

R code used on the PRIMO data for the function F2:

```
#Creation of the database containing the information useful for the model:  
#TM is the value of the biomarker TM  
#V3 is the value of the biomarker V3  
#tps is the time between the infection and the visit measure (in days)  
base=data.frame(id=base_primo$id, TM=base_primo$TM,  
                 V3=base_primo$V3, tps=base_primo$time)  
  
#Database transformation for the brms package  
dt=base%>% gather(gp, y, V3,TM) %>% mutate (gp=as.factor(gp))  
  
#Prior specifications  
prior1 <- prior(normal(0,10000), nln = "b", lb=0)+  
           prior(normal(0,10000), nln = "c", lb=0)+  
           prior(normal(0,10000), nln = "num", class="b", lb=0)+  
           prior(uniform(0,10000),nln = "num", class="sd", group = "id") +  
           prior(uniform(0,10000),nln = "num", class="sd")  
  
#Model specification with the function F2 and random effet in a1 (fixed parameter b1)  
fit_F2_a1 <- brm(bf(y ~ num*exp(-b*exp(-tps/c))),  
                   num~-1+gp+(gp+0|id),  
                   b~-1+gp,  
                   c~-1+gp,  
                   sigma~-1+gp,  
                   nl = TRUE),  
                   data = dt,prior = prior1,  
                   chains = 4,iter=20000, control=list(adapt_delta=0.9))  
  
#Model specification with the function F2 and random effet in a2 (fixed parameter b2)  
fit_F2_a2 <- brm(bf(y ~ b*exp(-num*exp(-tps/c))),  
                   num~-1+gp+(gp+0|id),  
                   b~-1+gp,  
                   c~-1+gp,  
                   sigma~-1+gp,  
                   nl = TRUE),  
                   data = dt,prior = prior1,  
                   chains = 4,iter=20000, control=list(adapt_delta=0.9))
```

```

#Model specification with the function F2 and random effet in a3 (fixed parameter b3)
fit_F2_a3 <- brm(bf(y ~ b*exp(-c*exp(-tps/num)),
  num~-1+gp+(gp+0|id),
  b~-1+gp,
  c~-1+gp,
  sigma~-1+gp,
  nl = TRUE),
  data = dt,prior = prior1,
  chains = 4,iter=20000, control=list(adapt_delta=0.99))

#WAIC calculation
WAIC_fit_F2_a1=waic(fit_F2_a1)
WAIC_fit_F2_a2=waic(fit_F2_a2)
WAIC_fit_F2_a3=waic(fit_F2_a3)

```

VPC calculation details for a given biomarker:

$$VPC = \frac{\sigma_a^2}{\sigma_a^2 + \sigma_\epsilon^2}$$

We need to calculate the variance of the random effect:

$$g_b(t, a) = b_1 \exp(-(b_2 + a_2) \exp(\frac{-t}{b_3}))$$

$$g_b(t, a) = b_1 \exp(-b_2 \exp(\frac{-t}{b_3}) - a_2 \exp(\frac{-t}{b_3}))$$

$$g_b(t, a) = b_1 \exp(-b_2 \exp(\frac{-t}{b_3})) \exp(-a_2 \exp(\frac{-t}{b_3}))$$

We note $\lambda = b_1 \exp(-b_2 \exp(\frac{-t}{b_3}))$ and $k(t) = \exp(\frac{-t}{b_3})$

$$g_b(t, a) = \lambda \exp(-a_2 k(t))$$

$$-a_2 k(t) \sim \mathbb{N}(0, \sigma_a^2 k(t)^2)$$

$$V(\exp(-a_2 k(t))) = \exp(\sigma_a^2 k(t)^2) (\exp(\sigma_a^2 k(t)^2) - 1)$$

$$V(g_b(t, a)) = \lambda^2 \exp(\sigma_a^2 k(t)^2) (\exp(\sigma_a^2 k(t)^2) - 1)$$

$$V(g_b(t, a)) = \lambda^2 \sigma_a^2 k(t)^2 \text{ if } \sigma_a^2 k(t)^2 \text{ is small}$$

For a given marker of our model:

$$VPC = \frac{\lambda^2 \sigma_a^2 k(t)^2}{\lambda^2 \sigma_a^2 k(t)^2 + \sigma_\epsilon^2}$$

R code used for the simulation design:

```
#We fixed n=272 individuals as in the PRIMO ANRS C06 cohort
n<-272

#We start by generating a number of measurements per patient according to
the distribution of Primo:
#134 patients with one measure [0,0.49[
#8 patients with two measures [0.49,0.497[
#12 patients with three measures [0.497,0.54[
#47 patients with four measures [0.54,0.71[
#13 patients with five measures[0.71,0.76[
#33 patients with six measures [0.76, 0.88[
#25 patients with seven measures [0.88,1]

nb_mesure=runif(n)

nb_mesure=replace(nb_mesure, nb_mesure<0.49,1)
nb_mesure=replace(nb_mesure, nb_mesure>=0.49 & nb_mesure<0.497,2)
nb_mesure=replace(nb_mesure, nb_mesure>=0.497 & nb_mesure<0.54,3)
nb_mesure=replace(nb_mesure, nb_mesure>=0.54 & nb_mesure<0.71,4)
nb_mesure=replace(nb_mesure, nb_mesure>=0.71 & nb_mesure<0.76,5)
nb_mesure=replace(nb_mesure, nb_mesure>=0.76 & nb_mesure<0.88,6)
nb_mesure=replace(nb_mesure, nb_mesure!=1 & nb_mesure!=2 & nb_mesure!=3 &
nb_mesure!=4 & nb_mesure!=5 & nb_mesure!=6 ,7)

#We generate the date of the first measurement
tps_J0=round(rlnorm(n, meanlog =4, sdlog = 0.50),0)

#We create the times t of measurements of individuals according
to the number of measurements:
time_individu=matrix(NA, ncol=n, nrow=7)
for (i in 1:n){
  time_individu[1,i]=tps_J0[i]
  if (nb_mesure[i]==2) {
    time_individu[2,i]=time_individu[1,i]+30
  } else if (nb_mesure[i]==3) {
    time_individu[2,i]=time_individu[1,i]+30
    time_individu[3,i]=time_individu[1,i]+3*30
  } else if (nb_mesure[i]==4) {
    time_individu[2,i]=time_individu[1,i]+30
    time_individu[3,i]=time_individu[1,i]+3*30
  }
```

```

        time_individu[4,i]=time_individu[1,i]+6*30
    } else if  (nb_mesure[i]==5) {
        time_individu[2,i]=time_individu[1,i]+30
        time_individu[3,i]=time_individu[1,i]+3*30
        time_individu[4,i]=time_individu[1,i]+6*30
        time_individu[5,i]=time_individu[1,i]+12*30
    } else if  (nb_mesure[i]==6) {
        time_individu[2,i]=time_individu[1,i]+30
        time_individu[3,i]=time_individu[1,i]+3*30
        time_individu[4,i]=time_individu[1,i]+6*30
        time_individu[5,i]=time_individu[1,i]+12*30
        time_individu[6,i]=time_individu[1,i]+18*30
    }
    else if  (nb_mesure[i]==7) {
        time_individu[2,i]=time_individu[1,i]+30
        time_individu[3,i]=time_individu[1,i]+3*30
        time_individu[4,i]=time_individu[1,i]+6*30
        time_individu[5,i]=time_individu[1,i]+12*30
        time_individu[6,i]=time_individu[1,i]+18*30
        time_individu[7,i]=time_individu[1,i]+24*30
    }
else {
    time_individu[1,i]=time_individu[1,i]
}

}
}

#We repeat the id of the individual according to the number of measurements
id=rep(1:n,nb_mesure)
base=data.frame(id=id)
vector_time=c(time_individu)

#We add the different times in the base:
base$tps=vector_temps

#We generate the values of TM and V3 for each time and for each individual:
b11<-70
b21<-65
b12<-3
b22<-4
b13<-200
b23<-100

