4:176–85.
- [52] Beatson GT. On the treatment of inoperable cases of carcinoma of the mamma: suggestions for a new method of treatment, with illustrative cases. *Trans Med Chir Soc Edin*. 1896;15:153–79.
- [53] Hercbergs A, Leith JT. Spontaneous remission of metastatic lung cancer following myxedema coma—an apoptosis-related phenomenon? *J Natl Cancer Inst*. 1993;85:1342–3.
- [54] Hercbergs AH, Ashur-Fabian O, Garfield D. Thyroid hormones and cancer: clinical studies of hypothyroidism in oncology. *Curr Opin Endocrinol Diabetes Obes*. 2010;17:432–6.
- [55] Moeller LC, Führer D. Thyroid hormone, thyroid hormone receptors, and cancer: a clinical perspective. *Endocr Relat Cancer*. 2013;20:R19–29.
- [56] Freindorf M, Furlani TR, Kong J, Cody V, Davis FB, Davis PJ. Combined QM/MM study of thyroid and steroid hormone analogue interactions with $\alpha\beta 3$ integrin. *J Biomed Biotechnol*. 2012;2012:959057.
- [57] Krashin E, Piekietko-Witkowska A, Ellis M, Ashur-Fabian O. Thyroid hormones and cancer: a comprehensive review of preclinical and clinical studies. *Front Endocrinol*. 2019;10:1664, <https://www.frontiersin.org/article/10.3389/fendo.2019.00059>.
- [58] Havaki S, Kouloukoussa M, Amawi K, Drosos Y, Arvanitis LD, Goutas N, et al. Altered expression pattern of integrin $\alpha\text{v}\beta 3$ correlates with actin cytoskeleton in primary cultures of human breast cancer. *Cancer Cell Int*. 2007;7:16.
- [59] Lin HY, Tang HY, Shih A, Keating T, Cao G. Thyroid hormone is a MAPK-dependent growth factor for thyroid cancer cells and is anti-apoptotic. *Steroids*. 2007;72:180–87.
- [60] Lin HY, Sun M, Tang HY, Lin C, Luidens MK, Mousa SA, et al. L-Thyroxine vs 3,5,3'-triiodo-L-thyronine and cell proliferation: activation of mitogen-activated protein kinase and phosphatidylinositol 3-kinase 1. *Am J Physiol Cell Physiol*. 2009;296:C980–91.
- [61] Mousa SA, Yalcin M, Bharali DJ, Meng R, Tang HY. Tetra iodo thyroacetic acid and its nanoformulation inhibit thyroid hormone stimulation of non-small cell lung cancer cells in vitro and its growth in xenografts. *Lung Cancer*. 2012;76:39–45.
- [62] Bergh JJ, Lin HY, Lansing L, Mohamed SN, Davis FB. Integrin $\alpha\text{v}\beta 3$ contains a cell surface receptor site for thyroid hormone that is linked to activation of mitogen-activated protein kinase and induction of angiogenesis. *Endocrinol*. 2005;146:2864–71.
- [63] Plow EF, Haas TA, Zhang L, Loftus J, Smith JW. Ligand binding to integrins. *J Biol Chem*. 2000;275:21785–88.
- [64] Zou W, Teitelbaum SL. Integrins, growth factors, and the osteoclast cytoskeleton. *Ann New York Acad Sci*. 2010;1192:27–31.
- [65] Roth P, Silgner M, Goodman SL, Hasenbach K, Thies S. Integrin control of the transforming growth factor- β pathway in glioblastoma. *Brain*. 2013;136:564–76.
- [66] Pinto M, Soares P, Ribatti D. Thyroid hormone as a regulator of tumor-induced angiogenesis. *Cancer Lett*. 2011;301:119–26.
- [67] Davis PJ, Davis FB, Mousa SA, Luidens MK, Lin HY. Membrane receptor for thyroid hormone: physiologic and pharmacologic implications. *Ann Rev Pharmacol Toxicol*. 2011;51:99–115.
- [68] Chiang AC, Massagué J. Molecular basis of metastasis. *New Eng J Med*. 2008;359:2814–23.
- [69] Haubner R, Wester HJ, Burkhart F, Schmidtke RS, Weber W, Goodman SL, et al. Glycosylated RGD-containing peptides: tracer for tumor targeting and angiogenesis imaging with improved biokinetics. *J Nuclear Med*. 2001;42:326–36.
