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Detection of *CagA*, *VacA*, *IceA1* and *IceA2* virulent genes in *Helicobacter pylori* isolated from gastric ulcer patients

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Abstract

Background: Virulence factors of *Helicobacter pylori* including *cagA*, *vacA*, *iceA* and their association with clinical manifestation varied widely with different subpopulations. The objective of the study was to determine the prevalence of *cagA*, *iceA1*, *iceA2*, *vacA*, *vacA s1/s2*, *vacA m1/m2*, Western type *cagA* and East Asian type *cagA* virulence genes in *H. pylori* isolated from gastric ulcer patients and evaluate the association of these genes with gender, age, smoking and alcohol consumption.

Methods: Gastric biopsy samples from 172 patients were collected. *H. pylori* virulence genes, *cagA*, *vacA*, *iceA1*, *iceA2*, *vacA s1/s2*, *vacA m1/m2*, Western type *cagA* and East Asian type *cagA* were detected using polymerase chain reaction (PCR).

Results: Of the gastric biopsy samples collected, 48.3% of samples grew *H. pylori*. The *vacA* (68.7%) was the predominant virulence gene detected and associated with male patients and patients within the age group of 31–40 years. The *cagA* was the second most common gene detected and significantly associated with alcoholic patients.

Conclusions: *H. pylori* infection rate was 48.3% and was associated with patients who were smokers or had a history of smoking. The majority of our isolates were positive for any one of the virulence genes tested indicating that these isolates were highly virulent in nature.

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Keywords: gastric ulcer; *H. pylori*; virulence genes.

Introduction

Helicobacter pylori is associated with gastrointestinal disorders including peptic ulcers, gastric cancer, chronic gastritis and lymphoma [1, 2]. Various factors such as age, ethnicity, geography and socio-economic status contribute to the varied prevalence of H. pylori [1]. H. pylori incidence tends to be low in many developed countries due to improved social-economic status, while it remains high in developing countries [3, 4]. H. pylori is highly prevalent in some countries; however, the disease severity varies from one region to another mainly due to the differences in the virulence factors among the strains [5]. Several virulence factors play an important role in the pathogenesis of the disease, which includs the cytotoxin-associated gene A (cagA), vacuolating cytotoxin A (vacA) and iceA genes [5–7]. The cagA gene encoded by the CagA protein is found in more than 50% of the *H. pylori* isolates. The cagA gene is associated with increased production of interleukin-8 (IL-8), mucosal inflammation, nuclear factor-kB activation and the development of peptic ulcer disease (PUD) and gastric cancer (GC). The cagA gene, a marker of the cag pathogenicity island has been associated with PUD, atrophic gastritis and adenocarcinoma [7–9]. About 60%– 70% of *H. pylori* strains tend to possess the cagA gene [8, 9]. The vacuolating toxin encodes the *vacA* gene, consists of three variable regions, the signal peptide encoding s region and two alleles (s1 and s2). The s1 region is further subdivided into s1a, s1b and s1c subtypes. The middle region (m region) of the vac gene has two alleles, m1 and *m*2 [10]. The activity of the *vac* gene was determined by the combination of s- and m-regions, the s1m1 combination was considered as the most virulent and produces a high amount of toxin, while the combination of s2m2 produces an inactive toxin [10, 11]. The cagA and vacA genes play an important role in determining the clinical outcomes of the infected patients [12]. The iceA gene induced by the contact

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with human gastric cells produces a high level of IL8 and is highly associated with PUD [13]. The *iceA* has two alleles, iceA1 and iceA2 [14].

The prevalence of *H. pylori* virulence factors including cagA, iceA and vacA and their association with clinical manifestation varied widely with different subpopulations [15–17]. However, studies regarding the prevalence of cagA and iceA, vacA genes are scarce. Hence, the present study determined the prevalence of cagA, iceA1, iceA2, vacA, vacA s1/s2, vacA m1/m2, Western type cagA and East Asian type *cagA* virulence genes in *H. pylori* isolated from gastric ulcer patients. The study also evaluated the association of these virulence genes with gender, age, smoking and alcohol consumption.

Materials and methods

One hundred and seventy-two consecutive gastric biopsy samples from 172 patients with a gastric ulcer, admitted to the endoscopic section at the Department of Gastroenterology, Jining First People's Hospital, Shandong Province and the Department of Gastroenterology, Third Affiliated Hospital of Shandong Medical College, Shandong Province, China between September 2015 and December 2017 were collected. Patients who received prior antibiotic therapy within the previous 4 weeks were excluded from the study. An informed consent was obtained from all patients or their legal heir. The hospital Ethics Committee approved the study. Using a sterile container sample was collected aseptically and immediately sent for *H. pylori* culture.

