

Fallacious hCG β in patients with choriocarcinoma

Zweifelhafte hCG β -Sekretion bei Patientinnen mit Chorionkarzinom

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Summary:

The serum level of hCG and total hCG β (hCG plus hCG β) was measured by a manual (hCG $_1$ and β_1) and an automated (hCG $_2$ and β_2) immunometric assay in two therapy-resistant patients with choriocarcinoma (ChCA). In contrast to hCG $_1$ and hCG $_2$, the results of β_1 and β_2 differed clearly. Total hCG β was always higher than hCG. The ratio of total hCG β /hCG in patient A with initially high serum hCG ranged between 1.1 and 5.3, while that of patient B with low serum hCG increased with time ranging between 1.1 and 550. The qualitative analysis of the hCG- and hCG β -activities was performed by gel chromatography of serum samples on Sephadex-G100 and Superdex-75 during various stages of the disease, and compared to the pattern of gel chromatography of nonradioactive hCG and hCG β or serum from a pregnant woman. After gel chromatography of serum samples from the two patients with ChCA only one fraction showed hCG-activity. There was also a variety of total hCG β -activity representing little and big-hCG or hCG β . The total hCG β activity measured by β_1 and β_2 in the fractions obtained by gel chromatography of serum samples showed different patterns. The hCG-activity in the gel chromatogram of the serum samples of patient B decreased with progress of the disease, whilst the total hCG β -activities increased in the eluates.

It was concluded that the discordant measurements of total hCG β in ChCA were probably not due to hCG β but to heterogeneity in the secretory pattern of hCG- and hCG β -like substances, which also varied with time. Moreover, the total hCG β -activities measured with different methods were shown to be considerably at variance.

Keywords:

hCG β – total hCG β – choriocarcinoma – gel chromatography – serum

Zusammenfassung:

Die Serumkonzentrationen von hCG und Gesamt-hCG β (hCG+hCG β) wurde mit manuellen (hCG $_1$ und β_1) und automatisierten (hCG $_2$ und β_2) immunometrischen Verfahren an Blutproben von zwei Frauen mit einem Therapie-resistenten Chorionkarzinom bestimmt. Die Ergebnisse des hCG $_1$ und hCG $_2$ unterscheiden sich kaum voneinander, während β_1 und β_2 große Unterschiede aufwiesen. Die Konzentration des Gesamt-hCG β lag jedoch stets höher als die des hCG. Das Verhältnis von Gesamt-hCG β /hCG schwankte bei der Patientin A (mit anfänglich hohem hCG) zwischen 1,1 und 5,3, bei Patientin B (mit konstant niedrigem hCG) zwischen 1,1 und 550. Bei der letzteren nahm die Serumkonzentration von Gesamt-hCG β , nachdem es zur Metastasierung gekommen war, bei Verwendung von β_1 , vor allem aber von β_2 , stark zu, während hCG $_1$ und hCG $_2$ gleichbleibend niedrig blieb (<26 IE/l).

Eine qualitative Analyse des Gesamt-hCG β und hCG im Serum wurde mit Hilfe der Gel-Chromatographie über Sephadex-G100 und Superdex 75 während verschiedener Stadien der Erkrankung durchgeführt, und mit dem Elutionsprofil nicht-radioaktiven hCG und hCG β bzw. dem Serum einer Schwangeren verglichen. Nach der Gel-Chromatographie der Sere dieser Patientinnen wies nur eine Fraktion hCG-Aktivität auf, deren Elutionsprofil dem des authentischen hCG oder Serum einer Schwangeren glich. Dazu kam noch ein weites Spektrum von Gesamt-hCG β -Aktivität, die jedoch nicht mit hCG β identisch war. Bei der Bestimmung der Gesamt-hCG β -Aktivität mit der β_1 - und β_2 -Methode in den Eluatens ergaben sich beträchtliche Unterschiede. Der Anteil des hCG im Gel-Chromatogramm von Patientin B nahm mit der Dauer der vierjährigen Erkrankung ab, während die mit der β_1 - und β_2 -Methode darstellbaren Aktivitäten zunahmen.

Es wird gefolgert, daß die im Vergleich zu hCG höhere Konzentration von Gesamt-hCG β im Serum von Frauen mit metastasierenden Chorionkarzinomen nicht durch hCG β verursacht wird, sondern durch ein Spektrum qualitativ und quantitativ verschiedener Substanzen, die auf Grund ihrer Heterogenität von verschiedenen Testsätzen in unterschiedlicher Weise erfaßt werden.

Schlüsselwörter:

hCG β – Gesamt hCG β – Chorionkarzinom – Gel-Chromatographie – Serum

Introduction

The determination of hCG in serum for diagnosing a choriocarcinoma (ChCA) and monitoring therapy can yield confounding results, particularly when unusually low values are being found [1–3]. Discordant measurements of hCG had i.e. been obtained in a case of ChCA when two different immunometric assays were used. This was thought to be due to variations in the secretion of hCG β [4]. A falsely-low hCG β value was also obtained because of a high dose hook effect in a patient with ovarian choriocarcinoma using an immunometric assay [5]. This is, however, a technical problem which can readily be solved by validating the assay. As the secretion of free β subunit of hCG (naturally occurring hCG β) and its ratio to hCG in patients with ChCA was found to be higher than during pregnancy [6–13], it was postulated that the determination of total hCG β (hCG plus hCG β) in an additive manner would be more advantageous than that of hCG or hCG β by a competitive immunoassay.