```

```

mu=c(0,0)
Sigma=matrix(c(4,2,2,6.25),ncol=2)
a<-MASS::mvrnorm(n=n,mu=mu,Sigma=Sigma)
alea=data.frame(id=1:272,a1=a[,1], a2=a[,2])

base=right_join(base,alea,by=c("id"))

TM_mean=NULL
V3_mean=NULL
for (i in 1: nrow(base)){
  TM_mean[i]=b11*exp(-(b12+base$a1[i])*exp(-base$tps[i]/b13))
  V3_mean[i]=b21*exp(-(b22+base$a2[i])*exp(-base$tps[i]/b23))
}

Sigma=diag(c(8.08,9.78))
for (i in 1:nrow(base)){
  base$TM[i]=MASS::mvrnorm(n=1,mu=c(TM_mean[i],V3_mean[i]),Sigma=Sigma)[1]
  base$V3[i]=MASS::mvrnorm(n=1,mu=c(TM_mean[i],V3_mean[i]),Sigma=Sigma)[2]
}

#We apply the biomarkers censorship:

base=base %>% mutate(TM = ifelse(TM >70, 70, TM))
base=base %>% mutate(V3 = ifelse(V3 > 70, 70, V3))

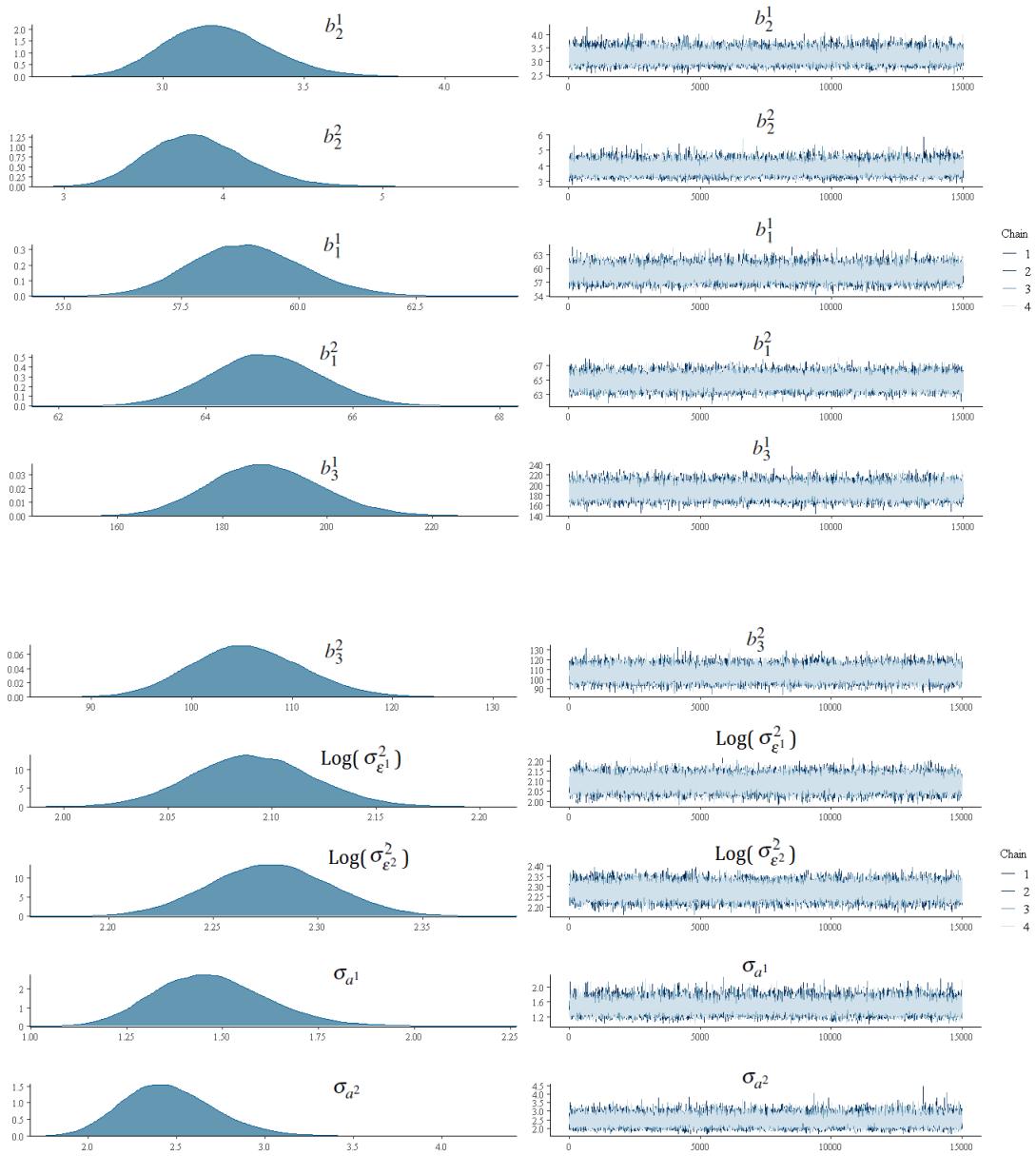
base=base %>% mutate(TM = ifelse(TM <0, 0 , TM))
base=base %>% mutate(V3 = ifelse(V3 < 0 , 0 , V3))

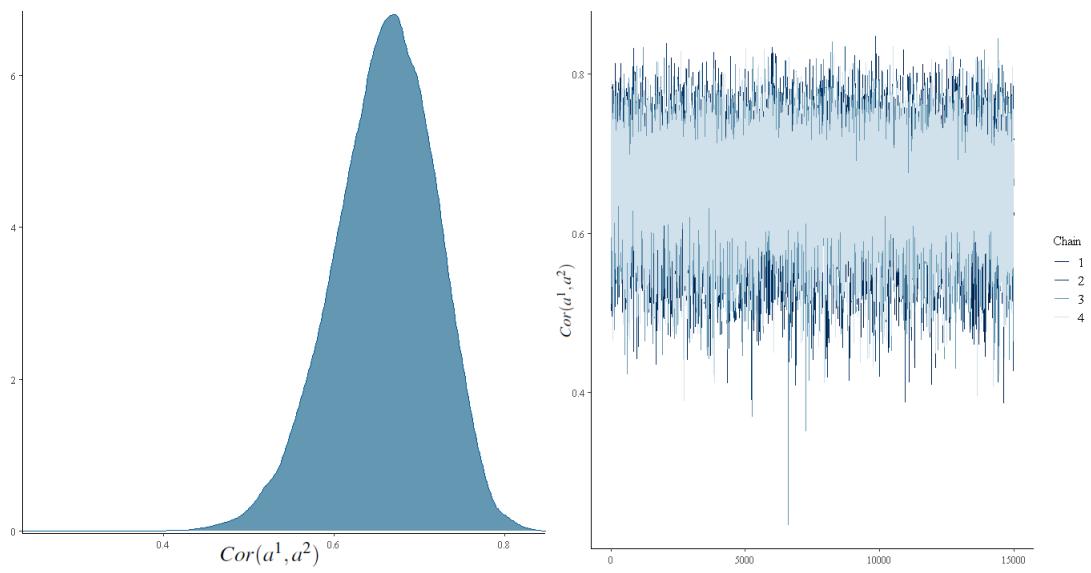
base$TM=round(base$TM,1)
base$V3=round(base$V3,1)

base

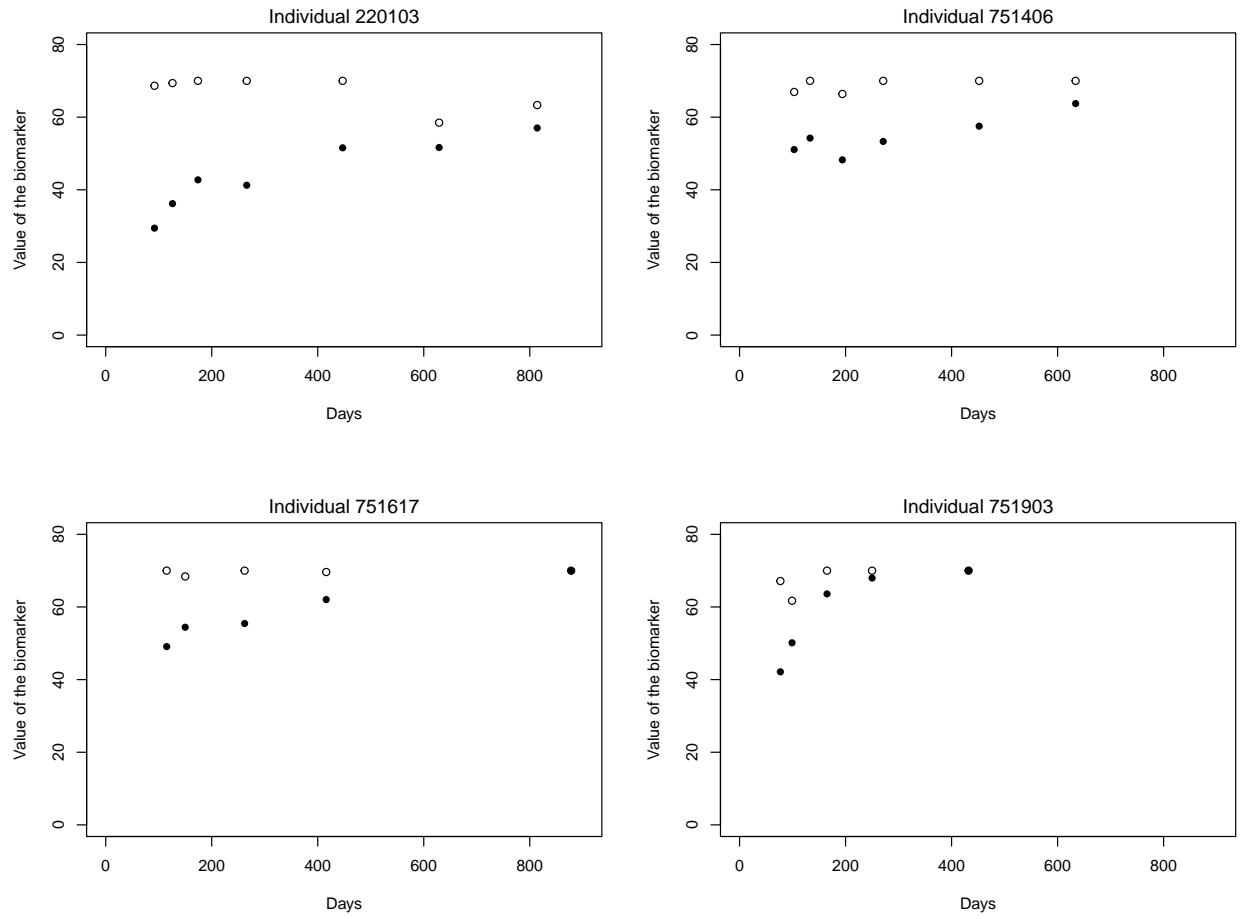
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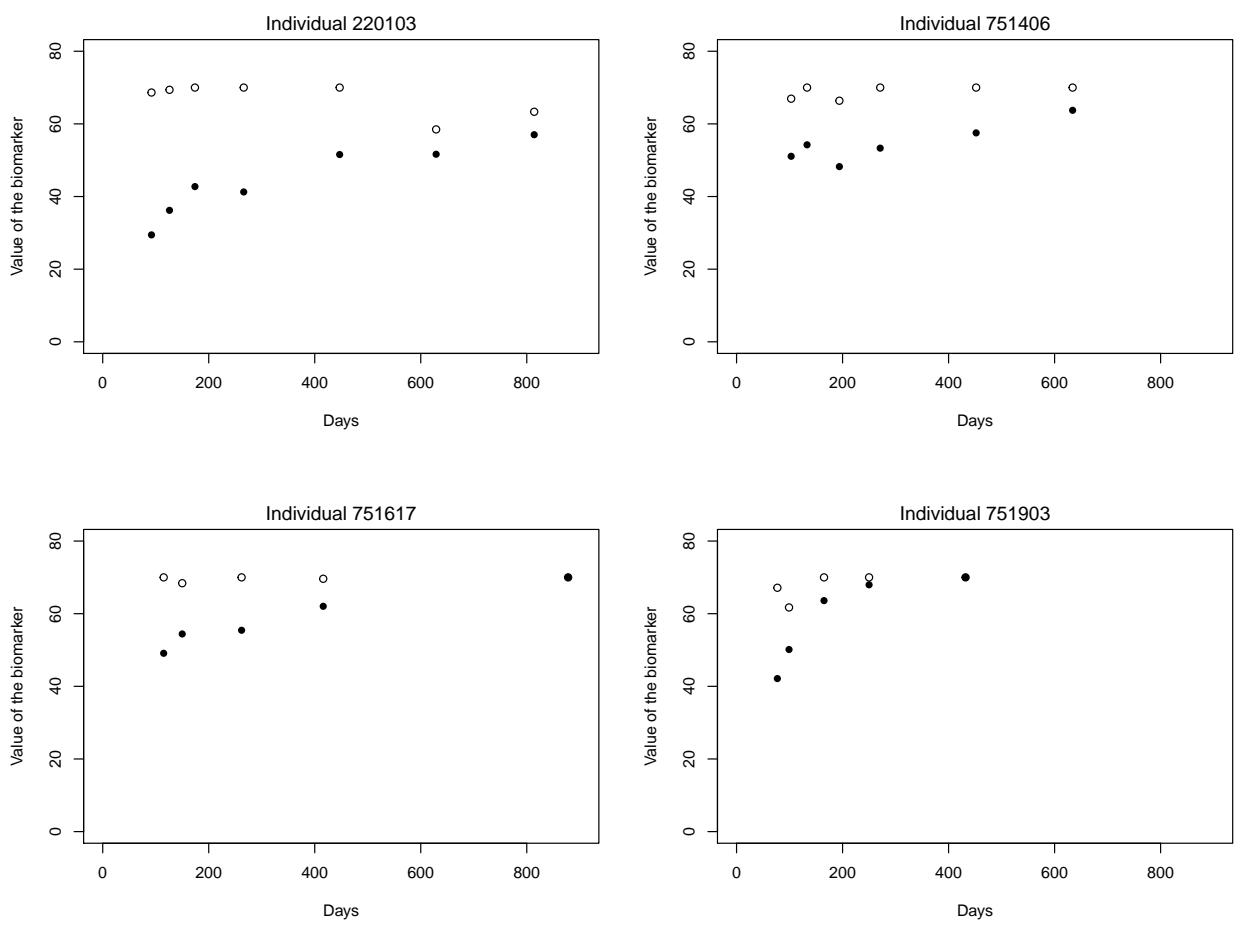
Trace plots and marginal posterior distributions for the PRIMO ANRS C06 results:





Evolution of the TM and V3 biomarkers for the four individuals who did not converge:





R code used and results for the univariate models:

```
#We load the 200 simulated databases
base_simul=readRDS( "base_primo_simul.rds")

#We define the sigmoid function of the estimate, namely the function F2
sigmoid.model <- function(psi, id, xidep) {
  tps <- xidep[, 1]
  A <- psi[id, 1]
  B <- psi[id, 2]
  C <- psi[id, 3]
  resp <- A*exp(-B*exp(-tps/C))
  return(resp)
}

#We estimate the parameters for the function F2 with the random
#effect in a2 for the biomarker TM
saemix_F2_A2_TM=list()
for (i in 1:200){

  base=base_simul[[i]]

  modele <- saemixData(name.data = base, name.group = "id",
                        name.predictors = "tps", name.response = "TM",
                        units = list(x = "Days"))

  #Parametrisation
  opt <- list(save = FALSE, save.graphs = FALSE)

  base.modelA2 <- saemixModel(model = sigmoid.model,
                                 description = "Sigmoid growth",
                                 psi0 = matrix(c(24.63, 3.48, 49.47), ncol = 3, byrow = TRUE,
                                               dimnames = list(NULL, c("A", "B", "C"))),
                                 covariance.model = matrix(c(0, 0, 0, 0, 1, 0, 0, 0, 0), ncol = 3, byrow = TRUE))

  saemix_F2_A2_TM[[i]]=saemix(base.modelA2, modele, opt)
}

#We estimate the parameters for the function F2 with the random
#effect in a2 for the biomarker V3
saemix_F2_A2_V3=list()
for (i in 1:200){

  base=base_simul[[i]]
```

```

modele <- saemixData(name.data = base, name.group = "id",
                      name.predictors = "tps", name.response = "V3",
                      units = list(x = "Days"))

base.modelA2 <- saemixModel(model = sigmoid.model,
                               description = "Sigmoid growth",
                               psi0 = matrix(c(24.63, 3.48, 49.47), ncol = 3, byrow = TRUE,
                               dimnames = list(NULL, c("A", "B", "C"))),
                               covariance.model = matrix(c(0, 0, 0, 0, 0, 1, 0, 0, 0, 0),
                               ncol = 3, byrow = TRUE))

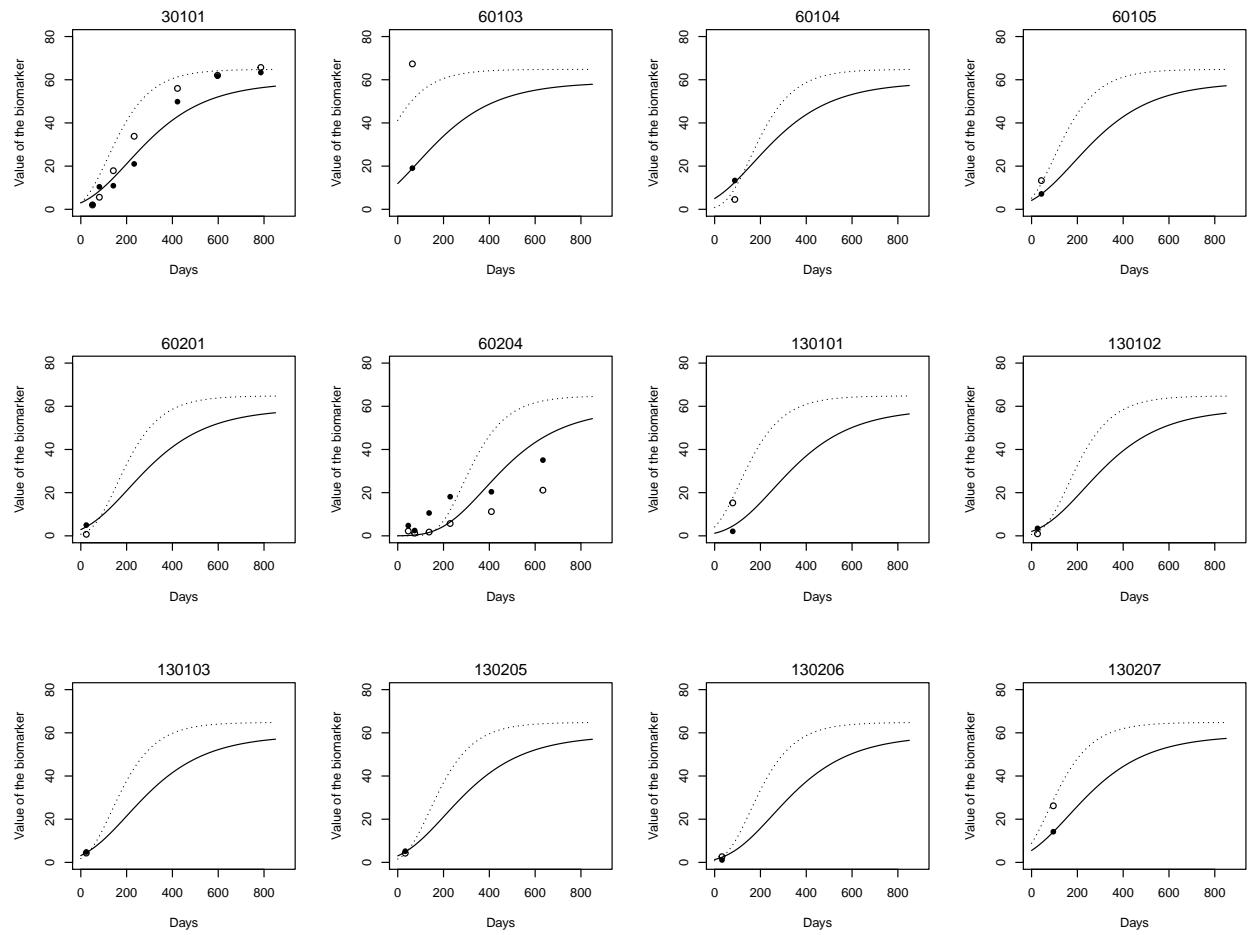
saemix_F2_A2_V3[[i]]=saemix(base.modelA2, modele, opt)
}

