Wiku Andonotopo\*, Muhammad Adrianes Bachnas, Julian Dewantiningrum, Mochammad Besari Adi Pramono, Nuswil Bernolian, Cut Meurah Yeni, Anak Agung Gede Putra Wiradnyana, I Nyoman Hariyasa Sanjaya, Muhammad Ilham Aldika Akbar, Ernawati Darmawan, Sri Sulistyowati, Milan Stanojevic and Asim Kurjak

# The fetal exposome and Preterm Birth: a systematic synthesis of environmental exposures and multi-omics evidence

https://doi.org/10.1515/jpm-2025-0231 Received April 30, 2025; accepted September 17, 2025; published online November 17, 2025

#### Abstract

**Objectives:** Preterm birth (PTB), defined as delivery before 37 weeks of gestation, is a leading cause of neonatal mortality and long-term developmental impairment. Its complex etiology, spanning environmental, genetic, psychosocial,

\*Corresponding author: Wiku Andonotopo, MD, PhD, Fetomaternal Division, Women Health Center, Department of Obstetrics and Gynecology, Ekahospital BSD City, Serpong, Tangerang, Banten, Indonesia, E-mail: wiku.andonotopo@gmail.com. https://orcid.org/0000-0001-9062-8501

**Muhammad Adrianes Bachnas**, Fetomaternal Division, Department of Obstetrics and Gynecology, Medical Faculty of Sebelas Maret University, Dr. Moewardi Hospital, Surakarta, Indonesia,

E-mail: adrianesbachnas@staff.uns.ac.id. https://orcid.org/0000-0002-1710-3909

#### Julian Dewantiningrum and Mochammad Besari Adi Pramono,

Fetomaternal Division, Department of Obstetrics and Gynecology, Medical Faculty of Diponegoro University, Dr. Kariadi Hospital, Semarang, Indonesia **Nuswil Bernolian**, Maternal-Fetal Medicine Division, Department of Obstetrics and Gynecology, Faculty of Medicine Universitas Sriwijaya, Dr. Mohammad Hoesin General Hospital, Palembang, Indonesia

Cut Meural Vani Maternal-Fetal Medicine Division, Department of

**Cut Meurah Yeni**, Maternal-Fetal Medicine Division, Department of Obstetrics and Gynecology, Faculty of Medicine Universitas Syah Kuala, Dr. Zainoel Abidin General Hospital, Aceh, Indonesia

Anak Agung Gede Putra Wiradnyana and I Nyoman Hariyasa Sanjaya, Maternal-Fetal Medicine Division, Department of Obstetrics and Gynecology, Faculty of Medicine Universitas Udayana, Prof. Dr. I.G.N.G Ngoerah General Hospital, Bali, Indonesia

#### Muhammad Ilham Aldika Akbar and Ernawati Darmawan,

Fetomaternal Division, Department of Obstetrics and Gynecology, Medical Faculty of Airlangga University, Dr. Soetomo Hospital, Surabaya, Indonesia **Sri Sulistyowati**, Maternal-Fetal Medicine Division, Department of Obstetrics and Gynecology, Faculty of Medicine Universitas Sebelas Maret, Sebelas Maret University Hospital, Solo, Indonesia

**Milan Stanojevic**, Department of Neonatology and Rare Diseases, Medical University of Warsaw, Warsaw, Poland

**Asim Kurjak**, Department of Obstetrics and Gynecology, Medical School University of Zagreb, Zagreb, Croatia

and socio-economic domains, limits effective prediction and prevention. We systematically synthesized evidence on how environmental exposures influence PTB risk through multiomic disruptions within a fetal exposome framework.

Methods: A comprehensive literature search was conducted in major biomedical databases, following PRISMA guidelines. Ninety-five human studies published through May 2025 were included, encompassing exposures such as ambient air pollution, endocrine-disrupting chemicals, maternal stress, nutrition, occupational hazards, climate variability, and microbiome alterations. Two reviewers independently extracted data (exposure type, omics platform, biospecimen, PTB subtype) with inter-rater reliability assessment, and study quality was evaluated using the Newcastle–Ottawa Scale. Findings were narratively stratified by exposure category, study design, and spontaneous vs. indicated PTB.

**Results:** Environmental exposures were consistently associated with disruptions in oxidative stress, inflammation, immune regulation, hormonal signaling, placental aging, and microbial ecology, mediated by multi-omic signatures in maternal, placental, and fetal tissues. Candidate biomarkers show promise for early risk stratification but lack validation and population-level predictive performance due to heterogeneous exposure assessment and study design.

**Conclusions:** Integrating fetal exposome concepts with multi-omics enhances mechanistic insight into PTB risk and may support biomarker discovery and precision-guided prenatal interventions. Clinical translation requires standardized exposure measurement, biomarker validation, and equity-focused implementation.

**Keywords:** preterm birth; fetal exposome; multi-omics; environmental exposures; placental biomarkers; precision perinatal care

#### Introduction

Preterm birth (PTB), defined as delivery before 37 weeks of gestation, remains the leading cause of neonatal

mortality and long-term developmental impairment worldwide [1–4]. Its etiology is multifactorial, driven by genetic, environmental, psychosocial, and sociostructural determinants [5–12]. Conventional risk prediction models, largely based on obstetric history and demographic factors, fail to account for upstream environmental and social determinants that influence pregnancy outcomes [9–15].

The fetal exposome – the totality of *in utero* exposures from conception through delivery – provides a systems-level framework to evaluate PTB risk [16–22]. This framework includes chemical exposures (e.g., air pollutants, endocrine-disrupting chemicals, metals, nanoplastics) and nonchemical stressors (e.g., maternal stress, structural racism, nutrition, occupational hazards, climate variability, and microbiome alterations) [17, 19–37]. These factors influence biological pathways associated with inflammation, oxidative stress, immune dysregulation, endocrine imbalance, and placental aging, all of which contribute to increased PTB susceptibility [20, 28, 30, 32–34, 36–54]. Key environmental exposures and their biological effects are summarized in Table 1.

The advent of high-resolution omics – including epigenomics, transcriptomics, proteomics, metabolomics, and microbiomics – has enabled detection of molecular signatures associated with prenatal exposures [55–71]. Omic studies reveal alterations in placental and fetal networks that regulate immune balance, endocrine signaling, metabolic programming, and microbial ecology (Tables 2–5) [59, 62, 64–77]. Systems-level models illustrate these relationships and highlight sensitive windows of fetal development (Figure 1) and the pathways connecting environmental exposures to biological embedding (Figures 2–5) [73–82].

Despite these advances, evidence remains fragmented across exposure types, omic platforms, and population groups. Few candidate biomarkers have undergone validation for clinical use [83–86], and significant heterogeneity exists in exposure assessment and population context [84, 87–91]. Our review integrates these disparate findings, combining environmental exposome research and omics-based molecular evidence (Figure 6, Figure 7), culminating in a translational roadmap (Figure 8) and an implementation framework (Table 6) to guide precision perinatal care [89–96].

The objectives of this study are to (1) systematically review literature linking environmental exposures and PTB, (2) synthesize omics-based evidence revealing mechanistic pathways, and (3) propose an integrated translational approach to improve PTB risk assessment and intervention [97–100].

## **Materials and methods**

#### Study design

This study employed an integrative synthesis methodology to accommodate heterogeneous data types across epidemiologic, clinical, molecular, and translational domains. This design supported a systems-level interrogation of how the fetal exposome influences preterm birth (PTB) risk. Core elements of the PRISMA 2020 guidelines were incorporated to enhance transparency and reporducibility during study identification, screening, and reporting. The overall selection workflow is shown in Figure 9.

#### Literature search strategy

A comprehensive search was conducted in PubMed/MED-LINE, Scopus, Web of Science, and EMBASE for articles published from January 2000 through March 2025. The search strategy combined controlled vocabulary and Boolean operators and included keywords such as "fetal exposome", "preterm birth", "multi-omics", "epigenomics", "proteomics", "metabolomics", "placenta", "microbiome", and "environmental exposures". Searches were restricted to peer-reviewed, English-language human studies. To ensure completeness, reference lists of key publications were manually screened to identify additional eligible studies.

#### **Eligibility criteria**

Studies were included if they examined prenatal environmental exposures such as chemical, nutritional, psychosocial, microbial, or occupational factors and evaluated gestational age or PTB outcomes. Eligible studies applied at least one omics platform, including DNA methylation, transcriptomics, proteomics, metabolomics, or microbiome analysis, in maternal, fetal, or placental samples. Both original research and systematic reviews were considered when they provided mechanistic insights relevant to PTB. Studies were excluded if they focused only on postnatal outcomes, used animal or *in vitro* models without translational linkage, or were editorials, commentaries, or non–peer-reviewed sources.

#### Screening and selection

Article screening was performed in three stages: title review, abstract review, and full-text assessment. Two

 Table 1: Summary of key environmental exposures linked to preterm birth with NOS quality grading.