The secretion of hCG and total hCG β was followed in 2 therapy-resistant patients with ChCA from the time of diagnosis to their demise after 7 months and 4 years, respectively. The quantitative determination of hCG and total hCG β in serum and the qualitative analysis by gel chromatography using Sephadex-G100 and Superdex-75 was carried out by means of a manual and automated immunometric assay during various stages of the disease.

Material and Methods

Patients

Patient A: the 28 years old patient was admitted as a case of emergency because of occlusion of the aorta by a chorionepithelioma, one year after a molar pregnancy had been terminated by suction evacuation. Even though intensive cytostatic therapy was carried out subsequent to initially successful surgical intervention using methotrexate and actinomycin D, the patient expired 7 months later from the sequelae of cerebral metastasis.

Patient B: 39 years old woman, Gravida 3, Para 1. The last pregnancy terminated as a spontaneous abortion 4 years ago. She was admitted to the hospital on 12 June, 1987 (Day 0) because of suspected ectopic pregnancy as the hCG-Test was positive and sonography failed to show an intrauterine chorionic vesicle. The histologic examination of the endometrial tissue obtained by curettage revealed a choriocarcinoma. After hysterectomy, hCG declined to unmeasurable levels (<1 IU/l). Two years later, an exploratory

laparotomy revealed the presence of metastases in paraaortal lymph nodes. Soon there were brain metastases, and on 7 June 1991, the patient succumbed to the disease.

Gel chromatography

The composition of hCG and hCG β in serum of these patients was characterized by gel chromatography on Sephadex-G100 and Superdex-75 (Pharmacia, Freiburg) and measurement of hCG and total hCG β in the eluates. The characteristics of the elution patterns and immunoactivities were compared to the patterns of gel chromatograms of nonradioactive authentic substances (hCG and hCG β), and of a serum sample obtained from a pregnant woman.

A column (1.6×90 cm) was packed with Sephadex-G100 ultrafine (uf) and eluted with PBS-buffer after addition of 0.1% NaN₃ and equilibrated at 4°C. The flow rate was 0.09 ml/min, and the total bed volume (V_t) was 168 ml. The Superdex-75 was a prepacked 1.6×60 column (HiLoad Superdex-75: Pharmacia, Freiburg). Elution was accelerated by means of a peristaltic pump with a flow rate of 0.85 ml/min. One ml of serum or 5 μ g hCG or hCG β were applied to the column and fractions of 1 ml collected for the determination of hCG and total hCG β (β_1 and β_2).

Chemicals

Nonradioactive hCG (immunological activity: 9000 IE/mg 1.IRP 75/537) and hCG β (chemically dissociated from hCG: containing hCG α <0.1%) were purchased from Boehringer Mannheim (Germany). Phosphate buffer saline (PBS) pH 7.4 and the proteins for calibration of the molecular weights were supplied by Serva (Heidelberg, Germany).

Immunoassay

The manual system for determination of hCG (hCG1) was a time-resolved fluoroimmunoassay (Delfia®-hCG: Pharmacia-LKB, Freiburg). The capture antibodies directed against a specific antigenic site on the β -subunit of hCG, and the europium-labelled antibodies directed against a specific antigenic site on the α -subunit were used as signal antibodies. The cross reaction to hCG β was 2.4% (probably due to contamination with hCG). The automated system for determination of hCG (hCG2) was an enzyme fluoroimmunoassay: Stratus®-hCG (Baxter, Munich) (Table 1).

The total hCG β was determined in a manual system (β_1) by means of a radioimmunoassay with oligoclonal

Table 1: Immunoassay systems used for quantitative and qualitative measurements.

analyte designation	hCG ₁	hCG ₂	total hCGβ _{β₁}	total hCGβ _{β₂}
cross reaction:				
hCG	100%	100%	100%	100%
hCGβ	2.4%	1.4%	100%	100%
LH,FSH	undetectable	undetectable	undetectable	undetectable
assay	time-resolved fluoro-immunometric	enzyme-fluoro-immunometric	radioimmunometric	microparticle enzyme-fluoro-immunometric
ligand	europium chelate	alkaline phosphatase + 4-methylumbelliferyl-phosphate	J ¹²⁵	alkaline phosphatase + 4-methylumbelliferyl-phosphate
standard range*)	0–10,000 IU/l	0–500 IU/l	0–1,000 IU/l	0–1,000 IU/l
sensitivity	<1 IU/l	2 IU/l	1.5 IU/l	2 IU/l
interassay CV**)	7.8%	6%	6%	6.5%
system	manual	automated	manual	automated
kit	Delfia®-hCG	Stratus®-hCG	hCG + β-IRMA	Total β-hCG IMx
supplier	Pharmacia	Baxter	Medgenix	Abbott