- [70] Pleasure JR, Pleasure D, Pleasure SJ. Ch. 133: trophic factor, nutritional, and hormonal regulation of brain development. In: Polin RA, Abman SH, Rowitch DH, Benitz WE, Fox WW, Editor. *Fetal and neonatal physiology*. Fifth Edition New York, USA: Elsevier; 2017. p. 1326–1333.e3. ISBN 9780323352147. doi: 10.1016/B978-0-323-35214-7.00133-5.
- [71] Scanlan TS, Suchland KL, Hart ME, Chiellini G, Huang Y, Kruzich PJ, et al. 3-Iodothyronamine is an endogenous and

- rapid-acting derivative of thyroid hormone. *Nat Med.* 2004;10:638–42.
- [72] Davis PJ, Goglia F, Leonard JL. Non-genomic actions of thyroid hormone. *Nature Reviews Endocrinol.* 2016;12:111–21.
- [73] Cody V, Davis PJ, Davis FB. Molecular modeling of the thyroid hormone interactions with $\alpha\beta 3$ integrin. *Steroids.* 2007;72:165–70.
- [74] Cohen K, Ellis M, Khoury S, Davis PJ, Hercbergs A, Ashur-Fabian O. Thyroid hormone is a MAPK-dependent growth factor for human myeloma cells acting via $\alpha\beta 3$ integrin. *Mol Cancer Res.* 2011;9:1385–95.
- [75] Wang XQ, Lindberg FP, Frazier WA. Neuronal roles of integrin-associated protein (IAP/CD47). *J Cell Biol.* 1999;147:389.
- [76] Lin HY, Tang HY, Keating T. Resveratrol is pro-apoptotic and thyroid hormone is anti-apoptotic in glioma cells: both actions are integrin and ERK-mediated. *Carcinogen.* 2008;29:62–9.
- [77] Rebbaa A, Chu F, Davis FB. Novel function of the thyroid hormone analog tetraiodo thyro acetic acid: a cancer chemo-sensitizing and anti-cancer agent. *Angiogen.* 2008;11:269–76.
- [78] Mousa SA, O'Connor LJ, Bergh JJ, Davis FB, Scanlan TS, Davis PJ. The proangiogenic action of thyroid hormone analogue GC-1 is initiated at an integrin. *J Cardiovasc Pharmacol.* 2005;46:356–60.
- [79] Bridoux A, Khan RA, Chen C, Chev   G, Cui H, Dyskin E, et al. Design, synthesis, and biological evaluation of bifunctional thyrointegrin inhibitors: new anti-angiogenesis analogs. *J Enzyme Inhib Med Chem.* 2011;26(6):871–82. doi: 10.3109/14756366.2011.557023.
- [80] Mousa SA, O'Connor L, Davis FB, Davis PJ. Pro-angiogenesis action of the thyroid hormone analog 3, 5-diiodothyropropionic acid (DITPA) is initiated at the cell surface and is integrin-mediated. *Endocrinol.* 2006;147:1602–7.
- [81] Davis PJ, Mousa SA. Uses of formulations of thyroid hormone antagonists and nanoparticulate forms thereof to increase chemo-sensitivity and radio-sensitivity in tumor or cancer cells. European Patent EP 2442800A2, EP 2662079 A1; 2010.
- [82] Bridoux A, Cui H, Dyskin E, Yalcin M, Shaker MA. Semi-synthesis and pharmacological activities of Tetrac analogs: angiogenesis modulators. *Bioorg Med Chem Lett.* 2009;19:3259–63.
- [83] Hall LC, Salazar EP, Kane SR, Liu N. Effects of thyroid hormones on human breast cancer cell proliferation. *J Steroid Biochem Mol Biol.* 2008;109:57–66.
- [84] Mousa SA, Thangirala S, Lin HY, Tang HY, Glinsky GV, Davis PJ. MicroRNA-21 and microRNA-15A expression in human breast cancer (MDA-MB-231) cells exposed to nanoparticulate tetraiodo thyroacetic acid (Nanotetrac). Meeting of the Endocrine Society; 2014. p. Abstract SUN-0472.