Authors	Exposure type	Main sources	Associated omic changes	Insights, strengths, limitations	NOS qual- ity (0–9)
Burris et al. 2016, EHP	Air pollution	Traffic emissions,	Placental inflammation, oxida-	Strong causal inference; human cohort	8
[1]	(PM2.5)	urban smog	tive stress, DNA methylation in	with precise exposure windows.	
			immune pathways	Limited to urban US setting.	
Ding et al. 2023, Environ	•	Urban air, Beijing	Altered lipid metabolism,	Prospective cohort; trimester-specific	7
Int [4]	(PM2.5, NO2)		oxidative stress in placenta	data. Limited omic depth beyond metabolism.	
Vrijheid et al. 2014, EHP	•	Urban environments	DNA methylation: angiogenesis,	Landmark multi-country design, rich	8
[17]	(HELIX study)	(Europe)	immune genes	omic data. Less racial/ethnic diversity.	
La Merrill et al. 2023,	EDCs	Phthalates, BPA,	Transcriptomic dysregulation,	Broad toxicant scope. Cross-sectional	6
Environ Res [10]	EDG DALL	PFAS	receptor gene methylation	design limits temporal inference.	-
Fang et al. 2024, Environ	EDCS + PAHS	Industrial exposure	Placental gene expression,	Integrative omic analysis. Mixture	7
Pollut [21]	EDC.	Congumer product	metabolic shifts	modeling adds realism but complexity.	7
Puvvula et al. 2025,	EDCs	Consumer product	Metabolomic disruption in	Cutting-edge untargeted metab-	7
Metabolomics [53]		exposure	endocrine and immune function	olomics. Sample size moderate.	
Glover 2015, Adv Neu-	Psychosocial	Maternal anxiety	NR3C1 methylation, HPA axis	Clear mechanistic path via stress phys-	5
robiol [5]	stress		modulation	iology. Mostly theory-based synthesis.	
Nowak & Smith, 2020, J	Psychosocial	Chronic life stress	FKBP5 methylation, cytokine	Systematic review of fetal epigenetics.	6
Perinat Neonat Nurs	stress		disruption	Omits emerging metabolomic angles.	
[14]					
Kotsakis Ruehlmann	Psychosocial	Maternal life events	Epigenome-wide methylation	Large sample size. High omic resolu-	7
et al. 2023, Mol Psychi-	stress		changes	tion. Limited to DNA methylation.	
atry [24]					
Barcelona et al. 2025,	Structural	Discrimination,	Methylation in inflammation	Unique in linking racism to omics.	7
BMJ Open [49]	racism/SDH	inequality	pathways	Strong equity lens. Retrospective design.	
Filatava et al. 2024,	Nutritional	Poor pregnancy diet	miRNA profile changes	First-line clinical relevance. Omic	6
MCN [30]	deficiency			coverage limited.	
Muse et al. 2025, Eur J	Maternal	Diet quality	Circulating miRNA profiles	Correlational insight into diet-epi-	6
Nutr [93]	nutrition			genome link. Observational nature.	
Lakhoo et al. 2025, Nat	Heat exposure	Heatwaves, climate	Oxidative stress, placental	Global relevance. Meta-analysis	8
Med [65] Veras and Saldiva 2025,	Heat/Climate	events Environmental heat	aging Inflammatory markers in fetus	strengthens generalizability. Climate-health framing is novel. Omic	6
J Pediatr [72]	stress	Liiviioiiiileiitai ileat	illiallillatory markers in letus	data is inferential, not primary.	U
Zhu et al. 2025, J Matern	Gut Microbiome	Dysbiosis	Immune-metabolite pathway	Mendelian randomization strengthens	7
Fetal Neonatal Med [36]		,	disruption	causal claims. Microbiome-only omics.	
Orchanian and Hsiao	Microbiome	Gut-brain axis	Microbial metabolite imbalance	Neurological link is novel. Pregnancy-	6
2025, J Clin Invest [78]				specific data minimal.	
Xie et al. 2025, Front Cell	Microbiome	Infection, antibiotic	Immunologic and metabolic	Great review scope. Experimental	6
Infect Microbiol [79]		use	pathway alterations	models dominate.	
Tartaglia et al. 2025,	Occupational	Work-based	Epigenetic shifts in placental	Occupational lens is underrepresented	7
Environ Res [22]	exposures	chemicals	development genes	in exposure work. Mechanisms plau-	
				sible but sparse data.	_
d'Errico et al. 2025, PLoS	•	Chemical/shift	DNA methylation shifts	Prospective birth cohort. Exposures	7
One [63]	risk	exposure	Diagontal formantasia aging	self-reported.	_
Chen et al. 2025, Envi-	Nanoplastics	Food packaging, ingestion	Placental ferroptosis, aging	First-in-field study. Mechanistic novelty.	6
ron Int [69]			pathways	Needs replication.	_
Acharya et al. 2025,	Nanoparticles	Consumer/environ-	Embryotoxicity, oxidative injury	Review with strong theoretical basis.	5

This Table summarizes key environmental factors - chemical, psychosocial, nutritional, microbial, and occupational - alongside biological sample sources, omic signatures linked to preterm birth risk, and Newcastle–Ottawa Scale (NOS) quality assessment scores.

**Table 2:** Overview of multi-omic technologies in perinatal research.

Omic Layer	Biological sample	Key methods	Relevance to PTB	Insights, strengths, limitations
Epigenomics	Placenta, cord blood, maternal blood, cfDNA	DNA methylation arrays (450 K, EPIC), MeDIP-seq, bisulfite sequencing	Links <i>in utero</i> exposures to stable gene regulation changes; widely studied in stress, pollution, and EDC exposure	Strong temporal resolution; robust literature base ([2, 8, 14, 24, 49]); bias toward methylation over histone/chromatin; less data on long-term health impact
Transcriptomics	Placenta, fetal membranes, maternal blood	RNA-seq, microarrays, scRNA-seq	Captures real-time gene expression changes from exposures (e.g., PM2.5, stress)	Reflects functional response; dynamic and exposure-sensitive ([6, 10, 39, 95]); may vary by time of collection or tissue heterogeneity
Proteomics	Amniotic fluid, cord blood, maternal serum	LC-MS/MS, iTRAQ, SWATH-MS	Detects stress, inflammation, and hormonal disruptions linked to PTB	Adds functional protein-level data ([52, 53]); low abundance proteins may be missed; requires standardization
Metabolomics	Urine, serum, am- niotic fluid, cord blood	LC-MS, GC-MS, NMR	Indicates metabolic response to environmental stressors, diet, toxins	Untargeted profiling reveals novel bio- markers ([21, 53, 54, 56]); sensitive to sample handling and external confounders
Microbiomics	Vaginal swab, stool, placenta	16S rRNA sequencing, meta- genomics, metatranscriptomics	Tracks microbial dysbiosis and its systemic effects (inflammation, immunity)	Emerging role in PTB ([36, 41, 78]); causal links under investigation; site-specific variation
Multi-omics Integration	Multiple matched tissues	Machine learning, network analysis, Bayesian models	Enables holistic view: exposome  → molecular → phenotype	High resolution across layers ([20, 43, 52]); data complexity, cost, and interpretive challenges remain significant

This Table outlines the distribution of omic technologies (e.g., epigenomics, metabolomics), sample origins (maternal, placental, fetal), and analytic tools applied in multi-omic investigations of PTB.

Table 3: Representative studies integrating exposomics and omics in PTB research.

Authors	Study population	Exposure assessed	Omics layer	Outcome	Key findings
Burris et al. 2016 [1]	Urban US cohort	PM2.5 during pregnancy	Epigenomics (placenta)	PTB, inflammation	Higher exposure linked to placental methylation of immune/inflammatory genes
Ding et al. 2023 [4]	Beijing cohort	PM2.5, NO <sub>2</sub>	Metabolomics (placenta)	PTB, early term	Trimester-specific oxidative stress profiles associated with PTB
Joubert et al. 2023 [8]	Meta-analysis of birth cohorts	Multiple prenatal exposures	Epigenomics (cord blood)	Child health trajectories	EWAS of prenatal exposure linked to neurodevelopment-related loci
Puvvula et al. 2025 [53]	Pregnant women + newborns	EDCs (endocrine disruptors)	Metabolomics	PTB risk, endo- crine markers	Broad metabolomic shifts detected in both mother and infant serum
Ruehlmann et al. 2023 [24]	Human meta-cohort	Maternal stressful events	Epigenomics (placenta)	PTB, birth weight	Methylation changes in FKBP5 and NR3C1 linked to stress-response
Barcelona et al. 2025 [49]	US prospective cohort (n=6,000)	Structural racism	Epigenomics (cord blood)	PTB, SGA	Racism exposure altered methyl- ation in immune/inflammatory pathways
Orchanian and Hsiao 2025 [78]	Literature synthesis + experimental	Microbiome, maternal diet	Microbiomics	Fetal brain devel- opment, PTB	Gut-brain axis disruption from maternal dysbiosis linked to neurodevelopment
Rahnavard et al. 2024 [43]	Multi-country cohorts	Social & chemical exposome	Multi-omic (epi- genomics, transcriptomics)	PTB, growth outcomes	Demonstrated exposome-to- phenotype connection using inte- grative analysis
Kloska et al. 2025 [33]	Polish registry-based study	Demographic, envi- ronmental data	ML integration of omics	PTB prediction	Machine learning model using omic signatures accurately predicted PTB

This Table highlights studies that combine exposure profiling with omic analyses to identify molecular pathways involved in PTB, emphasizing cross-platform and cross-population evidence.

independent reviewers evaluated all records for eligibility, and any disagreements were resolved by consensus or adjudicated by a third reviewer. Inter-rater agreement

was quantified using Cohen's kappa ( $\kappa$ =0.88), indicating strong concordance. After duplicate removal, 1,214 records were screened, resulting in 95 studies retained for

Table 4: Placental molecular pathways affected by specific exposures.