**) high-dose hook effect in this range not observed.

**) CV = coefficient of variation.

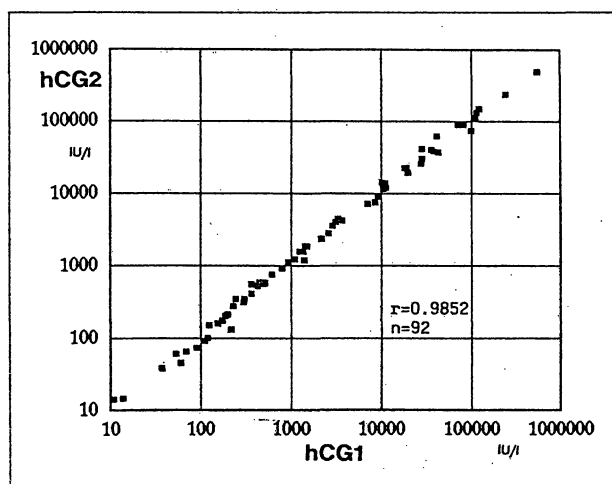


Figure 1: Concentration of hCG in serum samples from pregnant women and patients with chorioncarcinoma measured by a manual system (Delfia®-hCG = hCG₁) and an automated system (Stratus®-hCG = hCG₂).

antibodies (hCG+β-IRMA: Medgenix, Düsseldorf). The capture antibodies used for coating the tubes had a low affinity. Addition of J¹²⁵-labelled antibody (signal antibody) triggered the immunological reaction. The automated total hCGβ (β₂) was a microparticle enzyme immunometric-assay (Total hCGβ: Abbott-IMx, Wiesbaden) (Table 1).

The results of the measurements were expressed in IU/l hCG, calibrated against the 3rd International Standard WHO (IS no. 75/537), previously known as the 1st International Reference Preparation WHO (IRP no. 75/537). The characteristics of the kits used are listed in Table 1.

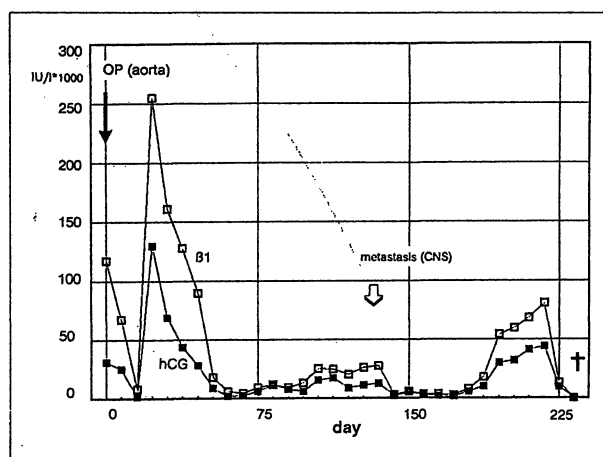


Figure 2: Quantitative measurements of hCG and total hCGβ (β₂-assay) in serum of patient A with initially high serum hCG during a period of 7 months.

Results

The measurement of hCG with the manual (hCG₁) and automated (hCG₂) system in 92 serum samples of patients with ChCA and pregnant women with a concentration ranging between 10–550000 IU/l showed a high degree of correlation (coefficient of correlation $r = 0.9853$) and did not differ significantly (paired t-test: $t = 0.342$, $p > 0.05$) (Fig. 1).

Follow-up of quantitative measurements in serum

Patient A: As expected the serum level of total hCGβ (β₂) was higher than that of hCG. After the chorionepithelioma had been removed from the aorta, three courses of treat-

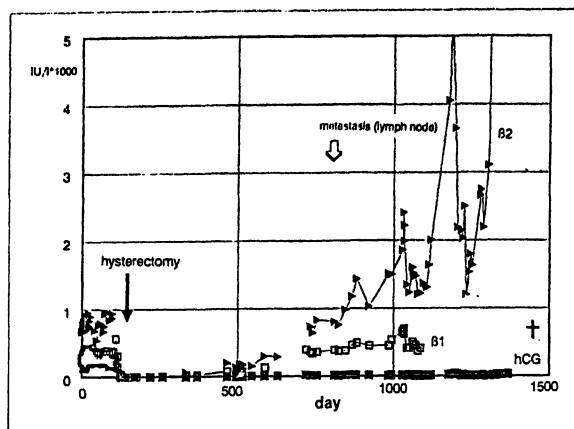


Figure 3: Quantitative measurements of hCG, total hCG β (β_1 -, β_2 -assay) in serum of patient B with permanently low serum hCG during a period of 4 years.

ment with methotrexate were carried out. This brought about just a temporary reduction of hCG and total hCG β (Fig. 2). The concentration of hCG in serum correlated well with that of total hCG β ($r = 0.96455$, $n = 32$, $p < 0.001$). It is shown in Fig. 2 that the serum level of hCG varied between 1,500 and 250,000 IU/l during the following 7 months of observation. The total hCG β and hCG ratio varied between 1.1 and 5.3.