Isolation and identification of H. pylori

The collected samples were aseptically dissected into two pieces for rapid urease test and H. pylori culture. Using the modified rapid urease agar medium, the rapid urease test was performed [18]. Briefly, after collection one part of the biopsy samples were immediately inoculated into urea agar and incubated at room temperature for 2 h. The change in color of the medium from yellow to red indicates a positive reaction. Helicobacter pylori culture was performed by inoculating the biopsy samples on the modified Columbia agar plates (Merck, Germany) supplemented with horse lysed blood (10%, v/v). The plates were then incubated in an anaerobic condition at 37 °C for 2–3 days under microaerobic conditions [19]. After incubation, based on the colony morphology, gram staining, positive oxidase, catalase, and urease tests the organisms were identified as H. pylori. The isolates were stored in Columbia broth supplemented with sterile glycerol (20%, v/v) at −70 °C until used.

DNA extraction

The H. pylori isolates were subjected to DNA extraction by the boiling lysis method, briefly, 200 µL of H. pylori culture was incubated at 95°C for 10 min, spun at 14,000 g for 10 min to collect the supernatant containing DNA. Both the DNA samples were stored at -20 °C until further analysis. DNA quantification and purity was determined using NanoDrop ND 1000 (Nano-Drop Technologies, Wilmington, DE, USA).

Detection of virulence genes

Detection of cagA, vacA, iceA1 and iceA2 virulence genes were performed using polymerase chain reaction (PCR). The primer sequences, cycling conditions and expected amplicon sizes are described in Table 1. Two duplex PCR was performed to detect the iceA1/iceA2 and cagA/vacA genes separately. PCR was done using a 50 µL master mix containing 5 µL of template DNA, 0.4 µM of each primer, 200 µM of dNTPs, one unit of Taq polymerase enzyme and 5 μL of 10× reaction buffer. After PCR, the amplicons were resolved in 1.5% agarose gel and visualized under a UV transilluminator.

Detection of caqA and VacA gene variance

The *cagA* and *vacA* variant genes, Western type *cagA*, East Asian cagA and vacA s1/s2 and vacA m1/m2 genes were detected by PCR. The primer sequences, cycling conditions and expected amplicon sizes were described in Table 1. Four different PCRs were performed to detect the Western type cagA, East Asian cagA, vacA s1/s2 and vacA *m1/m2* genes, respectively. The Western type *cagA* gene PCR was expected to yield two amplicons of sizes ~218-227 and ~174–177 bp. Similarly, the East Asian cagA was expected to yield two amplicons of size ~293-299 bp and ~455-461 bp. The PCR master mix was prepared as described above. After PCR, the amplicons were resolved in 1.8% agarose gel and visualized under a UV transilluminator.

Statistics

Continuous variables were presented as mean and ranges; categorical variables as numbers and percentages.

Table 1: Primer sequence of *H. pylori* virulence genes.

Genes	Primers	Size, bp	Cycling conditions	References
iceA1	F: 5'-GTGTTTTTAACCAAAGTATC-3'	246	95 °C, 1 min; 50 °C, 1 min; 72 °C, 1 min	[20]
	R: 5'-CTATAGCCATTATCTTTGCA-3'		(30 cycles)	
iceA2	F: 5'-GTTGGGTATATCACAATTTAT-3'	229/334ª		
	R: 5'-TTTCCCTATTTTCTAGTAGGT-3'			
cagA	F-5'-GATAACAGCCAAGCTTTTGAGG-3'	349	94°C, 1 min; 55°C, 1 min; 72°C, 1 min	[21]
	R-5'-CTGCAAAAGATTGTTTGG <i>CAGA</i> -3'		(35 cycles)	
vacA	F-5'-ATGGAAATACAACAAACACAC-3'	286		
	R-5'-CTGCTTGAATGCGCCAAAC-3'			
vacA s1/s2	F-5'-ATGGAAATACAACAAACACAC-3'	259/286	95 °C, 50 s; 58 °C, 50 s;	[1]
	R-5'-CTGCTTGAATGCGCCAAAC-3'		72°C,50 s (35 cycles)	
vacA m1/m2	F-5'-CAATCTGTCCAATCAAGCGAG-3'	567/642	95 °C, 50 s; 54 °C, 50 s;	
	R-5'-GCGTCAAAATAATTCCAAGG-3'		72°C,50 s (35 cycles)	
Western	F-5'-ACCCTAGTCGGTAATGGG-3'	~218-227 and	95 °C, 1 min; 52 °C, 1 min;	[22]
type <i>cagA</i>	R-5'-TGCCCTACAMCACCSAAACCAC-3'	~174-177	72 °C, 1 min (35 cycles)	
(cagT/cag W)	F-5'-AAAAATTGACCRACTCAATC-3'			
	R-5'-GCTTTAGCTTCTGAYACYGC-3'			
East Asian	F-5'-ACCCTAGTCGGTAATGGG-3'	~293-299	95°C, 1 min; 52°C, 1 min;	[5]
type <i>cagA</i>	R-5'-GCAATTTTGTTAATCCGGTC-3'	~ 455-461	72 °C, 1 min (35 cycles)	
(cagT/cag j)	F-5'-GCATCAGCAGGTAAAGGAGT-3'			
	R-5'-GCTTTAGCTTCTGAYACYGC-3'			

The primers yield a fragment of 229 or 334 bp depending on the presence of a repetitive sequence of 105 nucleotides codifying for 35 amino acids in some iceA2 alleles.