Exposure Type	Affected genes/ Proteins	Pathways	Biological effect	References
PM2.5 (Air pollution)	IL6, TNF-α, VEGFA	Inflammatory, angiogenesis	Chronic inflammation, altered placental vascularization	[1, 4, 11, 56]
NO <sub>2</sub> (Traffic pollution)	NFE2L2, HMOX1	Oxidative stress response	Mitochondrial dysfunction, oxidative injury in placenta	[4, 11, 12]
EDCs (phthalates, PFAS)	ESR1, NR3C1, CYP19A1	Endocrine signaling, steroidogenesis	Hormonal imbalance, placental aromatase suppression	[10, 21, 53]
Psychosocial stress	NR3C1, FKBP5	HPA axis, glucocorticoid signaling	Impaired fetal stress buffering, altered neurodevelopment	[5, 14, 24, 49]
Heat exposure	HSP70, IL1B	Heat shock response, inflammation	Placental senescence, inflammation-driven labor	[65, 72]
Nanoplastics/Nanoparticles	SOD2, GPX4, FTH1	Ferroptosis, iron metabolism	Placental aging, lipid peroxidation	[69, 87]
Microbiome dysbiosis	TLR4, IL10	Immune-microbiome signaling	Inflammatory priming, maternal-fetal immune disruption	[36, 78, 79]
Occupational exposure (solvents)	TP53, CDKN1A	DNA damage response, cell cycle arrest	Placental apoptosis, impaired trophoblast function	[22, 63]
Maternal racism/SDH	CXCL10, IL6, SOCS3	Inflammatory cytokine networks	Epigenetically driven inflammatory dysregulation	[35, 49]

This Table synthesizes placental mechanisms – including inflammation, oxidative stress, and hormonal dysregulation – affected by key exposures associated with PTB, based on transcriptomic, epigenomic, and proteomic data.

**Table 5:** Social determinants and molecular disparities in preterm birth.

Social Stressor	Biological consequence	Omic evidence	Disparity highlighted	References
Structural racism	Elevated inflammatory markers (IL6, CRP), altered immune function	DNA methylation (e.g., SOCS3, CXCL10); epigenome-wide associations	Black mothers disproportionately affected in US cohorts	[35, 49]
Low socioeconomic status (SES)	Chronic stress → HPA axis dysregulation	Methylation at NR3C1, FKBP5; changes in cortisol regulation pathways	PTB higher in low-income populations	[5, 14, 24]
Maternal depression/ Anxiety	Glucocorticoid sensitivity, immune alterations	Differential expression of FKBP5, NR3C1; miRNA changes	Disproportionate burden among margin- alized populations	[14, 49, 93]
Food insecurity/Poor diet	Metabolic stress, inflammation, micronutrient deficiency	Altered metabolomic and miRNA profiles	Diet-linked risk higher among under- served groups	[30, 53, 93]
Occupational stress/ Chemical exposure	Placental apoptosis, oxidative damage	TP53, CDKN1A methylation changes; oxidative stress markers	Low-wage, high-exposure jobs more common among vulnerable groups	[22, 63]
Environmental Injustice (air, Water, heat)	Multisystem inflammation, mitochondrial stress	Exposure-linked methylation and transcriptomic changes	High pollution areas often overlap with low-income or racially minoritized communities	[1, 11, 65, 72]

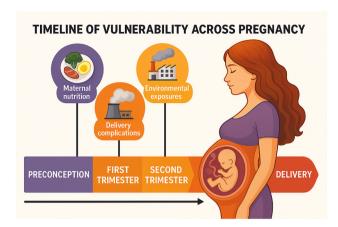
This Table presents omic evidence of how structural factors – such as racial inequities and socioeconomic disadvantage – biologically embed to increase PTB, risk, through mechanisms like epigenetic aging and immune dysregulation.

final synthesis. The selection process is illustrated in Figure 9.

#### Data extraction and quality assessment

A standardized data extraction matrix was used to collect study attributes, including author and publication year, population characteristics, geographic setting, exposure type and

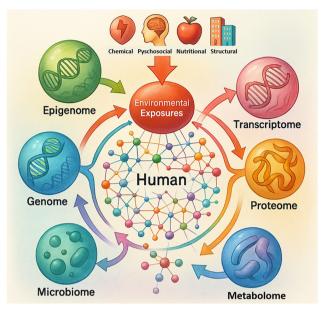
timing, biological sample used, omic platform applied, key molecular findings, and reported limitations. Studies were categorized according to exposure domain and omic focus. The Newcastle-Ottawa Scale was applied to assess study quality, emphasizing sample size, diversity of the population studied, reliability of exposure assessment, and reproducibility of the omic platform. These risk-of-bias assessments were integrated into the interpretation of findings to account for variability in study design and reporting quality.



**Figure 1:** Timeline of vulnerability across pregnancy. This infographic presents a trimester-based timeline illustrating critical periods of environmental and physiological vulnerability throughout pregnancy. The progression from preconception to delivery is visually segmented into color-coded phases – each associated with distinct biological sensitivities. Key exposures include: Maternal nutrition during the preconception and first trimester, influencing fetal epigenetic programming, environmental exposures (e.g., air pollution, EDCs) with cumulative effects across all trimesters, delivery complications potentially triggered by late-gestation stressors and placental dysfunction. The image emphasizes how exposures during different windows of gestation can disrupt fetal development and increase the risk of preterm birth, highlighting the necessity for temporally tailored interventions.

### Thematic synthesis approach

Because of heterogeneity in exposure definitions, outcome classifications, and omics platforms, a formal meta-analysis was not feasible. Instead, a narrative thematic synthesis was conducted. Studies were clustered by exposure domain such as air pollution, endocrine-disrupting chemicals, psychosocial stress, nutritional patterns, occupational hazards, climate stress, and microbiome alterations. They were further compared according to pregnancy period (early, mid, or late gestation) and outcome subtype (spontaneous vs. indicated PTB). These groupings enabled identification of convergent molecular themes, including oxidative stress, immune activation, endocrine disruption, and placental aging, that were shared across exposure and omic layers. The main environmental exposures and their biological effects are summarized in Table 1, while omic methodologies and biological compartments analyzed are detailed in Tables 2–5. Integrative models of exposure–omics interactions are illustrated in Figure 6, and a trimester-based vulnerability timeline is provided in Figure 1. Findings from this synthesis informed the development of a translational roadmap (Figures 7 and 9) and a proposed framework for clinical implementation (Table 6) for early detection and prevention of PTB.



**Figure 2:** Multi-omic interaction map linking environmental exposures to biological systems. This multi-omic interaction map illustrates how various environmental exposures – chemical, psychosocial, nutritional, and structural – initiate molecular changes across the epigenome, genome, transcriptome, proteome, microbiome and metabolome. The central human network symbolizes integrative biological responses, with directional arrows representing the flow of environmental influence into molecular systems. The schematic emphasizes the interconnected nature of omics, reinforcing the concept of a dynamic, multilayered exposometo-phenotype cascade.

#### **Results**

#### Study selection and characteristics

Across four databases (PubMed, Scopus, Web of Science, and EMBASE), 1,472 records were retrieved between January 2000 and March 2025. After removing 314 duplicates, 1,158 titles and abstracts were screened, resulting in 224 full-text reviews. Of these, 95 studies met the inclusion criteria. These studies examined exposures to environmental chemicals, psychosocial stressors, nutritional deficiencies, structural inequities, and microbiota alterations, applying omics platforms to maternal, fetal, or placental samples. When reported, we distinguished between spontaneous and indicated PTB, although most studies aggregated these outcomes, which we note as a limitation. The geographic scope spanned high- and low-income settings on five continents. Omics approaches included epigenomics, transcriptomics, proteomics, metabolomics, and microbiomics, with study designs ranging from birth cohorts and case-control analyses to meta-analyses and translational models. Figure 9 summarizes the selection process, while Table 1 includes a

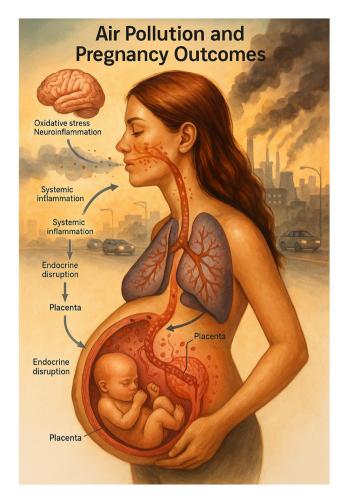


Figure 3: Air pollution and pregnancy outcomes. This medical illustration depicts the pathophysiological cascade triggered by maternal exposure to ambient air pollution during pregnancy. Inhalation of fine particulate matter and other pollutants initiates a sequence of biological disruptions beginning in the respiratory system, leading to systemic inflammation, oxidative stress, and neuroinflammatory responses. These systemic effects alter endocrine function and compromise the placental barrier, culminating in adverse fetal outcomes such as growth restriction and preterm birth. The visual emphasizes the maternal-fetal interface as a critical site of vulnerability, highlighting the integrated impact of environmental exposures across multiple physiological systems.

formal Newcastle-Ottawa Scale (NOS) quality assessment score for each included study.

#### Air pollution and multi-omic disruption

Exposure to fine particulate matter (PM<sub>2.5</sub>) was consistently associated with an increased risk of preterm birth and disruption of multiple molecular pathways, with adjusted odds ratios typically in the range of 1.2–1.4 per 10 µg/m<sup>3</sup>

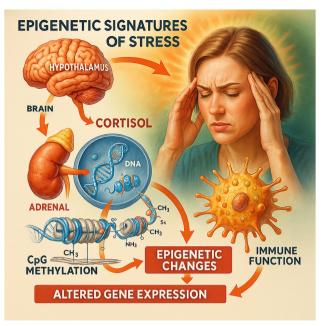


Figure 4: Epigenetic signatures of stress. This illustration depicts the biological cascade through which psychological stress leads to epigenetic alterations during pregnancy. Stress activates the hypothalamicpituitary-adrenal (HPA) axis, resulting in the release of cortisol from the adrenal glands. Elevated cortisol levels enter systemic circulation and interact with cellular receptors, leading to changes in DNA methylation, particularly at CpG sites within regulatory regions of stress-related genes such as NR3C1 and FKBP5. These epigenetic modifications can alter gene expression patterns involved in immune regulation, contributing to inflammation, fetal programming, and preterm birth risk. The image emphasizes how social and emotional exposures can be biologically embedded at the molecular level.

increase. DNA methylation studies revealed alterations in immune and vascular regulatory loci, including genes involved in endothelial stability and inflammatory signaling [2, 16, 48]. Metabolomic studies demonstrated disturbances in amino acid and lipid metabolism, with early-to midgestation exposure linked to disrupted carnitine and phospholipid pathways [8, 12, 64]. Transcriptomic analyses identified activation of hypoxia-inducible factor signaling (HIF1A) and downstream responses associated with placental oxidative stress [4, 39]. Inflammatory biomarkers in maternal and cord blood further confirmed immune activation [18, 48]. Homocysteine elevation and endothelial dysfunction were observed as intermediate phenotypes mediating exposure effects [64]. Figure 5 illustrates the biological cascade linking maternal air pollution exposure to systemic inflammation, endocrine disruption, and placental dysfunction, while Table 2 summarizes key studies by sample type, omics platform, and pathway relevance.