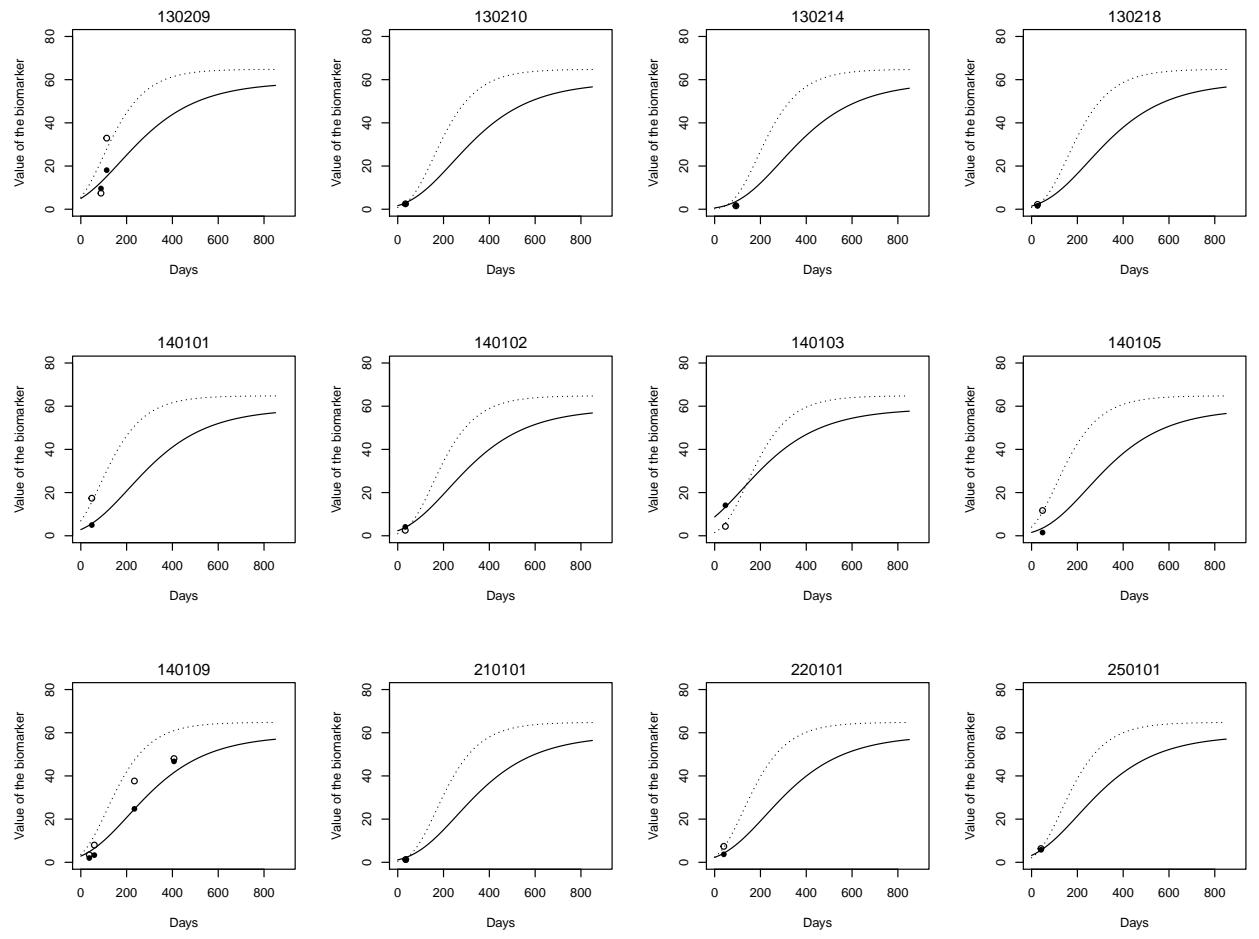
```

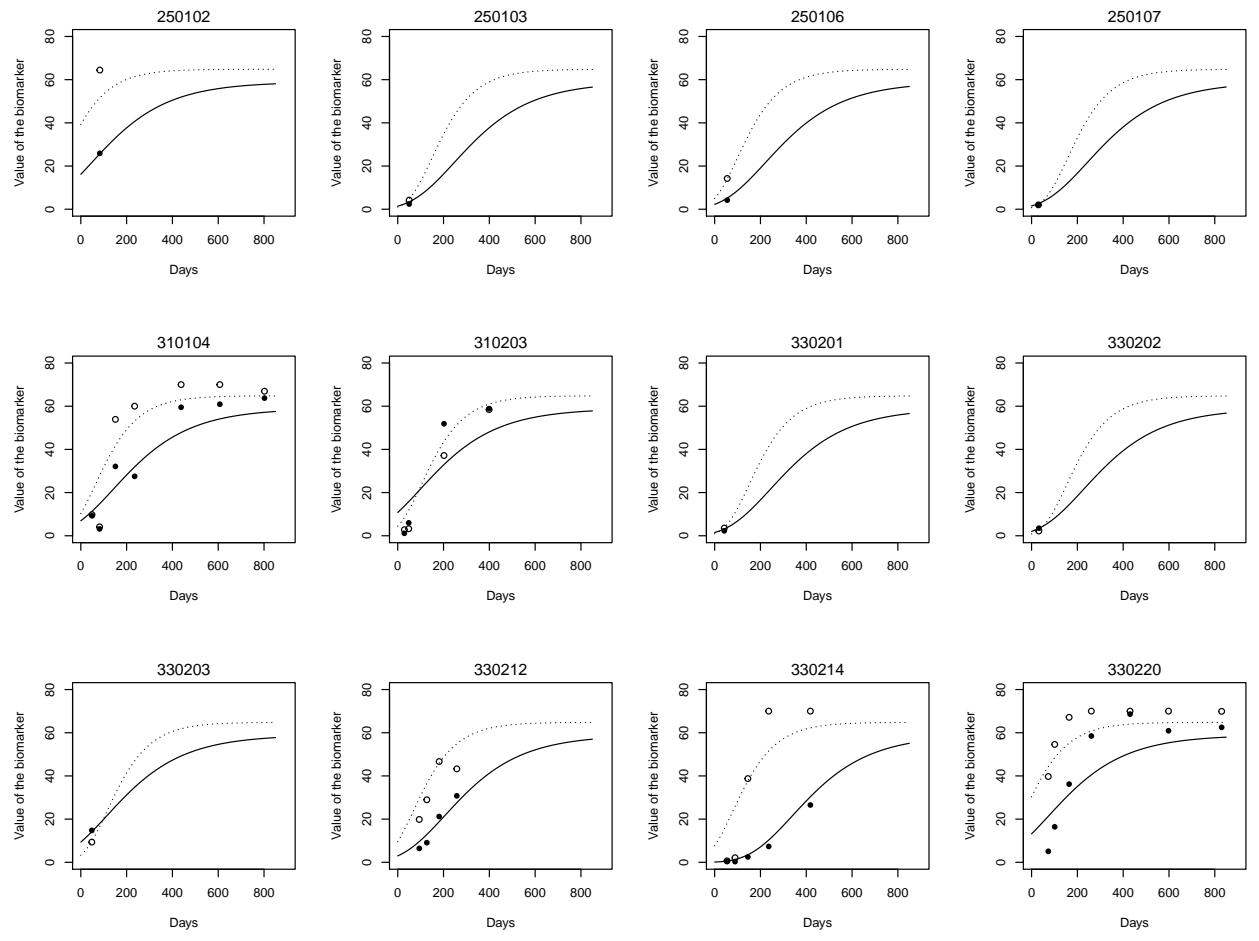
Table 1: True value of the parameters, mean, absolute relative bias (ARB), empirical root mean squared error (RMSE), and coverage rate (CR) of the credibility interval at 95% of