Chi-square tests were performed to determine the statistical significance using MINITAB statistical software (MINITAB ver. 13.1; MINITAB Inc., PA, USA). The data was considered statistically significant if the p-value was < 0.05.

Results

Of the 172 patients, 97 (56.4%) were male and 75 (43.6%) were female. The mean age was 46 ± 7.2 (range 19-63) years. Of the included patients, 113 (65.7%) were smokers or had a history of smoking and 108 (62.8%) were alcoholic patients. Demographic details of the patients are given in Table 2. Of the 172 gastric biopsy samples, 118 (68.6%) samples were positive for rapid urease test and 83 (48.3%) samples (48 from male and 35 from female) grew *H. pylori*. All the samples which isolated H. pylori were found to be positive for the rapid urease test. No significant association was found between H. pylori infection in male and female (27.9% vs. 20.3%; p = 0.714) patients and between alcoholic and non-alcoholic (30.2% vs. 18.0%; p = 0.971) patients. However, we observed a significant association between H. pylori infection and patients who were smokers or had a history of smoking (26.7%; p=0.006). We found that patients within the age group of 41–50 years were highly likely to be *H. pylori* positive (18.0%) followed

Table 2: Baseline characteristics of patients.

Variables	No. of patients
Male	97 (56.4%)
Female	75 (43.6%)
Mean age (SD) years	46±7.2
Age, years	
19-30	32 (18.6%)
31-40	53 (30.8%)
41-50	64 (37.2%)
51-63	23 (13.4%)
Smoker	113 (65.7%)
Non-smoker	59 (34.3%)
Alcoholic	108 (62.8%)
Non-alcoholic	64 (37.2%)

SD, standard deviation.

by patients within the age group of 31–40 years (14.5%). However, no significant association was found between the *H. pylori* infection and different age groups (p = 0.979).

Of the 83 isolates subjected to PCR, 57 (68.7%) isolates were positive for the vacA gene, 53 (63.9%) isolates were positive for the cagA gene, 46 (55.4%) isolates were positive for the iceA2 gene and 37 (44.6%) isolates were positive for the iceA1 gene. We found that the presence of vacA (49.4% vs. 19.3%; p = 0.000) and iceA2 (26.5% vs. 29%; p = 0.040) genes were significantly associated with male and female patients, respectively. When analyzed

Table 3: Detection and distribution of *H. pylori* virulence genes.

Variables	Culture positive (n=172)	Distribution of virulence genes among the isolates (n=83)				
		cagA	vacA	iceA1	iceA2	
Male	48 (27.9%)	34 (41.0%)	41 (49.4%)	20 (24.1%)	22 (26.5%)	
Female	35 (20.3%)	19 (22.9%)	16 (19.3%)	17 (20.5%)	24 (29.0%)	
χ²-Test	p = 0.714	p = 0.121	p = 0.000	p = 0.532	p = 0.040	
19-30 years	15 (8.7%)	8 (9.6%)	12 (14.5%)	7 (8.4%)	10 (12.0%)	
31-40 years	25 (14.5%)	21 (25.3%)	28 (33.7%)	18 (21.7%)	14 (16.9%)	
41-50 years	31 (18.0%)	17 (20.5%)	14 (16.9%)	6 (7.2%)	16 (19.3%)	
51-63 years	12 (7.0%)	7 (8.4%)	3 (3.6%)	7 (7.2%)	6 (7.2%)	
χ²-Test	p = 0.979	p = 0.095	p = 0.000	p = 0.010	p = 0.815	
Smoker	46 (26.7%)	28 (33.7%)	33 (39.8%)	19 (22.9%)	24 (29.0%)	
Non-smoker	37 (21.5%)	25 (30.1%)	24 (29.0%)	18 (21.7%)	22 (26.5%)	
χ²-Test	p = 0.006	p = 0.528	p = 0.502	p = 0.503	p = 0.507	
Alcoholic	52 (30.2%)	38 (45.8%)	37 (44.6%)	27 (32.5%)	31 (37.3%)	
Non-alcoholic	31 (18.0%)	15 (18.1%)	20 (24.1%)	10 (12.0%)	15 (18.1%)	
χ²-Test	p = 0.971	p=0.024	p = 0.528	p = 0.081	p = 0.076	

Bold values indicates statistical significant values.

between different age groups vacA (33.7%; p=0.000) and iceA1 (21.7%; p=0.010) genes were significantly associated with the age group of 31–40 years. The presence of cagA gene was significantly associated with alcoholic patients (45.8% vs. 18.1%; p = 0.024). There was no significant association between the tested virulence genes and smoking history of the patients (p > 0.05). We also found that there was no statistically significant association of the iceA2 gene in relation to gender, age, smoking and alcoholic history of the patients (p > 0.05). Details on the distribution of virulence genes among the isolates are given in Table 3.