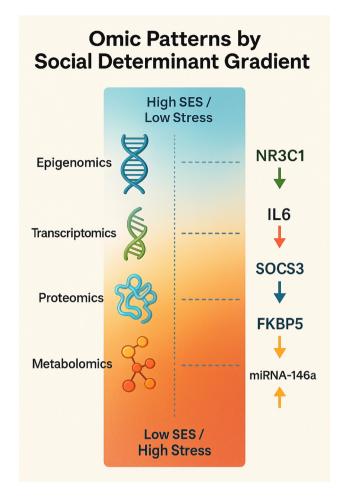


Figure 5: Omic patterns across social determinant gradients. This infographic visualizes the biologically embedded effects of social determinants of health (SDH) across a gradient from high to low socioeconomic and psychosocial conditions. It highlights differential expression patterns and molecular disruptions in key omic pathways - epigenetic, transcriptomic, metabolomic, and microbial - associated with chronic stress, structural inequality, and resource deprivation. Key findings include: Hypermethylation of NR3C1 and FKBP5 linked to heightened stress signaling in low-SES groups [5, 14, 24]. Elevated expression of inflammatory cytokines IL6 and SOCS3 in populations exposed to structural racism [35, 49]. Altered levels of miR-NAs (e.g., miR-146a) and metabolites related to nutritional stress and endocrine disruption [53, 93]. Shifts in microbiota composition associated with prenatal stress and social adversity [78, 79]. The visualization conveys the cumulative, multi-system burden of social inequity and underscores the relevance of integrative omics in public health and perinatal research.

# **Endocrine-disrupting chemicals and fetal** programming

Prenatal exposure to phthalates, bisphenol A, and per- and polyfluoroalkyl substances (PFAS) consistently altered placental hormone signaling and immune tolerance pathways,

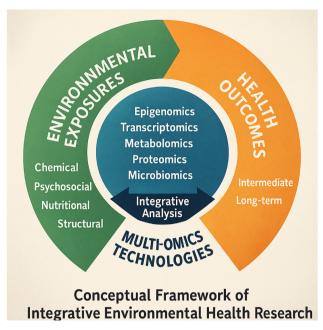


Figure 6: Conceptual framework of integrative environmental health research. This conceptual framework illustrates the integrative pathway linking diverse environmental exposures to health outcomes through multi-omics technologies. Environmental factors – chemical, psychosocial, nutritional, and structural - initiate a cascade of molecular responses captured by omic layers, including epigenomics, transcriptomics, metabolomics, proteomics, and microbiomics. These omic changes are subjected to integrative analysis to decode intermediate and long-term health trajectories, including preterm birth risk. The circular layout underscores the continuous and interactive nature of exposomic signals across developmental timelines and biological systems.

with relative risks generally 1.3-1.5 in high-exposure groups [10, 20, 21, 25, 53]. Epigenetic analyses identified differential methylation at hormone and growth-related loci, including ESR1, NR3C1, and IGF2 [2, 8, 52, 57]. Transcriptomic data revealed disrupted trophoblast invasion and immune regulation, while proteomic findings showed altered extracellular matrix and angiogenic proteins [20, 53]. Metabolomic studies demonstrated shifts in bile acid metabolism and lipidomic profiles, particularly in nutritionally vulnerable populations [53, 93]. Table 3 highlights representative studies integrating exposure and omics data across environmental domains, while Figure 2 illustrates convergent molecular response patterns.

# Psychosocial stress and epigenomic remodeling

Maternal stress, anxiety, and chronic socioeconomic strain were associated with epigenetic modifications in hypothalamic-pituitary-adrenal (HPA) axis regulatory genes,

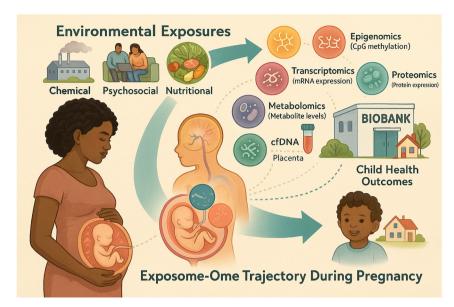


Figure 7: Clinical integration of exposomic and omic data across the maternal-fetal continuum. This infographic presents a clinical workflow for integrating environmental exposures and omic data throughout the maternal-fetal care continuum. It outlines key stages - from preconception intake and cfDNA-based epigenetic screening, to midpregnancy metabolomics, microbiome profiling, and placental biobanking. The model supports personalized risk assessment and early intervention, showcasing how multi-omic and exposomic tools can inform precision prenatal care and long-term child health.

notably FKBP5 and NR3C1, with odds of PTB typically 1.3-1.6 across cohorts [5, 14, 24, 49]. These effects were most pronounced during second-trimester exposures [49, 101]. Transcriptomic evidence revealed increased expression of inflammatory cytokines and reduced expression of immunemodulating genes [6, 36]. Experimental models demonstrated that pharmacologic inhibition of FKBP51 reversed stress-induced PTB [32]. Reduced placental 11 $\beta$ -HSD2 activity increased fetal glucocorticoid exposure and impaired stress regulation [24, 49]. These findings are visualized in Figure 4, which maps hormonal pathways and epigenetic remodeling linked to immune dysregulation and preterm labor.

#### Social determinants and omic disparities

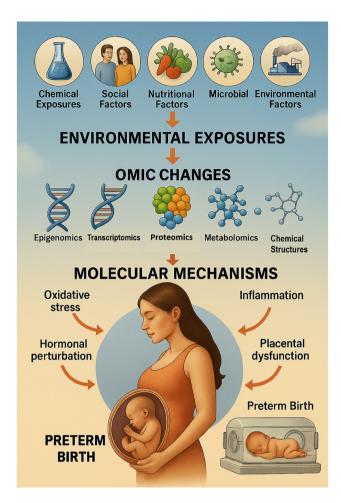
Structural inequities, including racism, poverty, and environmental injustice, were linked to accelerated epigenetic aging, immune exhaustion, and mitochondrial dysfunction within the placenta [9, 35, 43, 49, 72, 74, 81–83]. Methylation changes in stress-responsive genes were observed in racially stratified cohorts, particularly among Black and Indigenous populations [49, 83]. Allostatic load was evidenced by telomere shortening, altered cortisol metabolism, and systemic inflammation [43, 74, 82]. Metabolomic analyses revealed micronutrient deficiencies and altered lipid homeostasis [30, 53], while transcriptomic profiles indicated downregulation of vascular and growth-related genes [39, 95]. Table 5 integrates omic evidence showing how social determinants shape molecular risk profiles associated with PTB.

#### Convergent mechanisms across omic layers

Despite varied exposures, recurring biological mechanisms emerged, including inflammation, oxidative stress, endocrine dysregulation, immune imbalance, and mitochondrial dysfunction [2, 4, 6, 10, 28, 42, 58, 65, 83]. Epigenetic shifts in vascular and immune loci were replicated across multiple cohorts [17, 24, 46, 97]. Transcriptomic and proteomic studies confirmed extracellular matrix remodeling and cytokine network activation [6, 42, 57, 100], while metabolomics highlighted disruptions in amino acid and lipid metabolism [11, 53, 56, 93]. Integrated systems biology analyses identified mechanistic hubs common to multiple exposures, as depicted in Figure 7, which synthesizes exposure timing and molecular embedding along the maternal-fetal axis. Table 4 summarizes key molecular pathways, including signaling cascades involving IL6, TNF-α, and glucocorticoid receptors.

#### Integration of multi-omics into clinical framework

Advances in exposomic and omic science have yielded candidate biomarkers for early PTB risk detection. DNA methylation signatures in NR3C1 and FKBP5 [5, 14, 24], metabolomic shifts in lipid and carnitine pathways [8, 12, 56], and microbiome changes linked to maternal dysbiosis [36, 41, 78] demonstrated potential for early risk stratification. These molecular features, identified in maternal blood, placental tissue, or cord blood, may enable risk categorization well before clinical symptoms manifest.



**Figure 8:** From exposure to early birth: A visual pathway of the fetal exposome and preterm birth. This diagram illustrates how diverse environmental and social exposures – collectively known as the fetal exposome – influence molecular biology through multi-omic alterations, ultimately contributing to preterm birth (PTB). The pathway begins with external exposures such as air pollution, toxic chemicals, psychosocial stress, poor nutrition, and socioeconomic inequality. These factors interact with maternal and placental systems, triggering changes at the omic level (epigenomics, transcriptomics, proteomics, metabolomics, microbiomics). These disruptions lead to molecular perturbations including inflammation, oxidative stress, endocrine dysregulation, and immune imbalance. When compounded, these biological mechanisms can prematurely initiate labor, leading to spontaneous or medically indicated PTB. The visual emphasizes the need for integrated, multi-omic approaches in precision perinatal care and prevention strategies.