the estimated parameters for the 200 simulations

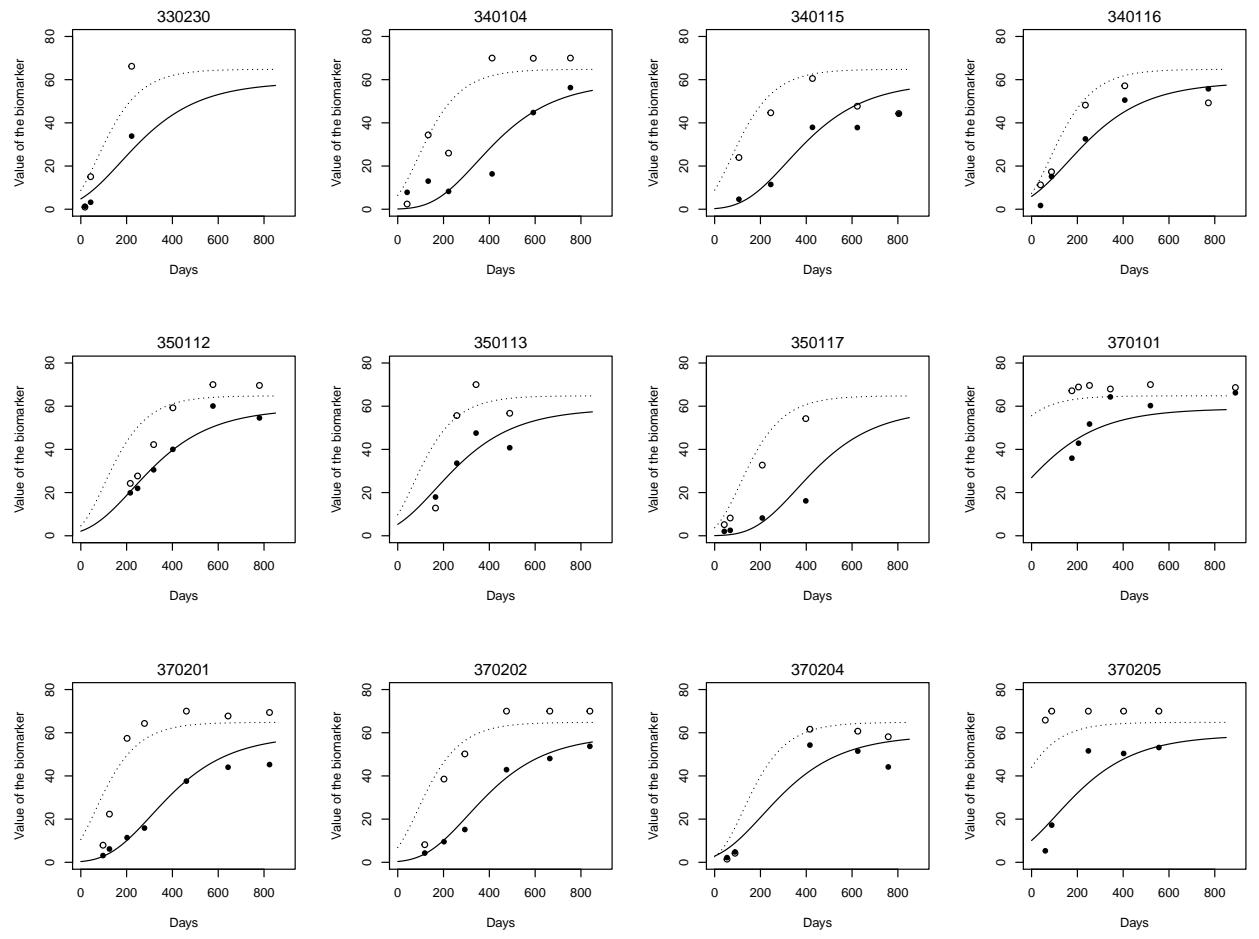
Parameters	True value	Mean	ARB	RMSE	CR
b_1^1	70	69.9	0.09%	0.38	96.5
b_2^1	3	3	0%	0.11	96.5
b_3^1	200	201.7	0.9%	3.5	87
b_1^2	65	65.3	0.46%	0.39	78
b_2^2	4	3.9	2.5%	0.19	88.5