Among the isolates tested for virulence genes, 20 isolates, each were positive for cagA and iceA2 genes alone, respectively. In the combinational analysis, we found that 18 (21.7%) isolates were positive for both *vacA* and *iceA2* genes, which was the most predominant combination detected. The cagA and vacA, cagA and iceA1 were the next most common combinations detected, respectively, in each of the 12 (14.5%) isolates. Three isolates were positive for the *cagA*, *vacA* and *iceA1* genes, three isolates were positive for the cagA, iceA1 and iceA2 genes. Another three isolates were positive for all the tested (cagA, vacA, iceA1, iceA2) virulence genes (Table 4). In the present study, 89.2% (74/83) of *H. pylori* isolates were positive for at least one of the tested virulence genes (cagA, vacA, iceA1, iceA2) which implies that a majority of the H. pylori isolates had virulence potential as evidenced by the detection of virulence genes.

Of the 83 isolates tested for vacA and cagA gene variance, 52 (62.7%) isolates were positive for the vacA s1/s2 gene, 61 (73.5%) isolates were positive for the vacA m1/ m2 gene, 12 (14.5%) isolates were positive for the Western type cagA gene and 56 (67.5%) isolates were positive for the East Asian type *cagA* gene. We found that the presence of vacA s1/s2 (42.2% vs. 20.5%; p=0.024) and East Asian type cagA (28.9% vs. 38.6%; p=0.004) genes were significantly associated with male and female patients, respectively. When analyzed between different age groups the vacA s1/s2 (28.9%; p=0.000) gene was significantly associated with the age group of 31–40 years. The presence of the vacA m1/m2 gene was significantly associated with non-smoking patients (42.2% vs. 31.3%; p = 0.000). The presence of the vacA s1/s2 gene was significantly associated with alcoholic patients (49.4% vs. 13.3%; p = 0.000). We also found that there was no statistically significant association of the Western type cagA gene in relation to gender, age, smoking and alcoholic history of the patients (p>0.05). Details on the distribution of vacA and cagA

Table 4: Detection and combination of H. pylori virulence genes.

Gene combinations	No. of isolates (n=83)	
cagA	20 (24.1%)	
vacA	13 (15.7%)	
iceA1	8 (9.6%)	
iceA2	20 (24.1%)	
cagA+vacA	12 (14.5%)	
cagA+iceA1	12 (14.5%)	
vacA+iceA1	6 (7.2%)	
vacA+iceA2	18 (21.7%)	
cagA+vacA+iceA1	3 (3.6%)	
cagA+iceA1+iceA2	3 (3.6%)	
vacA+iceA1+iceA2	2 (2.4%)	
cagA+vacA+iceA1+iceA2	3 (3.6%)	

Table 5: Distribution of vacA and cagA gene variance among the isolates (n=83).

Variables	Distribution of <i>vacA</i> and <i>cagA</i> gene variance among the isolates (n=83)				
	vacA s1/s2	vacA m1/m2	Western type <i>cagA</i>	East Asian type <i>cagA</i>	
Male	35 (42.2%)	32 (38.6%)	5 (6%)	24 (28.9%)	
Female	17 (20.5%)	29 (34.9%)	7 (8.4%)	32 (38.6%)	
χ²-Test	p = 0.024	p = 0.099	p = 0.220	p = 0.004	
19-30 years	8 (9.6%)	12 (14.5%)	1 (1.2%)	13 (15.7%)	
31-40 years	24 (28.9%)	21 (25.3%)	1 (1.2%)	15 (18.1%)	
41-50 years	11 (13.3%)	18 (21.7%)	8 (9.6%)	20 (24.1%)	
51-63 years	9 (10.8%)	10 (12%)	2 (2.4%)	8 (9.6%)	
χ²-Test	p = 0.000	p = 0.106	p = 0.101	p = 0.350	
Smoker	28 (33.7%)	26 (31.3%)	5 (3.6%)	29 (34.9%)	
Non-smoker	24 (28.9%)	35 (42.2%)	7 (10.8%)	27 (32.5%)	
χ²-Test	p = 0.706	p = 0.000	p = 0.322	p = 0.337	
Alcoholic	41 (49.4%)	39 (47%)	6 (7.2%)	36 (43.4%)	
Non- alcoholic	11 (13.3%)	22 (26.5%)	6 (7.2%)	20 (24.1%)	
χ^2 -Test	p = 0.000	p = 0.687	p = 0.327	p = 0.657	

Bold values indicates statistical significant values.

variance genes among the isolates are given in Table 5. Analysis of the *vacA* variance genes showed that five (6%) isolates amplified only the vacA s1 gene, seven (8.4%) isolates amplified only the *vacA* s2 gene, four (4.8%) isolates amplified only *vacA m1* gene and six (7.2%) isolates amplified the only vacA m2 gene, the presence of these genes did not differ significantly (p > 0.05).