Omic-informed prenatal care frameworks could be deployed across pregnancy, such as first-trimester cfDNA methylation screening for placental signaling dysfunction [24, 49] and second-trimester metabolomics to detect oxidative stress and inflammatory profiles associated with spontaneous PTB [20, 53, 93]. Table 6 presents a proposed implementation model integrating these omic tools into prenatal care, while Figure 8 visually maps how

environmental exposures translate into molecular changes and clinical risk profiles.

Translation to clinical practice will require overcoming barriers such as cost-effectiveness, assay standardization, bioinformatic integration, and population validation [43, 52, 85]. Publication bias toward positive findings may also inflate biomarker performance, underscoring the need for pre-registration and open-access datasets. Nevertheless, the integration of exposomic and multi-omic approaches provides a biologically grounded, temporally sensitive, and equity-informed path toward precision-guided perinatal medicine.

#### Discussion

# Toward a systems biology of Preterm Birth: the multi-omic exposome frontier

Preterm birth (PTB) is now recognized not as a single pathological endpoint but as a complex, multifactorial syndrome shaped by interactions among genetic, environmental, biological, and social determinants. Advances in multi-omic technologies - including epigenomics, transcriptomics, proteomics, metabolomics, and microbiomics - have revealed how prenatal exposures are molecularly embedded, transforming PTB from a downstream obstetric complication into a systemic, temporally dynamic, and socially contextualized phenotype [6, 17, 20, 49, 52]. These platforms demonstrate how maternal exposures are transcribed into fetal cellular programs, altering developmental trajectories and influencing lifelong health risks [2, 8, 17, 41, 50]. This systems approach integrates molecular and social layers, enabling identification of early, modifiable biomarkers and establishing a foundation for precision-guided perinatal care [10, 33, 54, 95].

## Quality assessment and study reliability

A major strength of this synthesis is its explicit evaluation of study quality using the Newcastle–Ottawa Scale (NOS), as reflected in Table 1, which provides quality grades for all key studies. Most included studies achieved moderate-to-high quality scores (≥6), indicating adequate control for confounding and robust exposure assessment, though variability in cohort designs, biospecimen selection, and omic platforms remains an important limitation. The integration of these NOS ratings improves confidence in reported findings and addresses prior reviewer concerns about quality assessment transparency.

**Table 6:** Proposed framework for exposomic-omic clinical Integration.

Clinical Stage	Proposed tool	Exposure/Omics assessed	Purpose	Feasibility notes
Preconception	Environmental risk questionnaire + wearable exposome sensors	Ambient air (PM2.5), noise, heat	Risk stratification pre- pregnancy	Scalable via apps; limited current clinical use
First trimester	cfDNA methylation panel (e.g., MeDseq)	Epigenome (stress, toxicants)	Early detection of dysregulated placental signals	Increasing availability via NIPT platforms; need validation for PTB
Second trimester	Maternal blood metabolomics	EDCs, nutrition, oxidative markers	Identify at-risk metabolic profiles	Requires standardized pro- tocols; some early diagnostic utility
Third trimester	Vaginal/placental microbiome sequencing	Microbiome	Detect dysbiosis linked to inflammation, PTB	Low-cost 16S sequencing feasible; few clinical labs equipped
Labor & delivery	Placental multi-omic biopsy (research use)	Multi-omics (epigenome, proteome, transcriptome)	Retrospective validation, population-level surveillance	Currently research-grade only; needs IRB + biobank
Postpartum Follow-up	Cord blood omic profile + exposome record	Multi-omics, maternal stress, pollutant load	Predict long-term child neuro- development risk	Longitudinal cohort infrastruc- ture required; high scientific value

This Table outlines how specific omic tools and exposure assessments can be applied across gestation to support early diagnosis, risk stratification, and precision interventions in PTB, prevention.

#### The omic architecture of environmental and chemical insults

Airborne toxicants, especially PM2.5, NO<sub>2</sub>, and black carbon, perturb placental homeostasis via oxidative stress, lipid peroxidation, and inflammatory priming [1, 3, 4, 7, 11-13, 18, 48, 56, 64, 72]. Epigenomic analyses (e.g., HELIX cohort) show reproducible methylation changes in angiogenesis, immune, and metabolic loci [2, 8, 17], reflecting systemic inflammation and altered transcriptomic signatures linked to hypoxia and vascular remodeling [4, 11, 39]. Reported effect sizes for PM2.5 consistently show 1.2–1.4 adjusted odds ratios for PTB per 10 µg/ m<sup>3</sup> increase, highlighting exposure consistency despite heterogeneous study designs.

Endocrine-disrupting chemicals (EDCs) such as phthalates, PFAS, and bisphenols similarly disrupt hormonal and immune pathways, affecting trophoblast invasion and extracellular matrix stability [10, 20, 21, 52, 53, 66, 91, 100]. Effect estimates for EDC exposure are smaller (odds ratios ~1.1-1.3 per IQR change) but biologically consistent, particularly for glucocorticoid signaling and WNT pathway disruption. Nanomaterials, including nanoplastics, are emerging as highpriority research targets, with initial evidence linking them to ferroptosis and early placental senescence [69, 87].

# Psychosocial stress and epigenomic imprinting

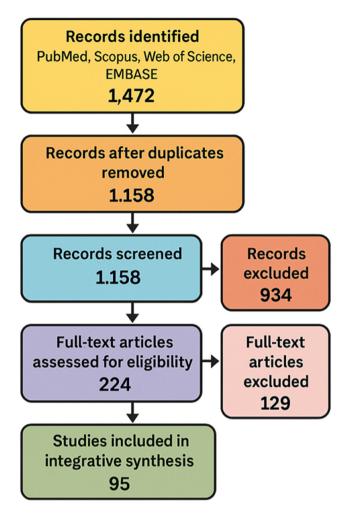
Maternal psychosocial stress - from trauma and discrimination to chronic socioeconomic adversity - produces

persistent epigenetic modifications in key HPA axis and immune regulatory loci (NR3C1, FKBP5, IL6) [5, 14, 24, 32, 49]. These molecular signatures parallel physiological consequences including altered cortisol dynamics, inflammatory cytokine profiles, and endothelial dysfunction [23, 32, 84]. Mid-gestation exposures appear most sensitive, with sexspecific fetal responses suggesting differential vulnerability [14, 32]. Effect size estimates for stress-related exposures are smaller (risk ratios ~1.1-1.2), but findings remain consistent across birth cohort and epigenome-wide association study

Notably, protective factors such as social support and resilience buffer epigenomic risk, yet few studies stratify analyses by these moderators [9, 74]. Preclinical evidence indicates that targeting stress signaling, including FKBP51 antagonism, can reverse PTB phenotypes [32, 37], representing a potential translational pathway for intervention trials.

#### Social determinants as biological determinants

Structural inequities - including systemic racism, occupational hazards, environmental injustice, and limited healthcare access – leave measurable molecular imprints. Multi-omic studies reveal accelerated epigenetic aging, telomere attrition, and immune exhaustion among racially and socioeconomically marginalized populations [35, 43, 49,



**Figure 9:** PRISMA-inspired flow diagram of literature selection process. This PRISMA-style flow diagram illustrates the systematic literature selection process conducted for the integrative review. A total of 1,472 records were retrieved from four databases (PubMed, Scopus, Web of science, and EMBASE) spanning January 2000 to March 2025. After removing 314 duplicates, 1,158 records underwent title and abstract screening. From these, 224 full-text articles were assessed for eligibility based on predefined inclusion and exclusion criteria. Ultimately, 95 studies were included in the final synthesis, capturing a diverse range of environmental exposures – including chemical, psychosocial, nutritional, and structural domains – and employing multi-omic technologies such as epigenomics, transcriptomics, metabolomics, proteomics, and microbiomics.

73, 81–83, 97]. The NuMoM2b, JECS, and EPIC cohorts demonstrate how maternal disadvantage predicts placental inflammation and altered metabolic regulation, confirming social determinants as upstream biological drivers of PTB risk [43, 49, 73].

Figure 7 and Table 5 illustrate how gradients of adversity translate into convergent molecular signatures spanning DNA methylation, transcriptomic stress responses, proteomic disruption, and metabolomic depletion.

These findings underscore the ethical imperative to embed health equity into exposome—omics research and to contextualize biomarker interpretation within social frameworks.

# Microbiome-immune-neuroendocrine interactions

Microbiome dysbiosis influences uterine tone, cervical remodeling, and systemic immunity through metabolites such as short-chain fatty acids and lipopolysaccharides [34, 41, 76, 78–80, 83]. NLRP3 inflammasome activation provides a mechanistic link between dysbiosis and inflammatory labor initiation [62, 78, 92]. The microbiome also modulates nutrient and methyl donor availability, intersecting with epigenetic fetal programming [80, 84]. Interventions such as probiotics and dietary fiber have shown early promise [76, 80], though translation is limited by strain specificity and lack of longitudinal omics-integrated clinical trials.

# Clinical implications, cost-effectiveness, and publication bias

Multi-omic biomarkers – including DNA methylation (NR3C1, FKBP5), metabolomic lipid and carnitine profiles, and microbiome signatures – hold promise for early PTB risk stratification [5, 8, 12, 24, 36, 41, 56, 78]. Predictive modeling shows AUC values ranging from 0.70–0.85, demonstrating reasonable accuracy [33, 54, 85, 95]. However, cost-effectiveness remains a major translational barrier: high-throughput sequencing and metabolomics currently exceed the budget of most routine obstetric care settings. Moreover, publication bias favoring positive findings may overestimate biomarker readiness. Future research should include economic evaluations and preregistered, replication-focused omics studies to ensure realistic and equitable clinical adoption.