b_3^2	100	101.9	1.9%	2.6	66
σ_{a^1}	2	1.8	10%	0.8	25
σ_{a^2}	2.5	2.2	12%	1.3	33
σ_{ϵ^1}	2.8	2.6	7%	0.2	30
σ_{ϵ^2}	3.1	3	3.5%	0.16	63

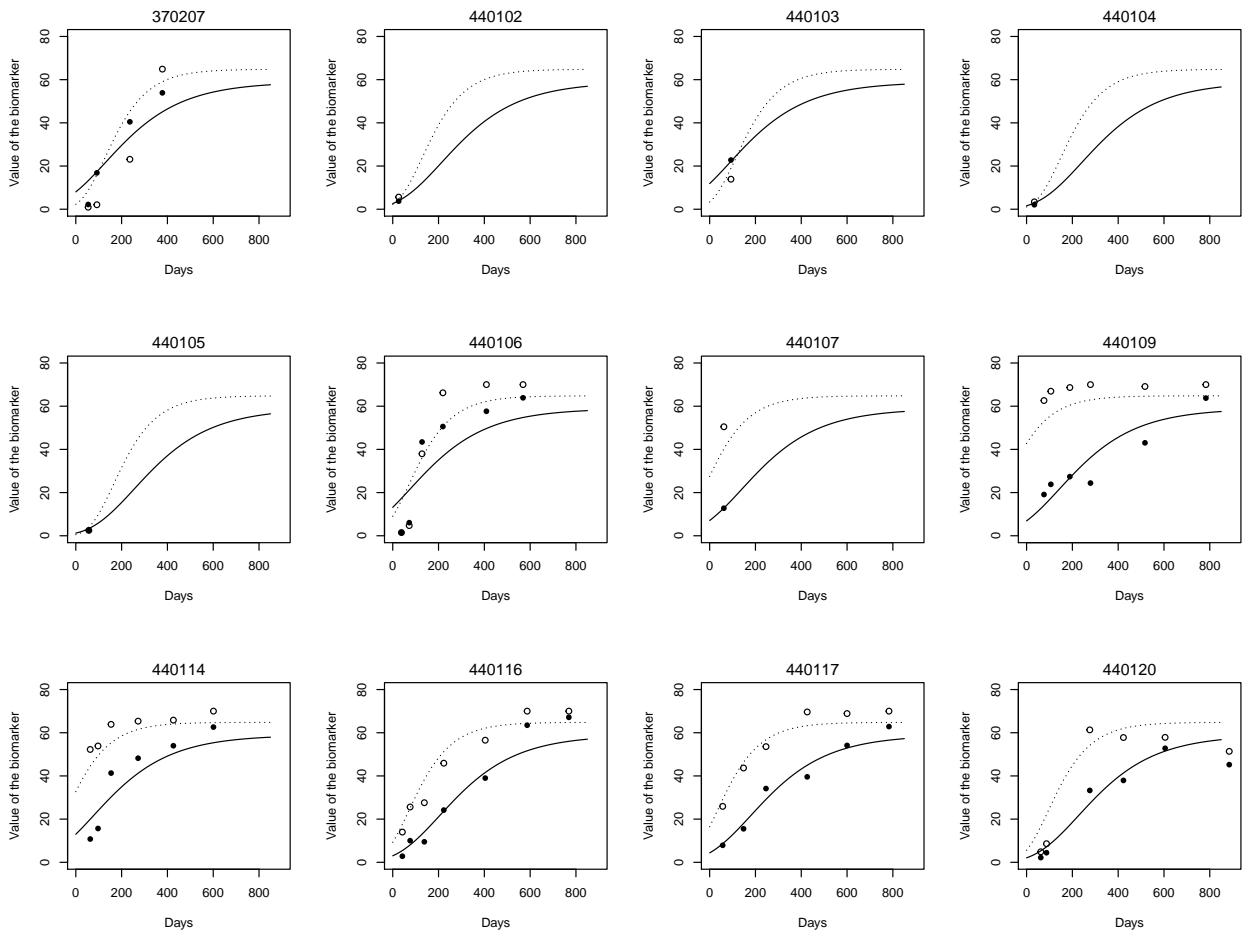
Values of the TM and V3 biomarkers with the observed (full circle for TM and