When compared between vacA and vacA s1/s2, vacA m1/m2 gene-positive isolates; a total of 32 (38.6%) isolates amplified the vacA s1/s2, vacA m1/m2 genes. Three isolates which were positive for the *vacA* gene did not amplify either vacA s1/s2 or vacA m1/m2 genes. Five and seven isolates which were positive for the vacA s1/s2 and vacA m1/m2 genes, respectively, did not amplify the vacA gene. When compared between cagA and cagA Western and East Asian type gene-positive isolates; four isolates which were positive for the *cagA* gene did not amplify either the cagA Western type or cagA East Asian type genes. Two and eight isolates which were positive for the cagA Western type and cagA East-Asian type genes, respectively, did not amplify the *cagA* gene.

Discussion

H. pylori, a causative agent of gastric ulcers, can lead to the development of GC [23]. Although the H. pylori prevalence

is declining in many developed countries [24, 25], it remains high in developing countries and ranges from 70% to 90% [3, 4, 26, 27]. The association of *H. pylori* with various gastrointestinal diseases makes it a major health problem, worldwide [1, 2, 28]. In our study, the H. pylori prevalence rate among patients with gastric ulcer was 48.3%. A study from Pakistan, reported the prevalence of *H. pylori* at a rate of 53% and 80%, respectively, in patients with duodenal ulcer and individuals presenting symptoms of various upper gastrointestinal diseases [29, 30]. This was higher than that reported (48.3%) in our study from gastric ulcer patients. Another study from Bangladesh conducted in patients with a gastric ulcer reported H. pylori prevalence at a rate of 75% which was higher than that reported in our study [26]. A study from Singapore which assessed *H. pylori* infection in patients with gastric ulcer using a Campylobacter-like organism (CLO) test and the histology reported a prevalence rate of 67.5%, which is higher than that reported in our study [31]. A study from Mongolia which included dyspeptic patients from all parts of Mongolia reported an overall H. pylori infection at a rate of 80%, which was higher than that reported in our study [32]. The prevalence of *H. pylori* in our study is lower than that reported from dyspeptic patients in Bhutan (86%) [33]. Among the dyspeptic patients, the prevalence of *H. pylori* in African countries including Morocco (75.5%), Ethiopia (65.7%) and Nigeria (93.6%) was higher than that reported in our study [34–36]. Due to rapid urbanization in China, the prevalence of H. pylori infection has decreased and could be a reason for the low prevalence of *H. pylori* in our study [24]. In our study, there was no significant association between the presence of *H. pylori* infection and gender (p = 0.714) and alcohol consumption (p = 0.971). Smoking is considered as one of the independent risk factors for *H. pylori* infection; in our study, we found a significant (p = 0.006) association between the presence of *H. pylori* and patients who were smokers or had a history of smoking [37].

H. pylori possess several virulence factors which are associated with various gastrointestinal diseases [5-7]. H. pylori adheres to the mucosa of the stomach, colonizes it, persists for a long time and leads to an inflammatory response. The colonization of H. pylori as such does not cause infection; however, it can increase the risk of developing various gastrointestinal diseases including gastric ulcer, GC and lymphoma [38]. Specific analysis of H. pylori virulence factors can predict post H. pylori infectious disorders [38]. In our study, vacA (68.7%) was the predominant gene detected among our isolates followed by cagA (63.9%), iceA2 (55.4%) and iceA1 (44.6%) genes. A meta-analysis which investigated

the association of H. pylori virulence factors with GC reported that the presence of *cagA* and *vacA* genes were significantly associated with GC [38]. In our study, 63.9% isolates were positive for the cagA gene which is higher (50%) than that reported in the biopsy samples collected from gastric patients from Northeast China [23]. The presence of cagA in our study is lower than that reported in India (90.9%), detected in the biopsy samples from gastric ulcer patients, which also reported a significant association between peptic ulcer and the presence of the cagA gene [39]. Another study from India reported that 77.27% of biopsy samples from acid-peptic disease were positive for the *cagA* gene, which was higher than that reported in our study. The study also reported that the presence of the cagA gene was significant in female patients; however, in our study there was no significant (p=0.121) difference found between the two genders [40]. Another study from Thailand reported that 98.2% of their gastric biopsy samples from dyspeptic patients were positive for the cagA gene [41]. In contrast, a lower rate of the cagA gene was reported in biopsy samples from gastric ulcer patients from Saudi Arabia (30%) and stool samples from asymptomatic individuals from Japan [1, 42]. The presence of the cagA in our study is much lower than that reported in H. pylori isolated from gastric biopsy samples from Korea (99%) [22].

We reported that 14.5% and 67.5% of our isolates were positive for Western and East Asian type cagA genes, respectively. As previously reported, East Asia type cagA was more prevalent in Asian countries, the majority of our isolates were positive for the East Asian type cagA gene compared to the Western type cagA gene [43]. In contrast, a study from Northeast China reported that 31% of their biopsy samples from gastric ulcer patients were positive for the Western type *cagA* gene which is higher and 20% of their samples were positive for the East Asian type *cagA* gene which is lower than that reported in our study [23]. In contrast, a study which determined the prevalence of H. pylori infection and genotypes of virulence factors in a Nepalese population reported, a higher rate of the Western type cagA gene (94.1%) compared to the East Asian type cagA gene (5.9%) [44]. The East Asian cagA is more virulent than Western type cagA [45, 46]. Additionally, the East Asian cagA gene plays a potential role in the development of gastric diseases caused by H. pylori infections [47]. Thus, the high presence of the East Asian *cagA* gene in our isolates indicates that our isolates are highly virulent in nature. Also, the significant association of the East Asian cagA gene with female patients (p=0.004) in our study indicates that they may be at high risk of *H. pylori* infection with significant clinical manifestation.