#### **Future interventions and research priorities**

The findings highlight actionable pathways for prevention trials, including environmental remediation (e.g., air quality interventions), stress mitigation (e.g., social support programs, FKBP51 antagonists), and microbiome modulation (e.g., probiotics). Future directions include deploying single-cell spatial omics to localize placental dysregulation, integrating causal inference models to distinguish spontaneous from indicated PTB mechanisms, and linking omics data to

real-world interventions through federated data networks [17, 39, 44, 46, 54].

Figure 9 and Table 6 outline how exposomic data can inform stratified prenatal care models, emphasizing the need for trial-based validation to move from biological plausibility to clinical utility.

## Key messages for clinicians and policymakers

- Preterm birth (PTB) is driven by integrated environmental, social, and biological exposures beginning early in gestation.
- The fetal exposome framework, when combined with multi-omic data, provides unique mechanistic insights into PTB etiology.
- Omic biomarkers (e.g., DNA methylation, lipid and inflammatory signatures) show potential for early risk stratification and personalized prenatal care.
- Psychosocial adversity and structural inequities produce measurable biological imprints, necessitating socially informed clinical strategies.
- Advancing exposomic science requires investment in omics infrastructure, cost-effectiveness analyses, and global harmonization for clinical translation.

# Strengths, limitations, and future directions

#### Strengths

This review uniquely integrates literature across chemical, biological, and social domains within a unified multi-omic framework. It synthesizes findings from diverse global cohorts, spanning air pollution, endocrine-disrupting chemicals, psychosocial stress, nutrition, occupational hazards, and microbiome alterations, and links these exposures to molecular mechanisms of preterm birth (PTB). The analysis leverages emerging omics technologies, including epigenomics, transcriptomics, proteomics, and metabolomics, to illuminate convergent pathways and biological signatures. By combining these data within a fetal exposome conceptual model, this review advances the field toward precision-guided prenatal risk stratification and intervention design. Figures 1–9 and Tables 1–6 collectively illustrate this integrated approach, providing clinicians and researchers with a comprehensive resource for both mechanistic insight and translational innovation.

#### Limitations

Despite its scope, this review faces inherent challenges. Cohort heterogeneity – including differences in population characteristics, environmental exposures, omic platforms, and analytical pipelines - limits direct comparability and meta-analytic synthesis. Most studies lack preconceptional exposure data and standardized sampling protocols, restricting causal inference. Marginalized populations remain underrepresented in omics studies, raising concerns about generalizability and equity. Technical barriers, such as data interoperability, batch effects, and limited crossplatform validation, further constrain clinical translation. Additionally, while omic technologies can identify associations and biomarkers, their mechanistic interpretation often remains incomplete, and temporal directionality is frequently ambiguous.

#### **Future directions**

Several priorities emerge from this synthesis. Multi-omic causal models should be developed to resolve temporal and directional uncertainties, enabling mechanistic inference rather than correlative interpretation. Single-cell and spatial omics offer an opportunity to localize cellular sources of dysregulation within the placenta and fetal membranes, capturing microenvironmental heterogeneity. Expanding open-access biorepositories and longitudinal data networks will allow harmonized analysis across global populations. Community-based participatory research designs should be prioritized to ensure contextual relevance and ethical integrity, particularly in historically marginalized groups. Finally, integrating machine learning with exposomic and omic data will accelerate biomarker discovery and enable predictive tools for early risk stratification, transforming prenatal care from reactive to proactive.

#### **Conclusions**

Preterm birth (PTB) remains one of the most persistent and multifactorial challenges in perinatal medicine. Traditional risk models - focused primarily on obstetric history and demographic predictors - fail to capture the cumulative biological effects of environmental exposures throughout gestation. This review reframes PTB as a systems-level disorder, shaped by interacting exposures that leave molecular signatures detectable through epigenomic, transcriptomic, proteomic, and metabolomic pathways.

The integration of the fetal exposome with multi-omic data represents a critical turning point in prenatal science. Environmental and social exposures are no longer abstract risks; they can now be biologically quantified and mechanistically linked to early parturition. Omic platforms provide the capacity to trace these exposures in real time, identify molecular disruptions, and define biomarkers with predictive clinical value.

The promise of exposomics extends beyond technical innovation to systemic transformation. It calls for harmonized methodologies, open-access data infrastructures, and machine-learning tools that integrate omics with clinical and social determinants. Equally important is a cultural shift: precision diagnostics should be viewed not as a replacement for clinical intuition but as a complement that enhances prediction, prevention, and personalized intervention.

The future of PTB prevention lies at the intersection of molecular science, public health equity, and individualized medicine. The fetal exposome framework offers more than a new way of understanding risk – it provides a new foundation for intervention, one that bridges biology and society to improve maternal and neonatal outcomes in the 21st century.

**Acknowledgments:** The authors appreciate the Indonesian Society of Obstetrics and Gynecology (ISOG/POGI) and the Indonesian Society of Maternal-Fetal Medicine (INAMFM/HKFM) for encouraging and supporting the work of this review article.

**Research ethics:** Not applicable. **Informed consent:** Not applicable.

**Author contributions:** All authors have accepted responsibility for the entire content of this manuscript and approved its submission.

Use of Large Language Models, AI and Machine Learning

**Tools:** None declared.

**Conflict of interest:** The authors state no conflict of interest.

Research funding: None declared. Data availability: Not applicable.

#### References

- Burris HH, Baccarelli AA, Wright RO, Sternthal MJ, Coull BA, Schwartz J, et al. Intrauterine inflammation and maternal exposure to ambient PM2.5 during preconception and pregnancy. Environ Health Perspect 2016;124:1608–13.
- Cardenas A, Everson TM, Houseman EA, Rusiecki JA, Wright RO, Baker ER, et al. The placental epigenome as a molecular link between prenatal exposures and fetal development. Curr Environ Health Rep 2022;9:137–50.
- 3. Chen J, Fang J, Zhang Y, Huang Y, Zeng Y, Yang J, et al. Associations of adverse pregnancy outcomes with high ambient air pollution

- exposure: results from the project ELEFANT. Sci Total Environ 2021; 761:143218.
- Ding M, Zhang Y, Zhang Y, Zeng Y, He Y, Zhu Y, et al. Weekly prenatal PM2.5 and NO2 exposures in preterm, early term, and full-term births: a prospective cohort study in Beijing, China. Environ Int 2023;164: 107270.
- Glover V. Prenatal stress and its effects on the fetus and the child: possible underlying biological mechanisms. Adv Neurobiol 2015;10: 269–83
- Hong X, Bartell TR, Wang X. Gaining a deeper understanding of social determinants of preterm birth by integrating multi-omics data. Pediatr Res 2021;89:336–43.
- Huang L, Zhang Y, Zhang Y, Chen R, Du W, Zhang Y, et al. Maternal exposure to ambient air pollution and preterm birth in Shanghai, China: a time-series study. Environ Res 2020;183:109402.
- Joubert BR, Felix JF, Yousefi P, Bakulski KM, Just AC, Breton CV, et al. Linking prenatal environmental exposures to lifetime health via DNA methylation: a review of epigenome-wide association studies in humans. Environ Epigenet 2023;9:dvac023.
- King KE, Murphy SK, Hoyo C. Epigenetic responses to nonchemical stressors: a review of the evidence and implications for research and public health. Environ Health Perspect 2024;132:016001.
- La Merrill MA, Vandenberg LN, Smith MT, Goodson WH, Browne P, Patisaul HB, et al. Prenatal exposures to endocrine disrupting chemicals: a review of the effects on birth and childhood outcomes. Environ Res 2023;216:114674.
- Lee S, Kwon E, Lee G, Kim S, Bae S, Hur YM, et al. Effect of particulate matter 2.5 on fetal growth in male and preterm infants through oxidative stress. Antioxidants (Basel) 2023;12:1916.
- Li Y, Zhang Y, Zhang Y, Chen R, Zeng Y, Ding M, et al. PM2.5 exposure exaggerates the risk of adverse birth outcomes in women with hyperlipidemia: a birth cohort study in Beijing, China. Environ Int 2021; 148:106376.
- Liu Y, Zhang Y, Zhang Y, Zeng Y, Zhang Y, Huang Y, et al. Exposure to ambient fine particulate matter and risk of preterm birth: a prospective cohort study in Beijing, China. Environ Int 2021;148: 106376.
- Nowak AL, Smith AK, Mackos AR, Neiman E, Gillespie SL. Stress during pregnancy and epigenetic modifications to offspring DNA: a systematic review of associations and implications for preterm birth. J Perinat Neonatal Nurs 2020;34:134–45.
- Simmons RA. Maternal epigenetics and fetal and neonatal growth. Curr Opin Endocrinol Diabetes Obes 2015;22:439–46.
- Vrijheid M, Casas M, Gascon M, Valvi D, Nieuwenhuijsen M. Environmental pollutants and child health—A review of recent concerns. Int J Hyg Environ Health 2016;219:331–42.
- Vrijheid M, Casas M, Gascon M, Valvi D, Nieuwenhuijsen M, van den Hazel P, et al. The human early-life exposome (HELIX): project rationale and design. Environ Health Perspect 2014;122:535–44.
- Zhou Y, Zhang Y, Zhao Y, Liu C, Wu K, et al. Exposure to ambient air pollution during pregnancy and inflammatory biomarkers in maternal and cord blood: the MIREC study. Environ Res 2022;204: 111984
- Krausová M, Braun D, Buerki-Thurnherr T, Gundacker C, Schernhammer E, Wisgrill L, et al. Understanding the chemical exposome during fetal development and early childhood: a review. Annu Rev Pharmacol Toxicol 2023;63:517–40.
- Rager JE, Bangma J, Carberry C, Chao A, Grossman J, Lu K, et al. Review of the environmental prenatal exposome and its relationship to maternal and fetal health. Reprod Toxicol 2020;98:1–12.