- [85] Zhao D, Tu Y, Wan L, Bu L, Huang T, Sun X, et al. In vivo monitoring of angiogenesis inhibition via down-regulation of mir-21 in a VEGFR2-luc murine breast cancer model using bioluminescent imaging. *PLoS One.* 2013;8:e71472.
- [86] Sun CY, She XM, Qin Y, Chu ZB, Chen L, Ai LS, et al. miR-15a and miR-16 affect the angiogenesis of multiple myeloma by targeting VEGF. *Carcinogen.* 2013;34:426–35.
- [87] Lin HY. COX-2 and p53 dependent apoptosis in head squamous cell cancer cell. *J Cell Biochem.* 2008;104:2131–42.
- [88] Lin HY, Sun M, Lin C, Tang HY, London D, Shih A, et al. Androgen-induced human breast cancer cell proliferation is mediated by discrete mechanisms in estrogen receptor-alpha-positive and -negative breast cancer cells. *J Steroid Biochem Mol Biol.* 2009;113:182–8.
- [89] Glinskii AB, Glinsky GV, Lin HY, Tang HY, Sun M, Davis FB, et al. Modification of survival pathway gene expression in human breast cancer cells by tetra iodo thyro acetic acid (Tetrac). *Cell Cycle.* 2009;8(21):3562–70.
- [90] Kalas W, Yu JL, Milsom C, Rosenfeld J, Benezra R, Bornstein P, et al. Oncogenes and angiogenesis: Downregulation of thrombospondin-1 in normal fibroblasts exposed to factors from cancer cells harboring mutant RAS. *Cancer Res.* 2005;65:8878–86.
- [91] Yang M, Chen T, Han C, Li N, Wan T, Cao X. Rab7b, a novel lysosome-associated small GTPase, is involved in monocytic differentiation of human acute promyelocytic leukemia cells. *Biochem Biophys Res Comm.* 2004;318:792–9.
- [92] Ren B, Yee KO, Lawler J, Khosravi-Far R. Regulation of tumor angiogenesis by thrombospondin-1. *Biochim Biophys Acta.* 2006;1765:178–88.
- [93] Mousa SA, Bergh JJ, Dier E. Tetra iodo thyro acetic acid, a small molecule integrin ligand, blocks angiogenesis induced by vascular endothelial growth factor and basic fibroblast growth factor. *Angiogen.* 2008;11:183–90.
- [94] Davis PJ, Davis FB, Mousa SA. Thyroid induced angiogenesis. *Curr Cardio Rev.* 2009;5:12–6.
- [95] Zi-Ming Z, Reynolds AB, Gaucher EA. The evolutionary history of the catenin gene family during metazoan evolution. *BMC Evolution Biol.* 2011;11:198.
- [96] Justilien V, Walsh MP, Ali SA, Thompson EA, Murray NR, Fields AP. The PRKCI and SOX2 oncogenes are co-amplified and cooperate to activate hedgehog signaling in lung squamous cell carcinoma. *Cancer Cell.* 2014;25:139–51.
- [97] Samar  ija I, Dekani   A, Humphries JD, Parad  ik M, Stojanovi   N, Humphries MJ et al. Integrin crosstalk contributes to the complexity of signaling and unpredictable cancer cell fates. *Cancers (Basel).* 2020;12(7):1910.
- [98] Cheng TM, Chang WJ, Chu HY, De Luca R, Pedersen JZ, Incerpi S, et al. Nano-strategies targeting the integrin $\alpha\beta 3$ network for cancer therapy. *Cells.* 2021;10(7):1684.
- [99] Go K, Sudha T, Davis PJ, Mousa S. Nano diamino propane tetrac and integrin $\alpha\beta 3$ expression in different cancer types: anti-cancer efficacy and safety. *Cancer Treat Res Comm.* 2021;28(1):100395.
- [100] Chin YT, He ZR, Chen CL, Chu HC, Ho Y, Su PY, et al. Tetrac and NDAT induce anti-proliferation via integrin $\alpha\beta 3$ in colorectal cancers with different K-RAS status. *Front Endocrinol (Lausanne).* 2019;10:130.
- [101] Eldar-Boock A, Blau R, Ryppa C, Baabur-Cohen H, Many A, Vicent MJ, et al. Integrin-targeted nano-sized polymeric systems for paclitaxel conjugation: a comparative study. *J Drug Target.* 2017;25:829–44.