empty circle for V3) and predicted trajectories (solid curve for TM and dotted line for V3) for all subjects of the PRIMO-ANRS C06 cohort

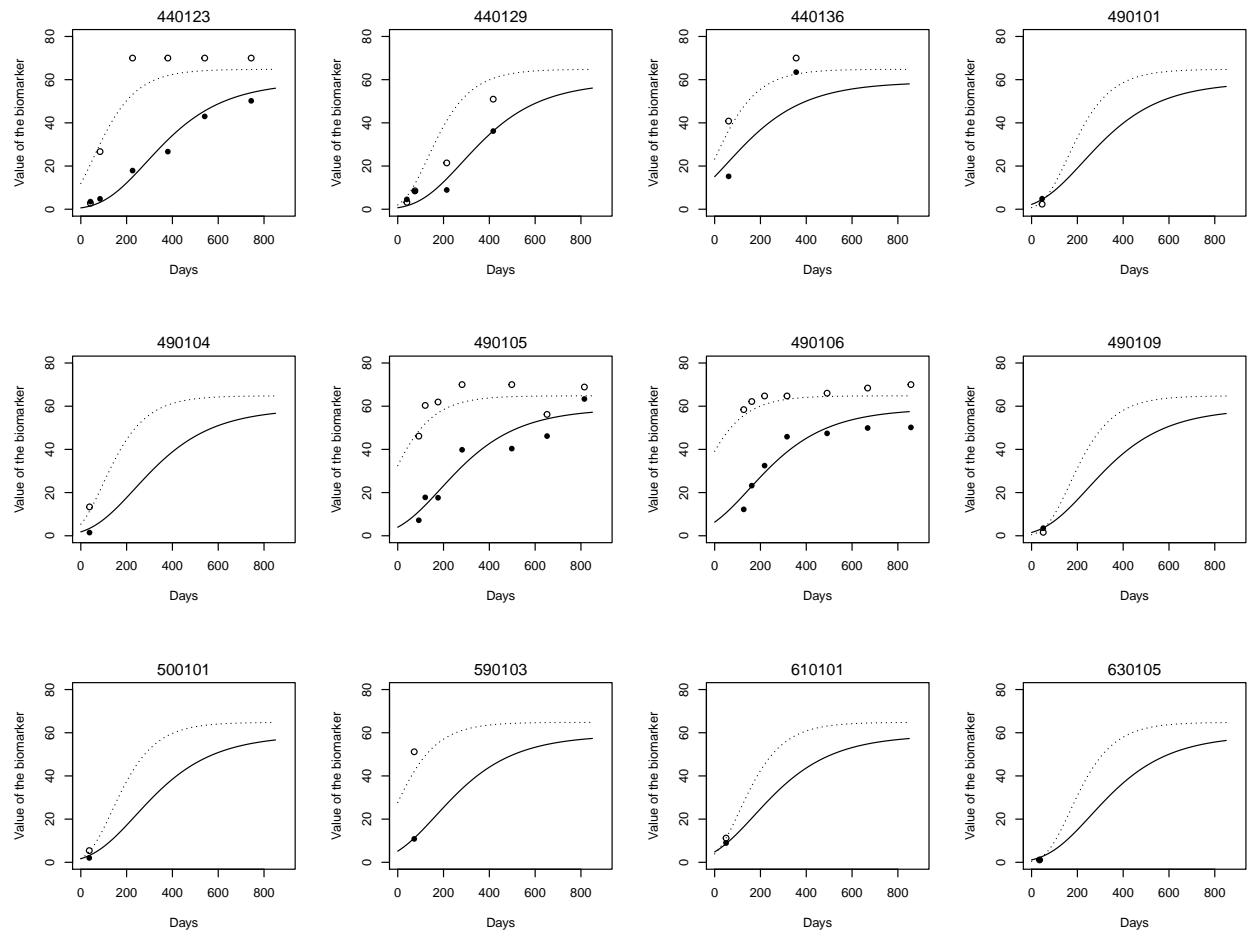


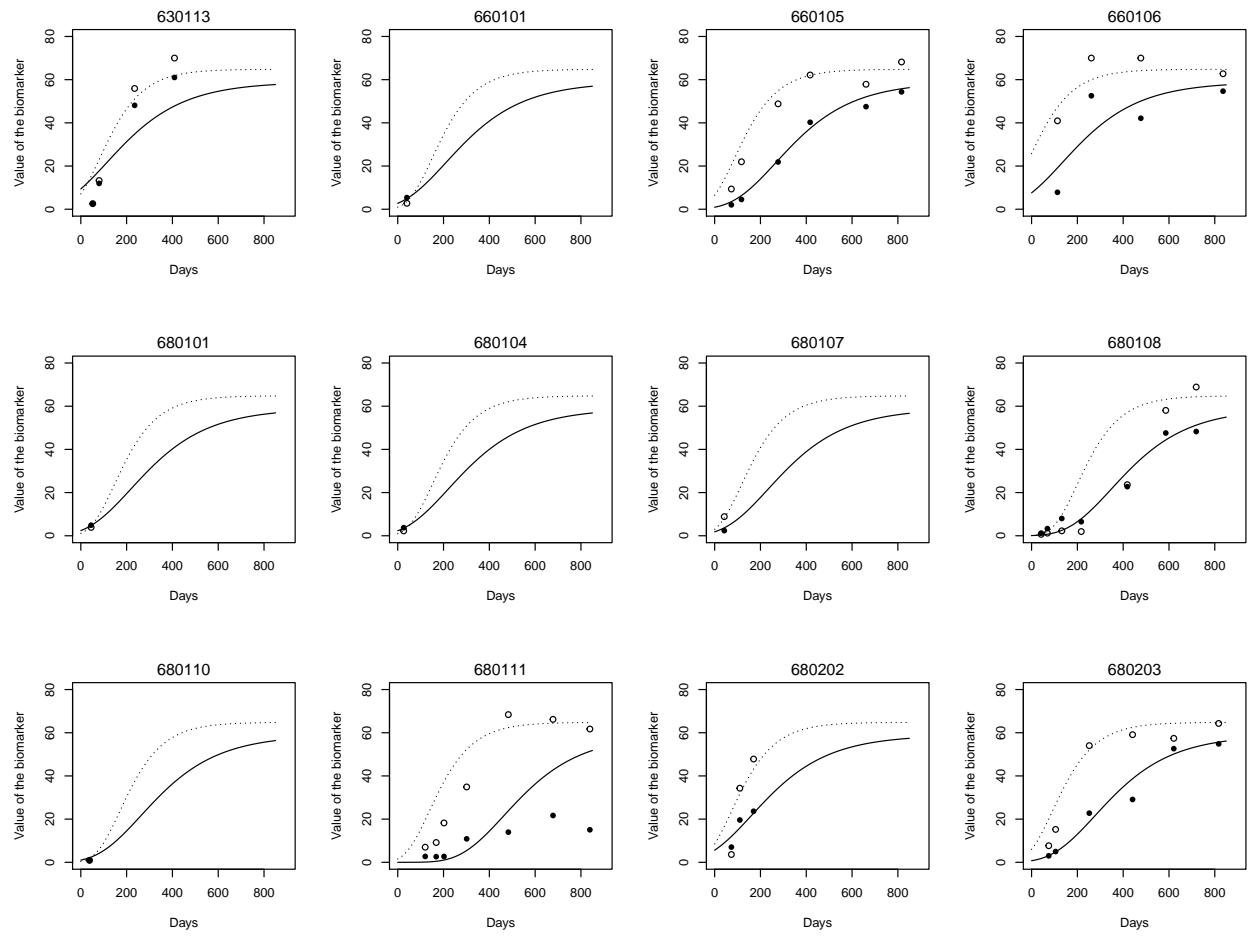


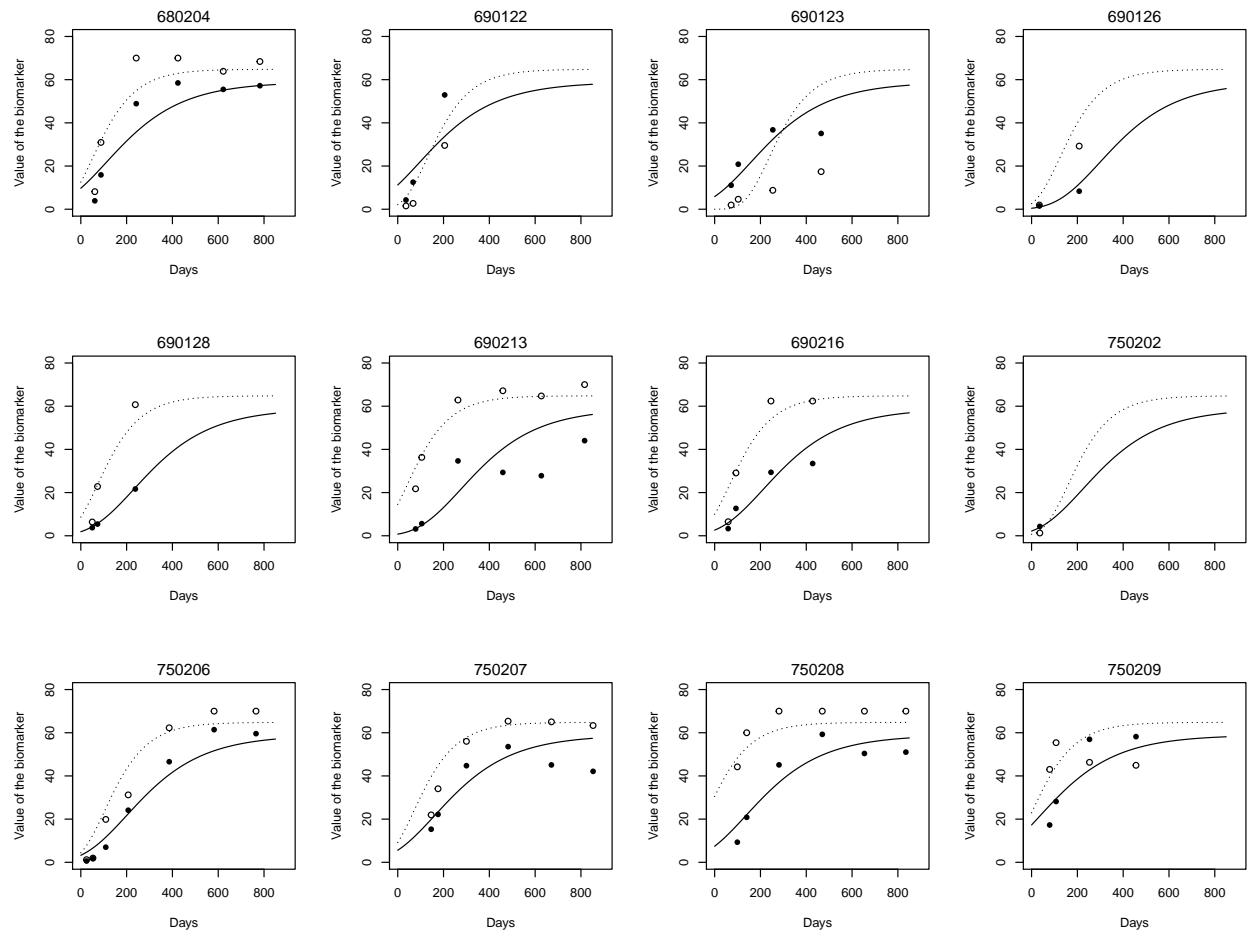