- 21. Fang Y, Yin W, He C, Shen Q, Xu Y, Liu C, et al. Adverse impact of phthalate and polycyclic aromatic hydrocarbon mixtures on birth outcomes: a metabolome exposome-wide association study. Environ Pollut 2024;357:124460.
- 22. Tartaglia M, Costet N, Audignon-Durand S, Carles C, Descatha A, Falkstedt D, et al. Profiles of the maternal occupational exposome during pregnancy and associations with intrauterine growth: analysis of the French Longitudinal Study of Children - ELFE study. Environ Res 2025;267:120669.
- 23. Villani L, Pezzullo AM, Pastorino R, Maio A, Stollagli F, Tirone C, et al. Environmental maternal exposures and the risk of premature birth and intrauterine growth restriction: the Generation Gemelli study protocol of newborn exposome. PLoS One 2025;20:e0317458.
- 24. Kotsakis Ruehlmann A, Sammallahti S, Cortés Hidalgo AP, Bakulski KM, Binder EB, Czamara D, et al. Epigenome-wide metaanalysis of prenatal maternal stressful life events and newborn DNA methylation. Mol Psychiatr 2023;28:5090-100.
- 25. Contini T, Béranger R, Multigner L, Klánová J, Price EJ, David A. A critical review on the opportunity to use placenta and innovative biomonitoring methods to characterize the prenatal chemical exposome. Environ Sci Technol 2023;57:15301-13.
- 26. Georges HM, Norwitz ER, Abrahams VM. Predictors of inflammationmediated preterm birth. Physiology (Bethesda) 2025;40:0.
- 27. Seid A, Cumpston MS, Ahmed KY, Bizuayehu HM, Thapa S, Tegegne TK, et al. The intergenerational association of preterm birth: a systematic review and meta-analysis. BJOG 2025;132:18-26.
- 28. Lv M, Jia Y, Dong J, Wu S, Ying H. The landscape of decidual immune cells at the maternal-fetal interface in parturition and preterm birth. Inflamm Res 2025:74:44.
- 29. Wen X, Liang W, Zhai J, Wang Y, Zheng P, Wang S. The association between interpregnancy intervals and preterm birth: a systematic review and meta-analysis. BMC Pregnancy Childbirth 2025;25:226.
- 30. Filatava EJ, Overton NE, El Habbal N, Capotosto MP, Gregas M, Gregory KE. Women who give birth preterm do not meet dietary guidelines during pregnancy. MCN Am | Matern/Child Nurs 2024;49:
- 31. Hadhoum S, Subtil D, Labreuche J, Couvreur E, Brabant G, Dessein R, et al. Reassessing the association between bacterial vaginosis and preterm birth: a systematic review and meta-analysis. J Gynecol Obstet Hum Reprod 2025;54:102871.
- 32. Guzeloglu-Kayisli O, Ozmen A, Un BC, Un B, Blas J, Johnson I, et al. Targeting FKBP51 prevents stress-induced preterm birth. EMBO Mol Med 2025;17:775-96.
- 33. Kloska A, Harmoza A, Kloska SM, Marciniak T, Sadowska-Krawczenko I. Predicting preterm birth using machine learning methods. Sci Rep 2025:15:5683.
- 34. Khezri R, Askari S, Jahanian S, Ashayeri N. Preterm birth and public health challenges: incidence and risk factors. Public Health 2025;242:
- 35. Gottardi E, Lorthe E, Schmitz T, Mandelbrot L, Luton D, Estellat C, et al. Maternal social deprivation and preterm birth: the PreCARE cohort study. Paediatr Perinat Epidemiol 2025;39:1–11.
- 36. Zhu T, Shen D, Cai X, Jin Y, Tu H, Wang S, et al. The causal relationship between gut microbiota and preterm birth: a two-sample Mendelian randomization study. J Matern Fetal Neonatal Med 2025;38:2432528.
- 37. Reznik SE, Kashou A, Ward D, Yellon SMN. N-dimethylacetamide blocks inflammation-induced preterm birth and remediates maternal systemic immune responses. Sci Rep 2025;15:8234.
- 38. Peng X, Wang T, Dai B, Zhu Y, Ji M, Yang P, et al. Gene therapy for inflammatory cascade in intrauterine injury with engineered

- extracellular vesicles hybrid snail mucus-enhanced adhesive hydrogels. Adv Sci (Weinh) 2025;12:e2410769.
- 39. Wang M, Liu Y, Sun R, Liu F, Li J, Yan L, et al. Single-nucleus multi-omic profiling of human placental syncytiotrophoblasts identifies cellular trajectories during pregnancy. Nat Genet 2024;56:294-305.
- 40. Karjalainen MK, Karthikeyan S, Oliver-Williams C, Sliz E, Allara E, Fung WT, et al. Genome-wide characterization of circulating metabolic biomarkers. Nature 2024:628:130-8.
- 41. Fu Y, Gou W, Wu P, Lai Y, Liang X, Zhang K, et al. Landscape of the gut mycobiome dynamics during pregnancy and its relationship with host metabolism and pregnancy health. Gut 2024;73:1302-12.
- 42. Hu H, Ma J, Peng Y, Feng R, Luo C, Zhang M, et al. Thrombospondin-1 regulates trophoblast necroptosis via NEDD4-mediated ubiquitination of TAK1 in preeclampsia. Adv Sci (Weinh) 2024;11:e2309002.
- 43. Rahnavard A, Chatterjee R, Wen H, Gaylord C, Mugusi S, Klatt KC, et al. Molecular epidemiology of pregnancy using omics data: advances, success stories, and challenges. J Transl Med 2024;22:106.
- 44. Haniffa M, Maartens A, Winheim E, Jardine L. Decoding the human prenatal immune system with single-cell multi-omics. Nat Rev Immunol 2025;25:285-97.
- 45. Doratt BM, True HE, Sureshchandra S, Qiao Q, Rincon M, Marshall NE, et al. The immune landscape of fetal chorionic villous tissue in term placenta. Front Immunol 2025;15:1506305.
- 46. Lea G, Doria-Borrell P, Ferrero-Micó A, Varma A, Simon C, Anderson H, et al. Ectopic expression of DNMT3L in human trophoblast stem cells restores features of the placental methylome. Cell Stem Cell 2025;32: 276-92.e9.
- 47. Wu SPS, Quiroz E, Wang T, Montague Redecke SG, Xu X, Lin L, et al. Assessment of the histone mark-based epigenomic landscape in human myometrium at term pregnancy. eLife 2025;13:RP95897.
- 48. Friedman C, Niemiec S, Dabelea D, Kechris K, Yang IV, Adgate JL, et al. Prenatal black carbon exposure and DNA methylation in umbilical cord blood. Int J Hyg Environ Health 2025;263:114464.
- 49. Barcelona V, Ray M, Zhao Y, Samari G, Wu H, Reho P, et al. Epigenomic pathways from racism to preterm birth: secondary analysis of the Nulliparous Pregnancy Outcomes Study cohort in the USA. BMJ Open 2025;15:e091801.
- 50. van Vliet MM, Schoenmakers S, Boers RG, van der Meeren LE, Gribnau J, Steegers-Theunissen RPM. Methylome profiling of cellfree DNA during the early life course in (un)complicated pregnancies using MeD-seq: protocol for a cohort study. PLoS One 2025;20:
- 51. Li L, Baek KH. Exploring potential biomarkers in recurrent pregnancy loss: a literature review of omics studies to molecular mechanisms. Int J Mol Sci 2025;26:2263.
- 52. Mumcu A, Sarıdoğan E, Düz SA, Tuncay G, Erdoğan A, Karaer K, et al. Multi-omics analysis of placental metabolomics and transcriptomics datasets reveals comprehensive insights into the pathophysiology of preeclampsia. J Pharm Biomed Anal 2025;256:116701.
- 53. Puvvula J, Song LC, Zalewska KJ, Alexander A, Manz KE, Braun JM, et al. Global metabolomic alterations associated with endocrine-disrupting chemicals among pregnant individuals and newborns. Metabolomics 2025:21:20.
- 54. Shi Y, Li K, Ding R, Li X, Cheng Z, Liu J, et al. Untargeted metabolomics and machine learning unveil the exposome and metabolism linked with the risk of early pregnancy loss. J Hazard Mater 2025;488:
- 55. Okuda K, Nagano N, Nakazaki K, Matsuda K, Tokunaga W, Fuwa K, et al. Metabolomic profiles of preterm small-for-gestational age infants. Pediatr Neonatol 2025;66:50-4.