- [102] Graf N, Bielenberg DR, Kolishetti N, Muus C, Banyard J, Farokhzad OC, et al. $\alpha\beta 3$ integrin-targeted PLGA-PEG nanoparticles for enhanced anti-tumor efficacy of a Pt (IV) prodrug. *ACS Nano.* 2012;6:4530–9.
- [103] Saraf P, Li X, Wrischnik L, Jasti B. In vitro and in vivo efficacy of self-assembling RGD peptide amphiphiles for targeted delivery of paclitaxel. *Pharm Res.* 2015;32:3087–101.

- [104] Zhang L, Su H, Wang H, Li Q, Li X, Zhou C, et al. Tumor chemoradiotherapy with rod-shaped and spherical gold nano probes: shape and active targeting both matter. *Theranost.* 2019;9:1893–908.
- [105] Maeda H, Tsukigawa K, Fang J. A retrospective 30 years after discovery of the enhanced permeability and retention effect of solid tumors: next-generation chemotherapeutics and photodynamic therapy—problems, solutions, and prospects. *Microcircul.* 2016;23(3):173–82.
- [106] Rajabi M, Godugu K, Sudha T, Bharali DJ, Mousa SA. Triazole modified tetraiodothyroacetic acid conjugated to polyethylene glycol: high affinity thyrointegrin $\alpha\text{v}\beta 3$ antagonist with potent anticancer activities in glioblastoma multiforme. *Bioconjug Chem.* 2019;30(12):3087–97.
- [107] Fujioka K, Godugu K, Mousa SA. Pharmacokinetics and biodistribution of a novel anticancer thyrointegrin $\alpha\text{v}\beta 3$ antagonist: triazole modified tetraiodothyroacetic acid conjugated to polyethylene glycol (P-bi-TAT). *AAPS Open.* 2021;7:2.
- [108] Reeves KJ, Hurrell JE, Cecchini M, van der Pluijm G, Down JM, Eaton CL, et al. Prostate cancer cells home to bone using a novel in vivo model: modulation by the integrin antagonist GLPG0187. *Int J Cancer.* 2015;136(7):1731–40.
- [109] Gramoun A, Shorey S, Bashutski JD, Dixon SJ, Sims SM, Heersche JN, et al. Effects of Vitaxin, a novel therapeutic in trial for metastatic bone tumors, on osteoclast functions in vitro. *J Cell Biochem.* 2007;102(2):341–52.
- [110] Hersey P, Sosman J, O'day S, Richards J, Bedikian A, Gonzalez R, et al. Etaracizumab Melanoma Study Group. A randomized phase 2 study of etaracizumab, a monoclonal antibody against integrin $\alpha\text{v}\beta 3$ + or - dacarbazine in patients with stage IV metastatic melanoma. *Cancer.* 2010;116(6):1526–34.
- [111] Khanna D, Tashkin DP, Wells AU, Seibold JR, Wax S, Vazquez-Mateo C, et al. STRATUS: a phase II study of abituzumab in patients with systemic sclerosis-associated interstitial lung disease. *J Rheumatol.* 2021;48(8):1295–8.
- [112] Hariharan S. Assessment of the biological and pharmacological effects of the $\alpha\text{v}\beta 3$, and $\alpha\text{v}\beta 5$ Integrin receptor antagonist, Cilengitide (EMD 121974), in patients with advanced solid tumors. *Ann Oncol.* 2007;18(8):1400–7.
- [113] Ji-Young B. Pharmacoproteomic analysis of a novel cell-permeable peptide inhibitor of tumor-induced angiogenesis. *Mol Cell Proteomics.* 2011;10(8):M110.005264.
- [114] Youngjin C. Site-specific inhibition of integrin $\alpha\text{v}\beta 3$ -vitronectin association by a Ser-Asp-Val sequence through an Arg-Gly-Asp-binding site of the integrin. *Proteomics.* 2010;10(1):72–80.
- [115] Wierzbicka-Patynowski I. Structural requirements of echistatin for the recognition of $\alpha\text{v}\beta 3$ and $\alpha 5\beta 1$ integrins. *J Biol Chem.* 1999;274(53):37809–14.
- [116] Xiaoyan Z. An integrin antagonist (MK-0429) decreases proteinuria and renal fibrosis in the ZSF1 rat diabetic nephropathy model. *Pharmacol Res Perspect.* 2017;5(5):e00354.