In our study, 68.7% isolates were positive for the vacA gene which was higher than that reported from India (4.5%) [40]. The vacA m1/m2 (73.5%) was the predominant vacA variant gene reported in our study. It has been reported that the presence of *vacA s1* and *m1* genotypes may indicate an increased risk of peptic ulcer and GC in Latin America and the Middle East [38]. In our study, 38.6% of isolates were positive for *vacA s1/s2* and *vacA m1/m2* genes indicating that the patients from whom these isolates were isolated are at a risk of GC in addition to the gastric ulcer they harbor. A study from Saudi Arabia reported the combined vacA s1/s2 gene in 30% of isolates and vacA m1/m2 in 0% of *H. pylori* isolates from gastric ulcer patients which were lower than that reported from our study [1]. The study reported that vacA s1/s2 was the predominant genotype in gastric ulcer patients; in contrast, vacA m1/m2 was the predominant genotype in our study [1]. We found that the vacA and vacA s1/s2 genes were significantly associated with male patients and patients within the age group of 23-40 years. Additionally, vacA s1/s2 was significantly associated with alcoholic patients. In our study, the cooccurrence of vacA variance and cagA variance gene combinations did not differ significantly (p > 0.05), hence were not discussed in detail.

In our study, 46 (55.4%) isolates were positive for the iceA2 gene and 37 (44.6%) isolates were positive for the iceA1 gene. Chomvarin et al. [41] who determined the association between the various genotypes of H. pylori and its clinical manifestations in Thai dyspeptic patients, reported that an overall 45.5% and 33.1% of the samples were positive for the *iceA1* and *iceA2* genes, respectively. Of this 35% and 40% of the gastric ulcer samples were positive for iceA1 and iceA2, respectively, which is lower than that reported in our study [41]. A study from Saudi Arabia which determined the prevalence of *cagA* and *iceA* genes from biopsy samples of patients with both gastric and peptic ulcers reported that 81.7% of the samples were positive for the iceA gene, of which 92.3% were from peptic ulcer and 75% were from gastritis cases [48]. We found that a majority of our isolates were positive for any one of the tested virulence genes indicating that these isolates are highly virulent in nature. As reported earlier, the presence of vacA and cagA genes is associated with GC, the high prevalence of these genes among our isolates may warrant a monitoring for GC in patients from whom these strains are isolated.

Conclusions

The H. pylori infection rate in our study was 48.3% and was associated with patients who were smokers or had a history of smoking. The vacA gene (68.7%) was the predominant virulence gene detected and was associated with male patients and patients within the age group of 31–40 years. The *cagA* gene was the second most common gene detected and was significantly associated with alcoholic patients. As reported earlier, the predominance of this gene warrants a close monitoring for GC in patients from whom these strains were isolated. The majority of our isolates were positive for any one of the virulence genes tested indicating that these isolates were highly virulent in nature.

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References

- 1. Bibi F, Alvi SA, Sawan SA, Yasir M, Sawan A, Jiman-Fatani AA, et al. Detection and genotyping of Helicobacter pylori among gastric ulcer and cancer patients from Saudi Arabia. Pak J Med Sci 2017;33:320-4.
- 2. Salama NR, Hartung ML, Muller A. Life in the human stomach: persistence strategies of the bacterial pathogen Helicobacter pylori. Nat Rev Microbiol 2013;11:385-99.
- 3. Goh KL, Chan WK, Shiota S, Yamaoka Y. Epidemiology of Helicobacter pylori infection and public health implications. Helicobacter 2011;16(Suppl. 1):1-9.
- 4. Roberts SE, Morrison-Rees S, Samuel DG, Thorne K, Akbari A, Williams JG. Review article: the prevalence of Helicobacter pylori and the incidence of gastric cancer across Europe. Aliment Pharmacol Ther 2016;43:334-45.
- 5. Yamaoka Y, Osato MS, Sepulveda AR, Gutierrez O, Figura N, Kim JG, et al. Molecular epidemiology of Helicobacter pylori: separation of H. pylori from East Asian and non-Asian countries. Epidemiol Infect 2000;124:91-6.
- 6. Feliciano O, Gutierrez O, Valdes L, Fragoso T, Calderin AM, Valdes AE, et al. Prevalence of Helicobacter pylori vacA, cagA, and iceA genotypes in Cuban patients with upper gastrointestinal diseases. BioMed Res Int 2015;2015:753710.
- 7. Suzuki R, Shiota S, Yamaoka Y. Molecular epidemiology, population genetics, and pathogenic role of Helicobacter pylori. Infect Genet Evol 2012;12:203-13.
- 8. Kuipers EJ, Perez-Perez GI, Meuwissen SG, Blaser MJ. Helicobacter pylori and atrophic gastritis: importance of the cagA status. J Natl Cancer Inst 1995;87:1777-80.