Patient B: During the first four months of observation prior to hysterectomy, the serum level of hCG and total hCG β fluctuated within a relatively narrow range:

hCG	130–260 IU/l
total hCG β (β_1)	250–420 IU/l
total hCG β (β_2)	670–950 IU/l

The serum concentration of total hCG β both in the β_1 - and β_2 -assay was higher than that of hCG, the β_2 exceeding the level of either one. After hysterectomy, a rapid decline in the concentration of hCG and total hCG β could be shown by all three methods (Fig. 3). Three months later, hCG, and hCG β rose minimally to levels ranging between 5 and 26 IU/l and remained at that until the end of the 4 years period of follow-up. In contrast, the level of total hCG β in the β_1 and β_2 -assay increased markedly for 2 years, the latter reaching a level of more than 10,000 IU/l during the final stage of the disease. The total hCG β (β_1 and β_2)/hCG ratio fluctuated before hysterectomy between 1.1 and 5.9. It increased rapidly to a value of 550:1 after lymphatic metastasis had occurred. There was a highly significant correlation between the level of β_1 and β_2 ($r = 0.9329$, $n = 58$), but neither one did correlate with hCG.

Qualitative analysis to characterize the composition of hCG and hCG β

Authentic substances and serum from a pregnant woman

Gel chromatography of nonradioactive hCG over Sephadex-G100 showed one main peak which was eluted in

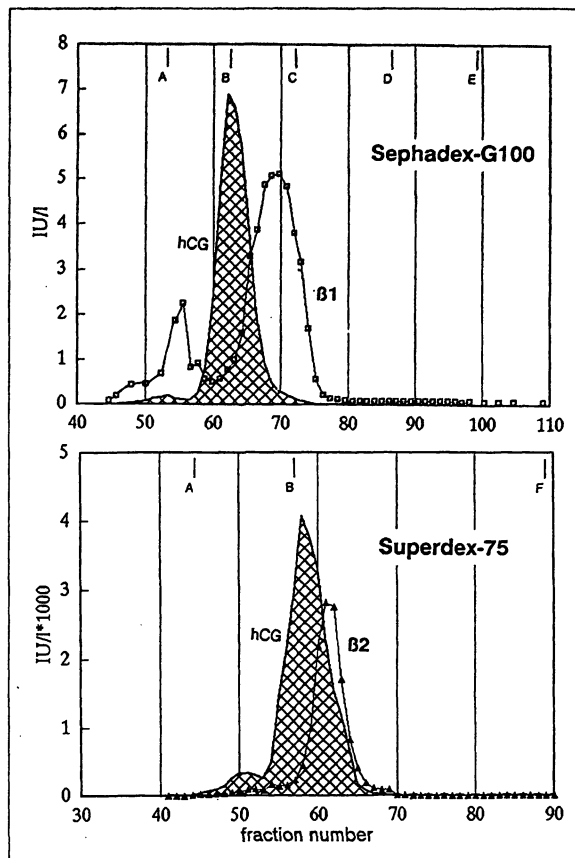


Figure 4: Elution profile of nonradioactive hCG and standard hCG β after gel chromatography on Sephadex-G100 ultrafine: column = 1.6×86 cm (flow rate 0.09 ml/min) (upper panel) and Hiload Superdex-75 column = 1.6×60 cm (flow rate 0.85 ml/min) (lower panel). PBS buffer pH 7.4 was used for elution. One milliliter of fractions were collected and the immunoactivity of hCG and total hCG β (β_1 - and β_2 -assay) were measured in the eluates. The lines above the versals indicate the peak of the fraction the calibrators for molecular weight (MW) were eluted: A = Ferritin (480,000), B = bovine serum albumin (67,000), C = Ovalbumin (43,000), D = Chymotrypsin (25,000), E = Ribonuclease A (13,700) and F = Cytochrom C (12,300).

fraction no. 62 and over Superdex-75 in fraction no. 57 when hCG was measured by means of the hCG-assay (Fig. 4). A small fraction with hCG-immunoactivity corresponding to a larger molecular weight (big-hCG) was also present both after gel chromatography over Sephadex-G100 or Superdex-75.

After gel chromatography of nonradioactive hCG β over Sephadex-G100 or Superdex-75, the peak of the main fraction was detected in fraction no. 70 and 61, respectively, when assayed by the β_1 or β_2 . The elution profile of hCG β over Sephadex-G100 showed another fraction with a peak in fraction no. 55 when assayed with the β_1 -kit. This fraction is probably similar to the large molecular form found in placental extracts, which showed cross reaction to antibodies to hCG and its subunits, but could not be

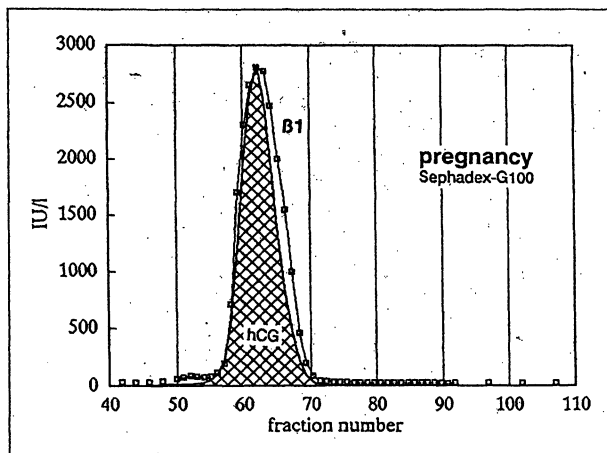


Figure 5: Elution profile of serum from a normal pregnant woman (11th week of pregnancy) after gel chromatography on Sephadex-G100, and the activity of hCG and total hCG β (β_1 -assay) in the fractions (see legend Fig. 4).

dissociated to subunits [14]. The elution profile after gel chromatography of serum from a pregnant woman on Sephadex-G100 showed only one main peak in fraction no. 62, when the samples were examined by both the hCG- and the β_1 -kit (Fig. 5).