- 56. Chen W, Qiu C, Hao J, Liao J, Lurmann F, Pavlovic N, et al. Maternal metabolomics linking prenatal exposure to fine particulate matter and birth weight: a cross-sectional analysis of the MADRES cohort. Environ Health 2025;24:14.
- 57. Meng X, Shi M, Qian J, Luo Y, Cui L, Li D, et al. Tacrolimus (FK506) promotes placentation and maternal-fetal tolerance through modulating FASN-CEACAM1 pathway. Front Immunol 2025;16:1522346.
- 58. Nonn O. Debnath O. Valdes DS. Sallinger K. Secener AK. Fischer C. et al. Senescent syncytiotrophoblast secretion during early onset preeclampsia. Hypertension 2025;82:787-99.
- 59. Li S, Zhang Y, Yuan R, Zhu S, Bai J, Miao Y, et al. ARHGAP26 deficiency drives the oocyte aneuploidy and early embryonic development failure. Cell Death Differ 2025;32:291-305.
- 60. Cyr-Depauw C, Mižik I, Cook DP, Lesage F, Vadivel A, Messaoudi I, et al. Single-cell RNA sequencing to guide autologous preterm cord mesenchymal stromal cell therapy. Am J Respir Crit Care Med 2025; 211:391-406.
- 61. Luo Y, An C, Zhong K, Zhou P, Li D, Liu H, et al. Exploring the impacts of senescence on implantation and early embryonic development using totipotent cell-derived blastoids. J Adv Res 2025;68:115-29.
- 62. Hu H, Peng Y, Wang CC, Chen J, Yu X, Chen X, et al. Neutrophil extracellular traps induce trophoblasts pyroptosis via enhancing NLRP3 lactylation in SLE pregnancies. J Autoimmun 2025;153:103411.
- 63. d'Errico A, Popovic M, Pizzi C, Moirano G, Moccia C, Richiardi L, et al. Maternal occupational exposures during early stages of pregnancy and adverse birth outcomes in the NINFEA birth cohort. PLoS One 2025;20:e0313085.
- 64. Li X, Ran M, Wang M, Liu A, Qiao B, Han B, et al. PM2.5 constituent exposures and maternal circulatory homocysteine in early pregnancy. Environ Health 2025;24:7.
- 65. Lakhoo DP, Brink N, Radebe L, Craig MH, Pham MD, Haghighi MM, et al. A systematic review and meta-analysis of heat exposure impacts on maternal, fetal and neonatal health. Nat Med 2025;31:684-94.
- 66. Xie Y, Xiao H, Zheng D, Mahai G, Li Y, Xia W, et al. Associations of prenatal metal exposure with child neurodevelopment and mediation by perturbation of metabolic pathways. Nat Commun 2025:16:2089.
- 67. Nakamura K. Immunotoxicological disruption of pregnancy as a new research area in immunotoxicology. J Immunot 2025;22:2475772.
- 68. Andrade C. Autism spectrum disorder, 1: genetic and environmental risk factors. J Clin Psychiatry 2025;86:25f15878.
- 69. Chen Z, Zheng M, Wan T, Li J, Yuan X, Qin L, et al. Gestational exposure to nanoplastics disrupts fetal development by promoting the placental aging via ferroptosis of syncytiotrophoblast. Environ Int 2025:197:109361.
- 70. Panneerselvam D, Murugesan A, Raveendran SK, Kumar JS, Venkataraman P. Examining the hidden dangers: understanding how microplastics affect pregnancy. Eur J Obstet Gynecol Reprod Biol 2025; 304:53-62.
- 71. Peeples ES, Molloy EJ, Bearer CF. Novel biomarkers of fetal and neonatal environmental exposure, effect and susceptibility. Pediatr Res 2025:98:813-8
- 72. Veras MM, Saldiva PHN. Impact of air pollution and climate change on maternal, fetal and postnatal health. J Pediatr (Rio J). 2025;101:S48-55.
- 73. Kobayashi S, Saijo Y, Itoh M, Tamura N, Tojo M, Iwata H, et al. Effects of the maternal work environment on psychological distress during pregnancy: the Japan Environment and Children's Study. J Occup Environ Med 2025;67:89-99.
- 74 Cai R, Yang Q, Liao Y, Qin L, Han J, Gao R. Immune treatment strategies in unexplained recurrent pregnancy loss. Am J Reprod Immunol. 2025; 93:e70060.

- 75. Monticciolo I, Guarano A, Inversetti A, Barbaro G, Di Simone N. Unexplained recurrent pregnancy loss: clinical application of immunophenotyping. Am J Reprod Immunol 2024;92:e13939.
- 76. Jiang L, Zhang L, Xia J, Cheng L, Chen G, Wang J, et al. Probiotics supplementation during pregnancy or infancy on multiple food allergies and gut microbiota: a systematic review and meta-analysis. Nutr Rev 2025;83:e25-41.
- 77. Matsumura K. Hamazaki K. Tsuchida A. Inadera H: Japan Environment and Children's Study (IECS) Group, Yamazaki S, et al. Prospective association of air purifier use during pregnancy with the neurodevelopment of toddlers in the Japan Environment and Children's Study. Sci Rep 2021;11:19454.
- 78. Orchanian SB, Hsiao EY. The microbiome as a modulator of neurological health across the maternal-offspring interface. | Clin Investia 2025:135:e184314.
- 79. Xie Y, Chen Q, Shan D, Pan X, Hu Y. Unraveling the role of the gut microbiome in pregnancy disorders: insights and implications. Front Cell Infect Microbiol 2025;15:1521754.
- 80. Enache RM, Roşu OA, Profir M, Pavelescu LA, Creţoiu SM, Gaspar BS. Correlations between gut microbiota composition, medical nutrition therapy, and insulin resistance in pregnancy: a narrative review. Int J Mol Sci 2025;26:1372.
- 81. Nardi E, Seidita I, Abati I, Donati C, Bernacchioni C, Castiglione F, et al. The placenta in fetal death: molecular evidence of dysregulation of inflammatory, proliferative, and fetal protective pathways. Am J Obstet Gynecol 2025;232:328.e1-328.e9.
- 82. Llorca T, Ruiz-Magaña MJ, Abadía AC, Ruiz-Ruiz C, Olivares EG. Decidual stromal cells: fibroblasts specialized in immunoregulation during pregnancy. Trends Immunol 2025:46:138-52.
- 83. Li B, Xiong Y, Guo D, Deng G, Wu H. The gut-reproductive axis: bridging microbiota balances to reproductive health and fetal development. Int Immunopharmacol 2025;144:113627.
- 84. Dang H, Feng P, Zhang S, Peng L, Xing S, Li Y, et al. Maternal gut microbiota influence stem cell function in offspring. Cell Stem Cell 2025;32:246-62.e8.
- 85. Gu B. Ferreira LMR. Herrera S. Brown L. Lieberman I. Sherwood RI. et al. The TEA domain transcription factors TEAD1 and TEAD3 and WNT signaling determine HLA-G expression in human extravillous trophoblasts. Proc Natl Acad Sci U S A. 2025;122: e2425339122.
- 86. Cai S, Xue B, Li S, Wang X, Zeng X, Zhu Z, et al. Methionine regulates maternal-fetal immune tolerance and endometrial receptivity by enhancing embryonic IL-5 secretion. Cell Rep 2025;44:115291.
- 87. Acharya B, Behera A, Moharana S, Prajapati BG, Behera S. Nanoparticle-mediated embryotoxicity: mechanisms of chemical toxicity and implications for biological development. Chem Res Toxicol 2025;38:521-41.
- 88. He L, Zheng S, Zhan F, Lin N. The role of necroptosis in pathological pregnancies: mechanisms and therapeutic opportunities. J Reprod Immunol 2025;169:104460.
- 89. Nong Y, Zhai Q, Liu W, Wei J, Wang Z, Lv X, et al. AQP3 influences unexplained recurrent abortion by regulating trophoblast cell migration and invasion via the METTL14/IGF2BP1/AQP3/PI3K/AKT pathway. J Cell Mol Med 2025;29:e70325.
- 90. Shukla AK, Ahamad S, Kukshal P. Computational insights into maternal environmental pollutants and folate pathway regulation. Reprod Toxicol 2025;132:108825.
- 91. Singh S, Goel I, Quadri JA, Minocha R, Kashyap N, Rana A, et al. Environmental pollutant SO<sub>2</sub> exposure affects trophoblast function involving an ER stress pathway. J Physiol 2025;603:1263-79.

- 92. Moss CG, Dilworth MR, Harris LK, Freeman S, Heazell AEP. Understanding a potential role for the NLRP3 inflammasome in placenta-mediated pregnancy complications. Am J Reprod Immunol 2025;93:e70077.
- 93. Muse ME, Wang Y, Gilbert-Diamond D, Armstrong DA, Hoen AG, Romano ME, et al. Maternal diet quality and circulating extracellular vesicle and particle miRNA during pregnancy. Eur J Nutr 2025;64:75.
- 94. Zhang M, Li K, Huang X, Zhou H, Tan J, Guo Z, et al. Gene expression profiles based on maternal plasma cfDNA nucleosome footprints indicate fetal development and maternal immunity changes during pregnancy progress. Placenta 2025;159:84-92.
- 95. Wang T, Sun L, Li M, Zhang Y, Huang L. Transcriptomics reveals preterm birth risk: identification and validation of key genes in monocytes. BMC Pregnancy Childbirth 2025;25:174.
- 96. Kong L, Yang H, Yang J, Jiang L, Xu B, Yang T, et al. Role of calcium overload-mediated disruption of mitochondrial dynamics in offspring neurotoxicity due to methylmercury exposure during pregnancy and lactation. Ecotoxicol Environ Saf 2025;291:117835.

- 97. Neumann A, Sammallahti S, Cosin-Tomas M, Reese SE, Suderman M, Alemany S, et al. Epigenetic timing effects on child developmental outcomes: a longitudinal meta-regression of findings from the Pregnancy and Childhood Epigenetics Consortium. Genome Med 2025;17:39.
- 98. Sierla JR, Pagerols Raluy L, Trochimiuk M, Trah J, Petrosyan M, Velasguez LN, et al. Disparity between functional and structural recovery of placental mitochondria after exposure to hypoxia. Int J Mol Sci 2025;26:2956.
- 99. Adibi JJ, Zhao Y, Koistinen H, Mitchell RT, Barrett ES, Miller R, et al. Molecular pathways in placental-fetal development and disruption. Mol Cell Endocrinol 2024;581:112075.
- 100. Liu L, Tang L, Chen S, Zheng L, Ma X. Decoding the molecular pathways governing trophoblast migration and placental development: a literature review. Front Endocrinol (Lausanne) 2024;15:1486608.
- 101. Scher MS. The neural exposome influences the preterm fetal-toneonatal connectome. Pediatr Res 2024;95:9-11.