- 9. Yong X, Tang B, Li BS, Xie R, Hu CJ, Luo G, et al. Helicobacter pylori virulence factor CagA promotes tumorigenesis of gastric cancer via multiple signaling pathways. Cell Commun Signal 2015:13:30.
- 10. Cover TL, Blanke SR. Helicobacter pylori VacA, a paradigm for toxin multifunctionality. Nat Rev Microbiol 2005;3:320-32.
- 11. Atherton JC, Cao P, Peek RM, Jr., Tummuru MK, Blaser MJ, Cover TL. Mosaicism in vacuolating cytotoxin alleles of Helicobacter pylori. Association of specific vacA types with cytotoxin production and peptic ulceration. I Biol Chem 1995:270:17771-7.
- 12. Sayehmiri F, Kiani F, Sayehmiri K, Soroush S, Asadollahi K, Alikhani MY, et al. Prevalence of cagA and vacA among Helicobacter pylori-infected patients in Iran: a systematic review and meta-analysis. J Infect Dev Ctries 2015;9:686-96.
- 13. Aghdam SM, Sardari Z, Safaralizadeh R, Bonyadi M, Abdolmohammadi R. Moghadam MS, et al. Investigation of association between oipA and iceA1/iceA2 genotypes of Helicobacter pylori and gastric cancer in Iran. Asian Pac J Cancer Prev 2014;15:8295-9.
- 14. Shiota S, Suzuki R, Yamaoka Y. The significance of virulence factors in Helicobacter pylori. J Dig Dis 2013;14:341-9.
- 15. Erdogdu C, Saribas Z, Akyon Yilmaz Y. Detection of cagA and vacA genotypes of Helicobacter pylori isolates from a university hospital in Ankara region, Turkey. Turk J Med Sci 2014;44:126-32.
- 16. Hussein NR, Mohammadi M, Talebkhan Y, Doraghi M, Letley DP, Muhammad MK, et al. Differences in virulence markers between Helicobacter pylori strains from Iraq and those from Iran: potential importance of regional differences in H. pylori-associated disease. J Clin Microbiol 2008;46:1774-9.
- 17. Yakoob J, Abid S, Abbas Z, Jafri W, Ahmad Z, Ahmed R, et al. Distribution of Helicobacter pylori virulence markers in patients with gastroduodenal diseases in Pakistan. BMC gastroenterol
- 18. Shahidi MA, Fattahi MR, Farshad S, Alborzi A. Validation of an in-house made rapid urease test kit against the commercial CLO-test in detecting Helicobacter pylori infection in the patients with gastric disorders. J Res Med Sci 2012;17:212-6.
- 19. Farshad S, Japoni A, Shahidi MA, Hosseini M, Alborzi A. An improvement in isolation and preservation of clinical strains of Helicobacter pylori. Trop Gastroenterol 2011;32:36-40.
- 20. van Doorn LJ, Figueiredo C, Sanna R, Plaisier A, Schneeberger P, de Boer W, et al. Clinical relevance of the cagA, vacA, and iceA status of Helicobacter pylori. Gastroenterology 1998;115:58-66.
- 21. Kalaf EA, Al-Khafaji ZM, Yassen NY, Al-Abbudi FA, Sadwen SN. Study of the cytoxin-associated gene a (CagA gene) in Helicobacter pylori using gastric biopsies of Iraqi patients. Saudi J Gastroenterol 2013;19:69-74.
- 22. Yamaoka Y, Kodama T, Gutierrez O, Kim JG, Kashima K, Graham DY. Relationship between Helicobacter pylori iceA, cagA, and vacA status and clinical outcome: studies in four different countries. J Clin Microbiol 1999;37:2274-9.
- 23. Aziz F, Chen X, Yang X, Yan Q. Prevalence and correlation with clinical diseases of Helicobacter pylori cagA and vacA genotype among gastric patients from Northeast China. BioMed Res Int 2014;2014:142980.
- 24. Nagy P, Johansson S, Molloy-Bland M. Systematic review of time trends in the prevalence of Helicobacter pylori infection in China and the USA. Gut Pathog 2016;8:8.