Patient A: The fraction with hCG activity in the eluates after gel chromatography of serum from day 89 and day 198 on Sephadex-G100 or Superdex-75 showed a peak in fraction no. 62 and 58 respectively (Fig. 6). In contrast to this, the total β_1 and β_2 -activities in the eluates showed a broad spectrum, which was due to the presence of a cohort of smaller and larger sized hCG-like moieties (ca. 30,000 and 100,000 daltons). The qualitative analysis of the eluates with the β_1 - and β_2 -assay showed a rather different pattern in detection of the hCG-like moieties in the gel chromatography of serum from day 198.

Patient B: The fraction with hCG activity in the eluates after gel chromatography of serum day 33 on Sephadex-G100 showed a peak in fraction no. 62. There was minimal hCG-activity in serum obtained on day 370 but none in serum from day 1329 (Fig. 7). In contrast to this, the pattern of hCG-moieties detected with the β_1 - and β_2 -kit in the eluates after gel chromatography of serum from day 33, day 370 or day 1329 showed a different and broader spectrum. The cohort of substances detected by β_1 - and β_2 -kit represented smaller and larger sized hCG or hCG β -moieties (20,000–100,000 daltons). The total hCG β -moieties after gel chromatography detected by the β_1 - and β_2 -kit showed a qualitative different pattern.

Discussion

It could be shown, that the concentration of total hCG β measured by 2 different methods (β_1 and β_2) was higher than that of hCG in serum of the patients with ChCA. The higher ratio of total hCG β /hCG was, however, not to be

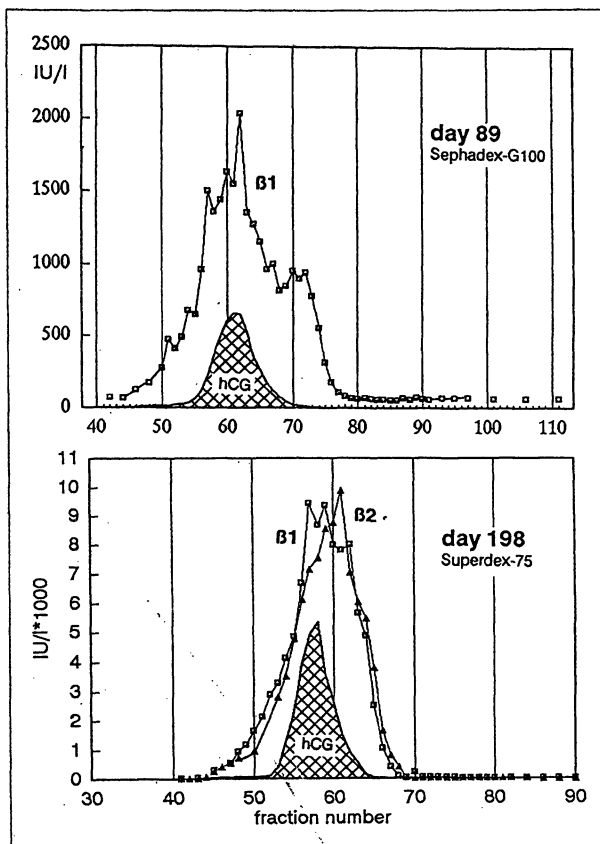


Figure 6: Elution profile of serum from patient A on day 89 (Sephadex-G100, upper panel) and on day 198 (Superdex-75, lower panel), and the activity of hCG and total hCG β (β_1 - and β_2 -assay) in the fractions (see legend Fig. 4).

caused by a higher production rate of hCG β , but by the presence of a certain number of other hCG-moieties. There was a quantitative and qualitative difference discernible in the concentrations of the hCG and hCG β species depending on the type (β_1 and β_2) of assay being used. Contrary to that, no difference between hCG1 and hCG2 could be shown.

It has become customary in clinical practice to use the measurement of hCG in the diagnosis of early pregnancy, and that of the free β -subunit (hCG β) as a tumor-marker. A competitive radioimmunoassay (RIA) using anti-hCG β and 125 I-labelled hCG or hCG β , can detect either free hCG β or intact hCG. This method is, however, not suited for the additive measurement of hCG and hCG β . The immunometric assay with mixed monoclonal antibodies is a specific and sensitive method for determination of hCG [15] or hCG β [16]. Falsely-positive or falsely-negative results have, however, been observed in an immunometric assay for hCG β in the presence of high levels of hCG [17]. On the other hand, the sum of hCG plus hCG β can be measured quantitatively using a sophisticated mixture of monoclonal antibodies as it is available in the β_1 and β_2 -assay.