- 25. Yu X, Yang X, Yang T, Dong Q, Wang L, Feng L. Decreasing prevalence of Helicobacter pylori according to birth cohorts in urban China. Turk J Gastroenterol 2017;28:94-7.
- 26. Habib AM, Alam MJ, Rudra B, Quader MA, Al-Forkan M. Analysis of Helicobacter pylori prevalence in Chittagong, Bangladesh, based on PCR and CLO test. Microbiol Insights 2016;9:47-50.
- 27. Nimri LF, Matalka I, Bani Hani K, Ibrahim M. Helicobacter pylori genotypes identified in gastric biopsy specimens from Jordanian patients. BMC Gastroenterol 2006:6:27.
- 28. Hooi JK, Lai WY, Ng WK, Suen MM, Underwood FE, Tanyingoh D, et al. Global prevalence of Helicobacter pylori infection: systematic review and meta-analysis. Gastroenterology 2017;153:420-9.
- 29. Taj Y, Essa F, Kazmi SU, Abdullah E. Sensitivity and specificity of various diagnostic tests in the detection of Helicobacter pylori. I Coll Physicians 2003:13:90-3.
- 30. Yakoob J, Jafri W, Jafri N, Islam M, Abid S, Hamid S, et al. Prevalence of non-Helicobacter pylori duodenal ulcer in Karachi, Pakistan. World J Gastroenterol 2005;11:3562-5.
- 31. Vu C, Ng YY. Prevalence of Helicobacter pylori in peptic ulcer disease in a Singapore hospital. Singapore Med J 2000;41:478-81.
- 32. Khasag O, Boldbaatar G, Tegshee T, Duger D, Dashdorj A, Uchida T, et al. The prevalence of Helicobacter pylori infection and other risk factors among Mongolian dyspeptic patients who have a high incidence and mortality rate of gastric cancer. Gut Pathogens 2018;10:14.
- 33. Dorji D, Dendup T, Malaty HM, Wangchuk K, Yangzom D, Richter JM. Epidemiology of Helicobacter pylori in Bhutan: the role of environment and Geographic location. Helicobacter 2014;19:69-73.
- 34. Benajah DA, Lahbabi M, Alaoui S, El Rhazi K, El Abkari M, Nejjari C, et al. Prevalence of Helicobacter pylori and its recurrence after successful eradication in a developing nation (Morocco). Clin Res Hepatol Gastroenterol 2013;37:519-26.
- 35. Mathewos B, Moges B, Dagnew M. Seroprevalence and trend of Helicobacter pylori infection in Gondar University Hospital among dyspeptic patients, Gondar, North West Ethiopia. BMC Res Notes 2013;6:346.
- 36. Olokoba AB, Gashau W, Bwala S, Adamu A, Salawu FK. Helicobacter pylori infection in Nigerians with dyspepsia. Ghana Med J 2013;47:79-81.
- 37. Zhu Y, Zhou X, Wu J, Su J, Zhang G. Risk factors and prevalence of helicobacter pylori infection in persistent high incidence area

- of gastric carcinoma in Yangzhong City. Gastroenterol Res Pract 2014;2014:481365.
- 38. Pormohammad A, Ghotaslou R, Leylabadlo HE, Nasiri MJ, Dabiri H, Hashemi A. Risk of gastric cancer in association with Helicobacter pylori different virulence factors: a systematic review and meta-analysis. Microb Pathog 2018;118:214-9.
- 39. Jeyamani L, Jayarajan J, Leelakrishnan V, Swaminathan M. CagA and VacA genes of Helicobacter pylori and their clinical relevance. Indian J Pathol Microbiol 2018;61:66-9.
- 40. Pandya HB, Agravat HH, Patel JS. Prevalence of specific Helicobacter pylori cagA, vacA, iceA, ureC genotypes and its clinical relevance in the patients with acid-peptic diseases. J Clin Diagn Res 2017;11:Dc23-6.
- 41. Chomvarin C, Namwat W, Chaicumpar K, Mairiang P, Sangchan A, Sripa B, et al. Prevalence of Helicobacter pylori vacA, cagA, cagE, iceA and babA2 genotypes in Thai dyspeptic patients. Int J Infect Dis 2008;12:30-6.
- 42. Hirai I, Sasaki T, Fujimoto S, Moriyama T, Azuma T, Yamamoto Y. A method for assessment of Helicobacter pylori genotype using stool specimens. FEMS Immunol Med Microbiol 2009:56:63-6.
- 43. Kersulyte D, Mukhopadhyay AK, Velapatino B, Su W, Pan Z, Garcia C, et al. Differences in genotypes of Helicobacter pylori from different human populations. J Bacteriol 2000;182:3210-8.
- 44. Miftahussurur M, Sharma RP, Shrestha PK, Suzuki R, Uchida T, Yamaoka Y. Molecular epidemiology of Helicobacter pylori infection in Nepal: specific ancestor root. PLoS One 2015;10:e0134216.
- 45. Higashi H, Tsutsumi R, Muto S, Sugiyama T, Azuma T, Asaka M, et al. SHP-2 tyrosine phosphatase as an intracellular target of Helicobacter pylori CagA protein. Science (NY) 2002;295:683-6.
- 46. Naito M, Yamazaki T, Tsutsumi R, Higashi H, Onoe K, Yamazaki S, et al. Influence of EPIYA-repeat polymorphism on the phosphorylation-dependent biological activity of Helicobacter pylori CagA. Gastroenterology 2006;130:1181-90.
- 47. Azuma T. Helicobacter pylori CagA protein variation associated with gastric cancer in Asia. J Gastroenterol 2004;39:97-103.
- 48. Kadi RH, Halawani EM, Abdelkader H. Prevalence of H. pylori strains harbouring cagA and iceA virulence genes in Saudi patients with gastritis and peptic ulcer disease. Microbiol Discov 2014;2:2.