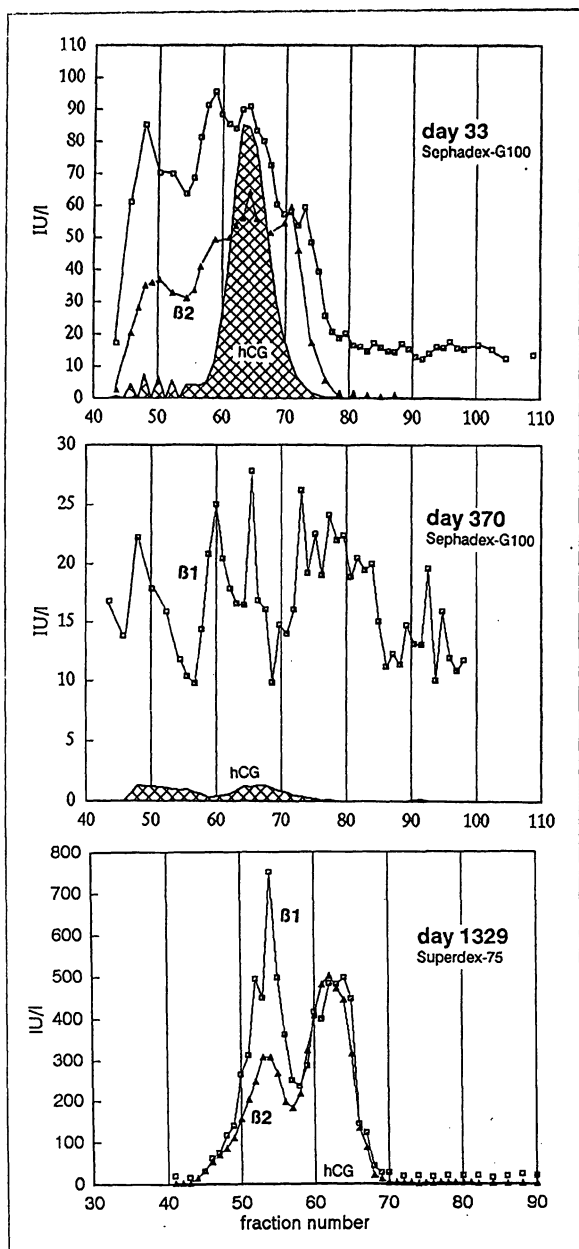


Figure 7: Elution profile of serum from patient B after gel chromatography on day 33 and day 370 (Sephadex-G100) and on day 1329 (Superdex-75). The immunoactivity of hCG, total hCG β (β_1 - and β_2 -assay) were measured in the fractions (see legend Fig. 4).

The minute amounts of authentic hCG found in the blood and urine of healthy and nonpregnant women detected by immunometric assays, represent a physiologic phenomenon of unknown relevance. It appears that the genome for hCG is capable of expressing itself outside the trophoblast in various organs such as the pituitary [18], liver, kidney etc. [19–21] and also in some malignant tumours during all phases of life. In postmenopausal women who

produce physiologically large amounts of FSH and LH, the secretion of hCG into urine is also somewhat higher than in menstruating subjects or men [22, 23]. For diagnostic purposes, a cut-off level of 5 IU/l is commonly used to minimize the risk of "false-positive" results. Another inherent technical problem of immunometric assays is the occurrence of false low levels of hCG caused by the high dose hook effect [5]. It can easily be overcome in manual systems by diluting the sample and in automated systems by a built-in computerized break.

There is growing evidence that the hCG-species produced by the placenta differ to a great deal from those secreted by a choriocarcinoma. Discordant results were e.g. obtained in a case of ChCA, when an immunometric assay for hCG and total hCG β was used [4]. In contrast, no significant difference was found when serum hCG was measured during pregnancy with an immunometric assay for hCG or total hCG β [24]. The discrepancy was probably due to the higher production rate of the naturally occurring hCG β in ChCA than during pregnancy, resulting in a higher ratio of hCG β /hCG [6–13]. Nevertheless, immunochemical staining of the tumor for hCG β in the patient with positive total hCG β in serum was found to be negative [4]. The biological activity of hCG in serum of patients with ChCA was relatively higher than the immunological activity as compared to that during pregnancy [13]. This indicates that hCG β produced by the tumor was not identical with standard hCG β (chemically dissociated). Moreover, there was a striking difference in the prevalence of larger molecular species of hCG, and a higher acidic charge in serum than in urine from a patient with ChCA [25]. The molecular heterogeneity of hCG and its subunits was also apparent when plasma and other biological fluids such as ascites, urine, and tumor extracts from oncologic patients were analyzed [26]. Apart from differences in molecular size [25, 27], median charge [28], and amino-acid sequence [29], there were particular quantitative and qualitative differences in the composition of the sugar chains not only in the N-linked but also in the O-linked oligosaccharide chain as in hCG obtained from ChCA and pregnant individuals [30, 31, 32]. Similarly, the naturally occurring hCG α from normal pregnant women was found to be different than the chemically dissociated hCG α [33–35].

The results of the present study corroborate the observations of others [13, 26, 27, 31, 32, 34, 36] that the difference between total hCG β and hCG from ChCA is not identical with chemically dissociated hCG β , but represents a whole cohort of hCG and hCG β moieties without a constant pattern. This emphasizes the fact that there are considerable qualitative and quantitative difference between the kits provided for the measurement of total hCG β . Finally, as postulated by Hay [37], the ratio of "hCG β "/hCG seems to increase with the advent of the disease, although the concentration of hCG remain quite low (Patient B).

References

1. Bakri, Y., Lee, Y. H., Jahshan, A. E., Lewis, G. C. (1982) Uterine choriocarcinoma with negative specific serum radioimmunoassay for human chorionic gonadotropins. *Gynecol. Oncol.* 14, 112-118.
2. Lemonnier, M. C., Glezerman, M., Vauclair, R., Audet-Lapointe, P. (1986) Choriocarcinoma associated with undetectable levels of human chorionic gonadotropin. *Gynecol. Oncol.* 25, 48-52.
3. O'Reilly, S. M., Rustin, G. J. S. (1993) Mismanagement of choriocarcinoma due to a false low hCG measurement. *Int. J. Gynecol. Cancer* 3, 186-188.
4. Remaley, A. T., Senior, M. B., Goodman, D. B. P. (1989) Discordant hCG measurements in a patient with a carcinoma. *Clin. Chem.* 35, 1259.
5. Wheeler, C. A., Davis, S., Degefu, S., Thorneycroft, I. H., O'Quinn, A. G. (1990) Ovarian choriocarcinoma: A difficult diagnosis of an unusual tumor and a review of the hook effect. *Obstet. Gynecol.* 75, 547-549.
6. Vaitukaitis, J. L., Ebersole, E. R. (1976) Evidence for altered synthesis of human chorionic gonadotropin in gestational trophoblastic tumors. *J. Clin. Endocrinol. Metab.* 42, 1048-1055.
7. Dawood, M. Y., Saxena, B. B., Landesman, R. (1977) Human chorionic gonadotropin and its subunits in hydatidiform mole and choriocarcinoma. *Obstet. Gynecol.* 50, 172-181.
8. Gaspard, U. J., Reuter, A. M., Deville, J. L., Vrindts-Gevaert, Y., Bagshawe, K. D., Franchimont, P. (1980) Serum concentration of human chorionic gonadotropin and its alpha and beta subunits. 2. Trophoblastic tumours. *Clin. Endocrinol.* 13, 319-329.
9. Cole, L. A., Hartle, R. J., Laferla, J. J., Ruddon, R. W. (1983) Detection of the free beta subunit of human chorionic gonadotropin (hCG) in cultures of normal and malignant trophoblast cells, pregnancy sera, and sera of patients with choriocarcinoma. *Endocrinology* 113, 1176-78.
10. Shapiro, A. I., Wu, T. F., Ballon, S. C., Lamb, E. J. (1985) Use of an immunoradiometric assay and a radioimmunoassay for the detection of free β -human chorionic gonadotropin. *Obstet. Gynecol.* 65, 545-549.
11. Fan, C., Goto, S., Furuhashi, Y., Tomoda, Y. (1987) Radioimmunoassay of serum free β -subunit of human chorionic gonadotropin in trophoblastic disease. *J. Clin. Endocrinol. Metab.* 64, 313-318.
12. Ozturk, M., Berkowitz, R., Goldstein, D., Bellet, D., Wands, J. R. (1988) Differential production of human chorionic gonadotropin and free subunits in gestational trophoblastic disease. *Am. J. Obstet. Gynecol.* 158, 193-198.
13. Wide, L., Hobson, B. (1987) Some qualitative differences of hCG in serum from early and late pregnancies and trophoblastic diseases. *Acta Endocrinol.* 116, 465-472.
14. Good, A., Ramos-Urbe, M., Ryan, R. J., Kempers, R. D. (1977) Molecular forms of human chorionic gonadotropin in serum, urine and placental extracts. *Fertil. Steril.* 28, 846-850.
15. Ehrlich, P. H., Moyle, W. R. (1983) Cooperative immunoassays: ultrasensitive assays with mixed monoclonal antibodies. *Science* 221, 279-281.
16. Bellet, D., Bidart, J. M., Jolivet, M., Tartar, A., Caillaud, J. M., Ozturk, M., Strugo, M. C., Audibert, F., Gras-Masse, H., Assicot, M., Bohuon, C. (1984) A monoclonal antibody against a synthetic peptide is specific for the free native human chorionic gonadotropin β -subunit. *Endocrinology* 115, 330-336.
17. Ozturk, M., Bellet, D., Manil, L., Hennen, G., Frydman, R., Wands, J. (1987) Physiological studies of human chorionic gonadotropin (hCG), α hCG, and β hCG as measured by specific monoclonal immunoradiometric assays. *Endocrinology* 120, 549-558.
18. Matsuura, S., Ohashi, M., Chen, H. C., Shownkeen, R. C., Stockell Hartree, A., Reichert Jr, L. E., Stevens, V. C., Powell, J. E. (1980) Physicochemical and immunological characterization of an hCG-like substance from human pituitary glands. *Nature* 286, 740-741.
19. Braunstein, G. D., Kamdar, V., Rasor, J., Swaminathan, N., Wade, M. E. (1979) *J. Clin. Endocrinol. Metab.* 58, 170-175.
20. Yoshimoto, Y., Wolfson, A. R., Hirose, F., Odell, W. D. (1979) Human chorionic gonadotropin-like material: Presence in normal human tissues. *Am. J. Obstet. Gynecol.* 134, 729-733.
21. Goldsmith, P. C., McGregor, W. G., Raymore, W. J., Kuhn, R. W., Jaffe, R. B. (1983) Cellular localization of chorionic gonadotropin in human fetal kidney and liver. *J. Clin. Endocrinol. Metab.* 57, 654-661.
22. Armstrong, E. G., Ehrlich, P. H., Birken, S., Schlatterer, J. P., Siris, E., Hembree, W. C., Canfield, R. E. (1984) Use of a highly sensitive and specific immunoradiometric assay for detection of human chorionic gonadotropin in urine of normal, nonpregnant, and pregnant individuals. *J. Clin. Endocrinol. Metab.* 59, 867-874.
23. Akar, A. H., Gervasi, G., Blacker, C., Wehmann, R. E., Blithe, D. L., Nisula, B. C. (1990) Human chorionic gonadotrophin-like and β -core-like materials in postmenopausal urine. *J. Endocrinol.* 125, 477-484.
24. Dericks-Tan, J. S. E. (1990) Sekretion von hCG β in der Schwangerschaft: Ein Artefakt? *Geburtsh. Frauenheilk.* 50, 439-442.
25. Nishimura, R., Kitajima, T., Hasegawa, K., Takeuchi, K., Mochizuki, M. (1989) Molecular forms of human chorionic gonadotropin in choriocarcinoma serum and urine. *Jpn. J. Cancer Res.* 80, 968-974.
26. Hattori, M., Yoshimoto, Y., Matsukura, S., Fujita, T. (1980) Qualitative and quantitative analyses of human chorionic gonadotropin and its subunits produced by malignant tumors. *Cancer* 46, 355-361.
27. Nishimura, R., Utsunomiya, T., Ide, K., Kitajima, T., Chen, H. C., Strobel, J. L., Hussa, R. O., Mochizuki, M. (1987) Characterization of human chorionic gonadotropin in urine of patients with trophoblastic diseases by Western blotting using specific antibodies. *Jpn. J. Cancer Res. (Gann)* 78, 833-839.
28. Yazaki, K., Yazaki, C., Wakabayashi, K., Igarashi, M. (1980) Isoelectric heterogeneity of human chorionic gonadotropin: presence of choriocarcinoma specific components. *Am. J. Obstet. Gynecol.* 138, 189-194.
29. Nishimura, R., Shin, J. T., Ji, L., Middaugh, C. R., Kruggel, W., Lewis, R. V., Ji, T. H. (1986) A single amino-acid substitution in an ectopic α -subunit of a human carcinoma choriogonadotropin. *J. Biol. Chem.* 261, 10475-10477.
30. Mizuochi, T., Nishimura, R., Derappe, C., Taniguchi, T., Hamamoto, T., Mochizuki, M., Kobata, A. (1983) Structures of the asparagine-linked sugar chains of human chorionic gonadotropin produced in choriocarcinoma. *J. Biol. Chem.* 258, 14126-14129.
31. Mizuochi, T., Nishimura, R., Taniguchi, T., Utsunomiya, T., Mochizuki, M., Derappe, C., Kobata, A. (1985) Comparison of carbohydrate structure between human chorionic gonadotropin present in urine of patients with trophoblastic diseases and healthy individuals. *Jpn. J. Cancer Res. (Gann)* 76, 752-759.
32. Cole, L. A. (1987) The O-linked oligosaccharide structure are striking different on pregnancy and choriocarcinoma HCG. *J. Clin. Endocrinol. Metab.* 65, 811-813.
33. Fein, H. G., Rosen, S. W., Weintraub, B. D. (1980) Increased glycosylation of serum human chorionic gonadotropin and subunits from eutopic and ectopic sources: Comparison with placental and urinary forms. *J. Clin. Endocrinol. Metab.* 50, 1111-1120.
34. Nishimura, R., Hamamoto, T., Utsunomiya, T., Mochizuki, M. (1983) Heterogeneity of free α -subunit in term placenta. *Endocrinol. Jpn.* 30, 663-669.
35. Posillico, E. G., Handwerker, S., Tyrey, L. (1985) Human chorionic gonadotropin α -subunit of normal placenta: characterization of synthesis and association with β -subunit. *Biol. Reprod.* 32, 1101-1108.
36. Nishimura, R., Ide, K., Utsunomiya, T., Kitajima, T., Yuki, Y., Mochizuki, M. (1988) Fragmentation of the β -subunit of human chorionic gonadotropin produced by choriocarcinoma. *Endocrinology* 123, 420-425.
37. Hay, D. L. (1988) Histological origins of discordant chorionic gonadotropin secretion in malignancy. *J. Clin. Endocrinol. Metab.* 66, 557-564.

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