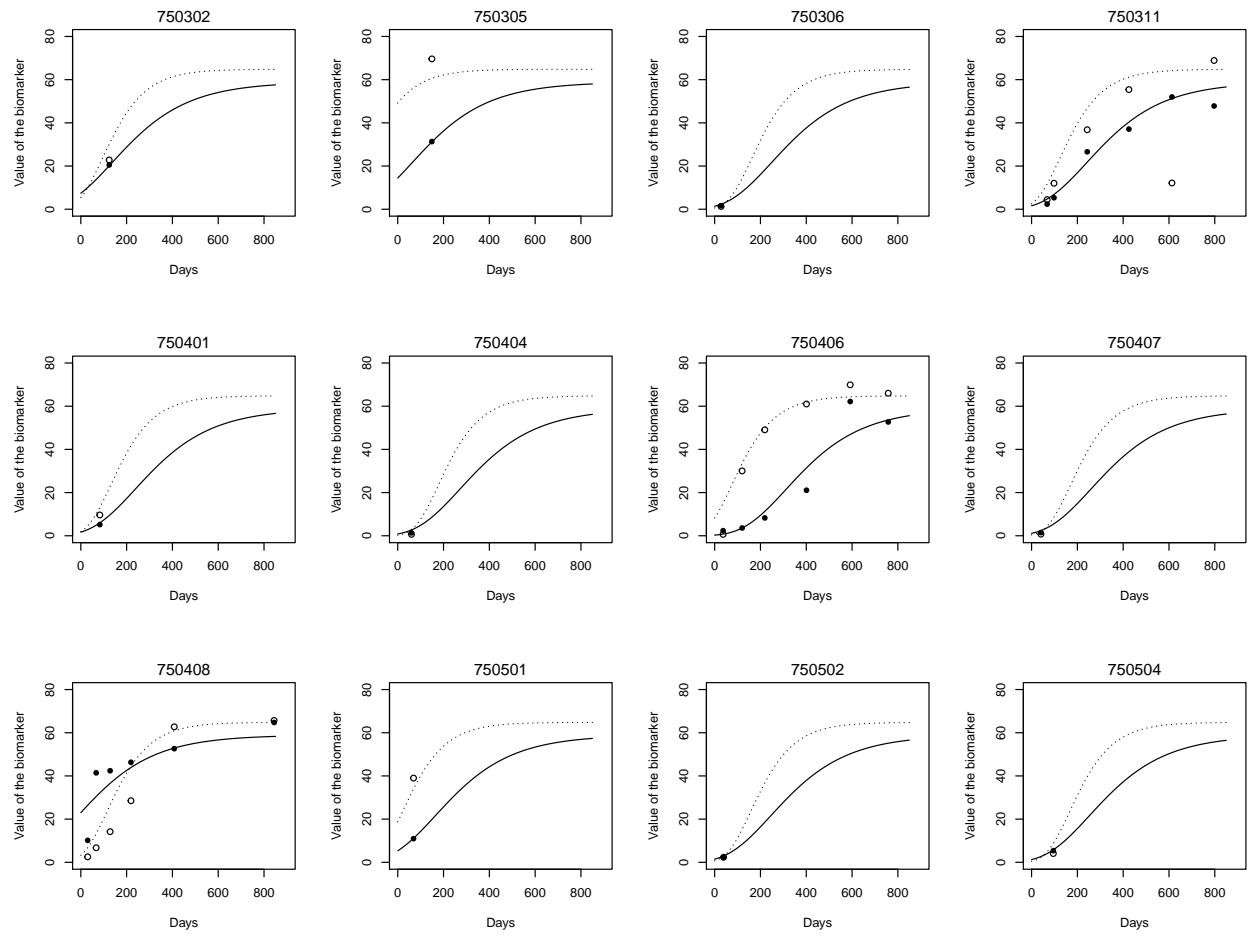


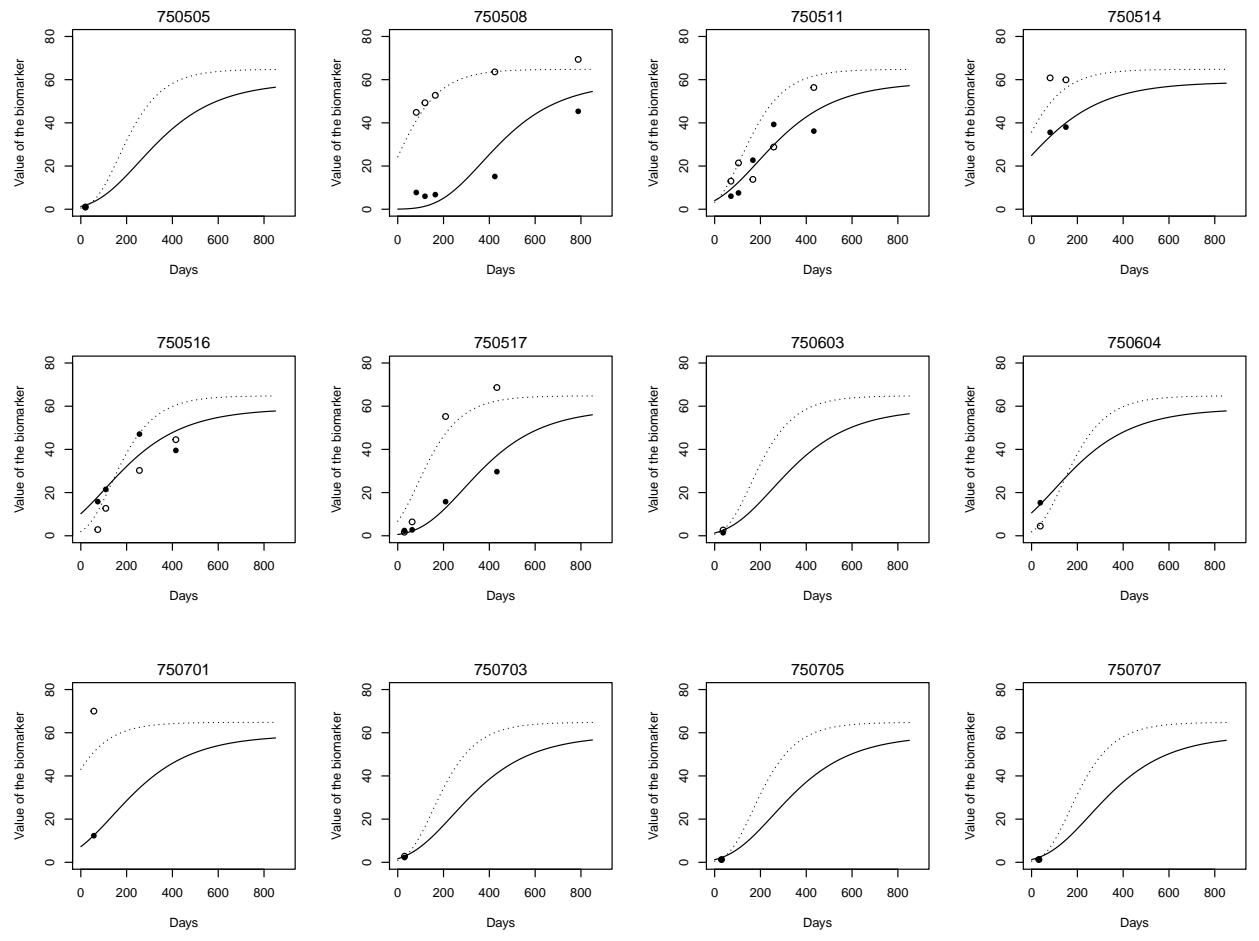


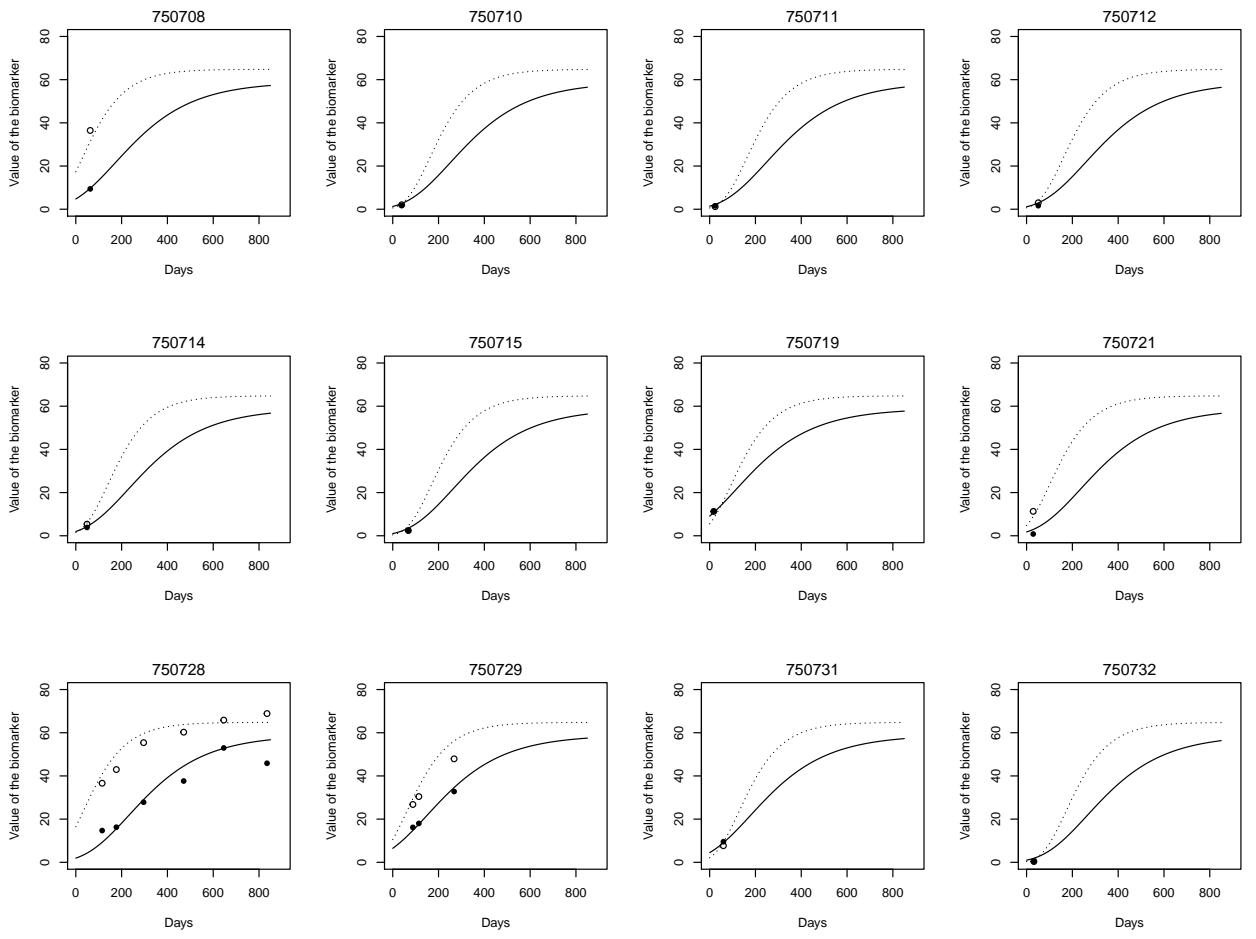


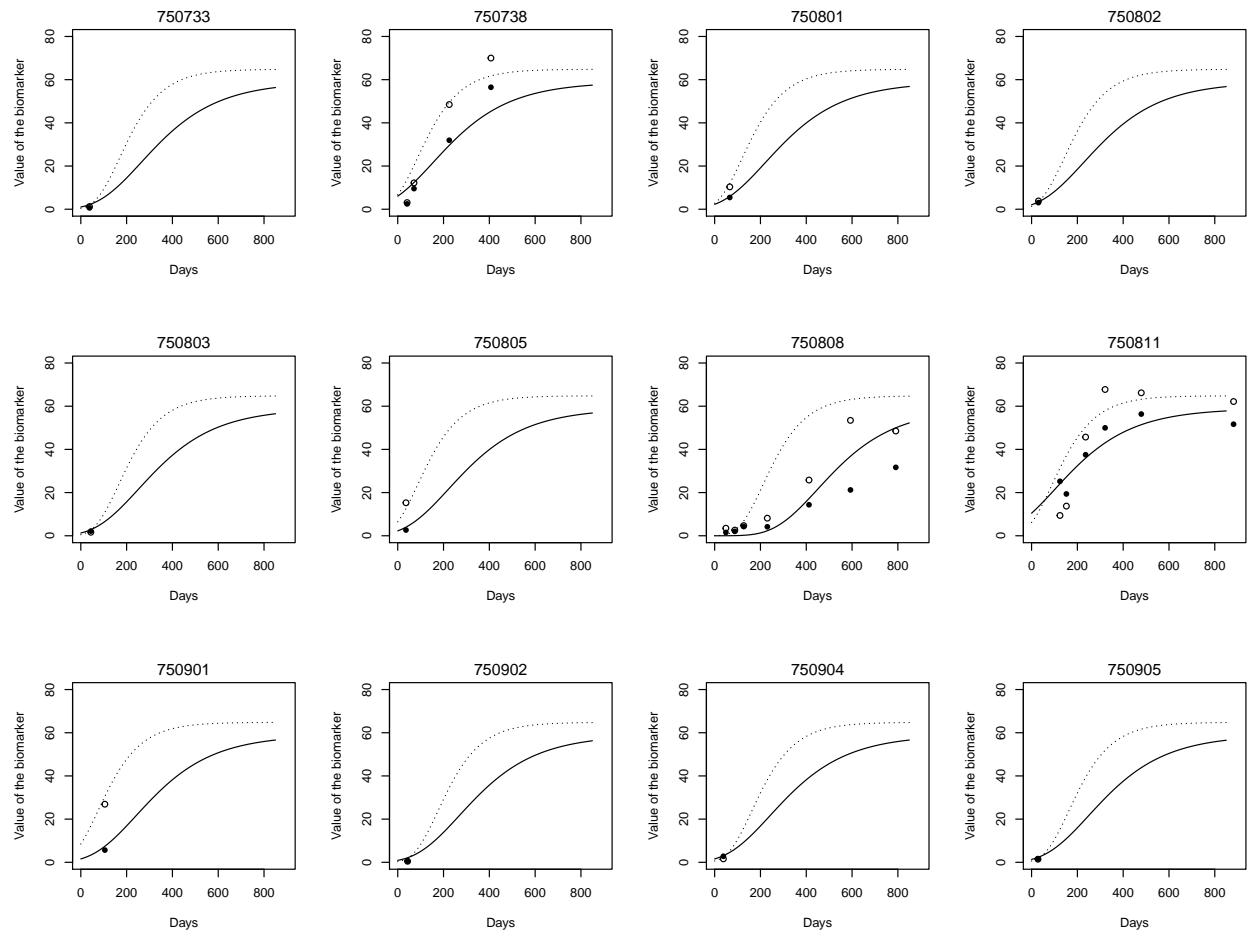


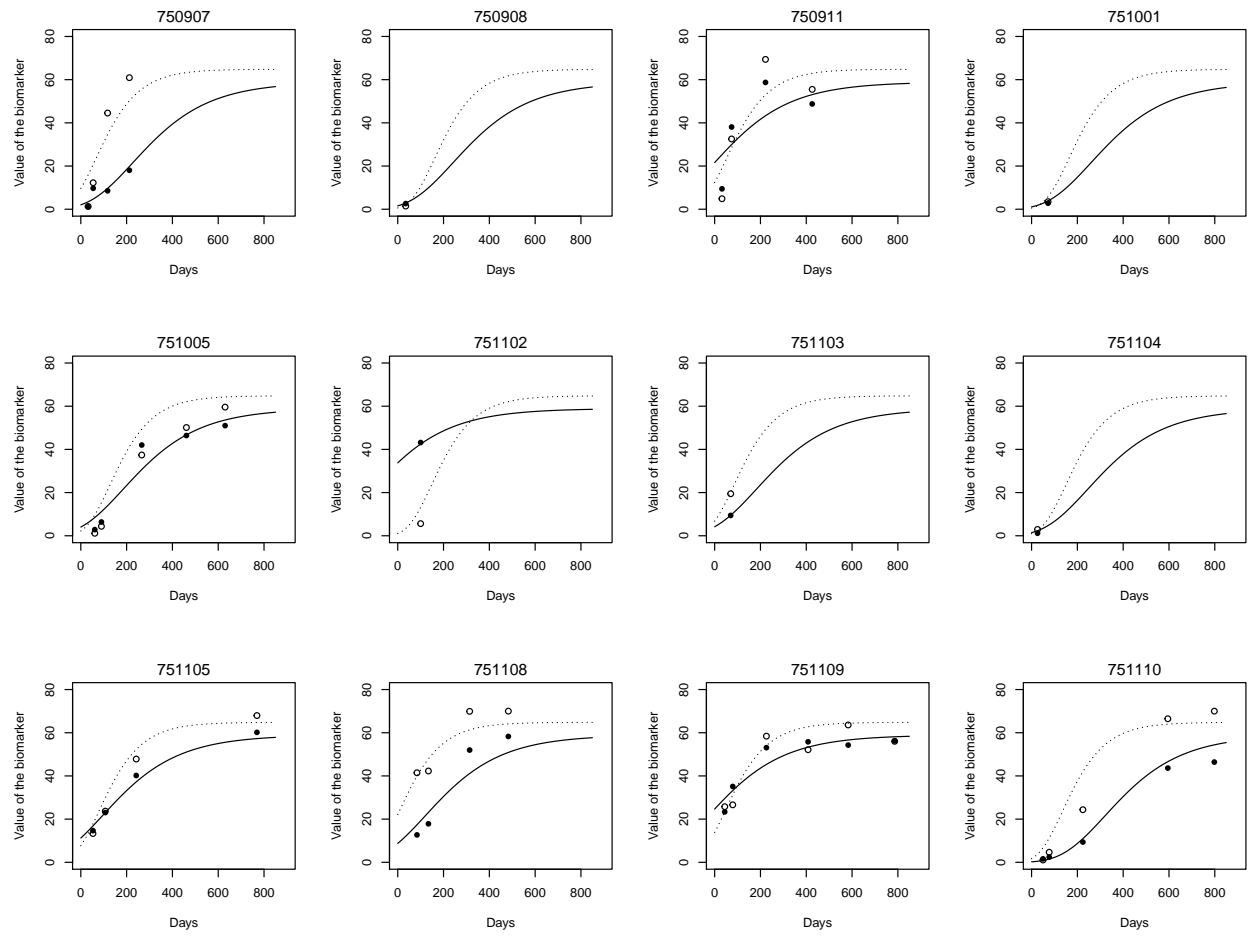


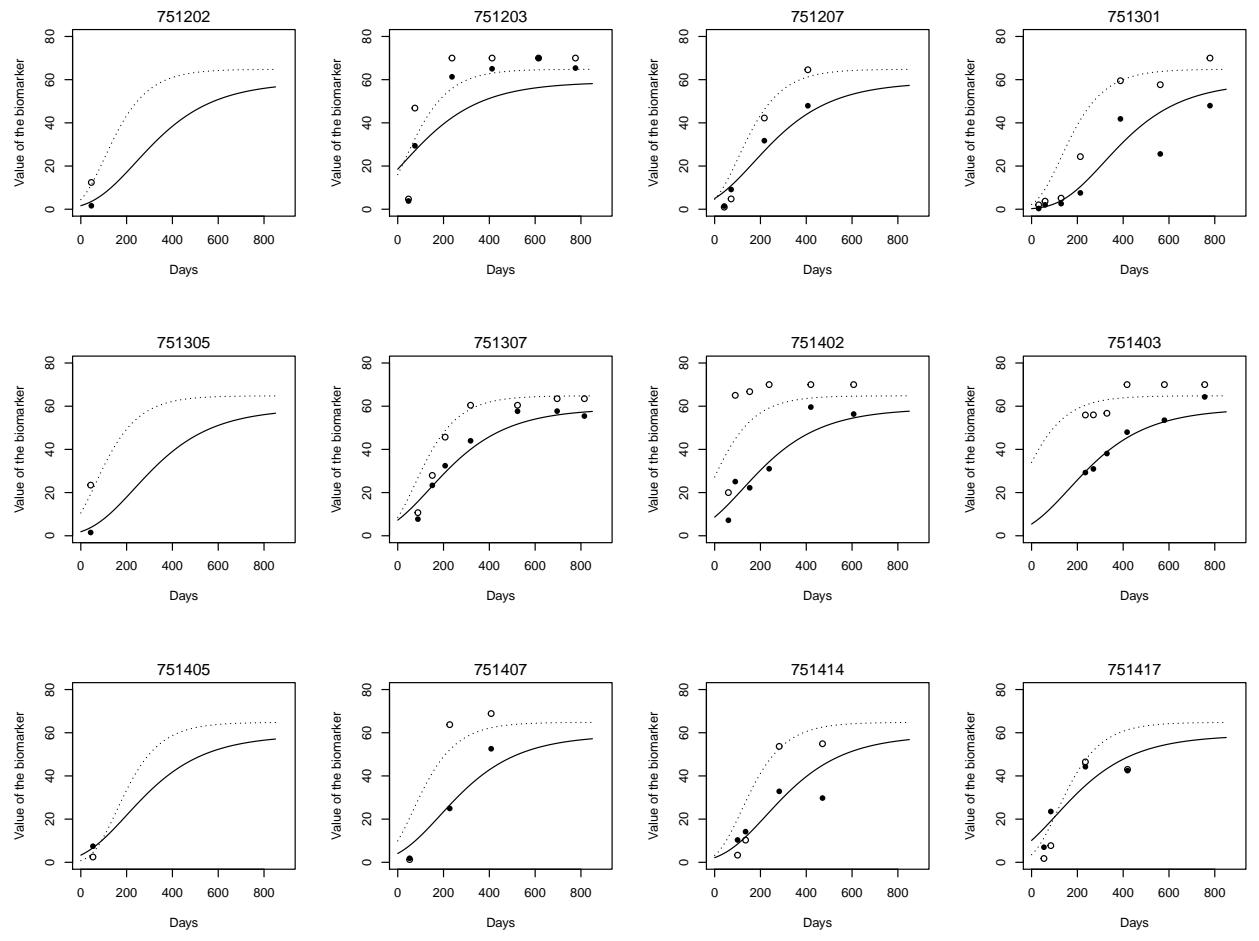


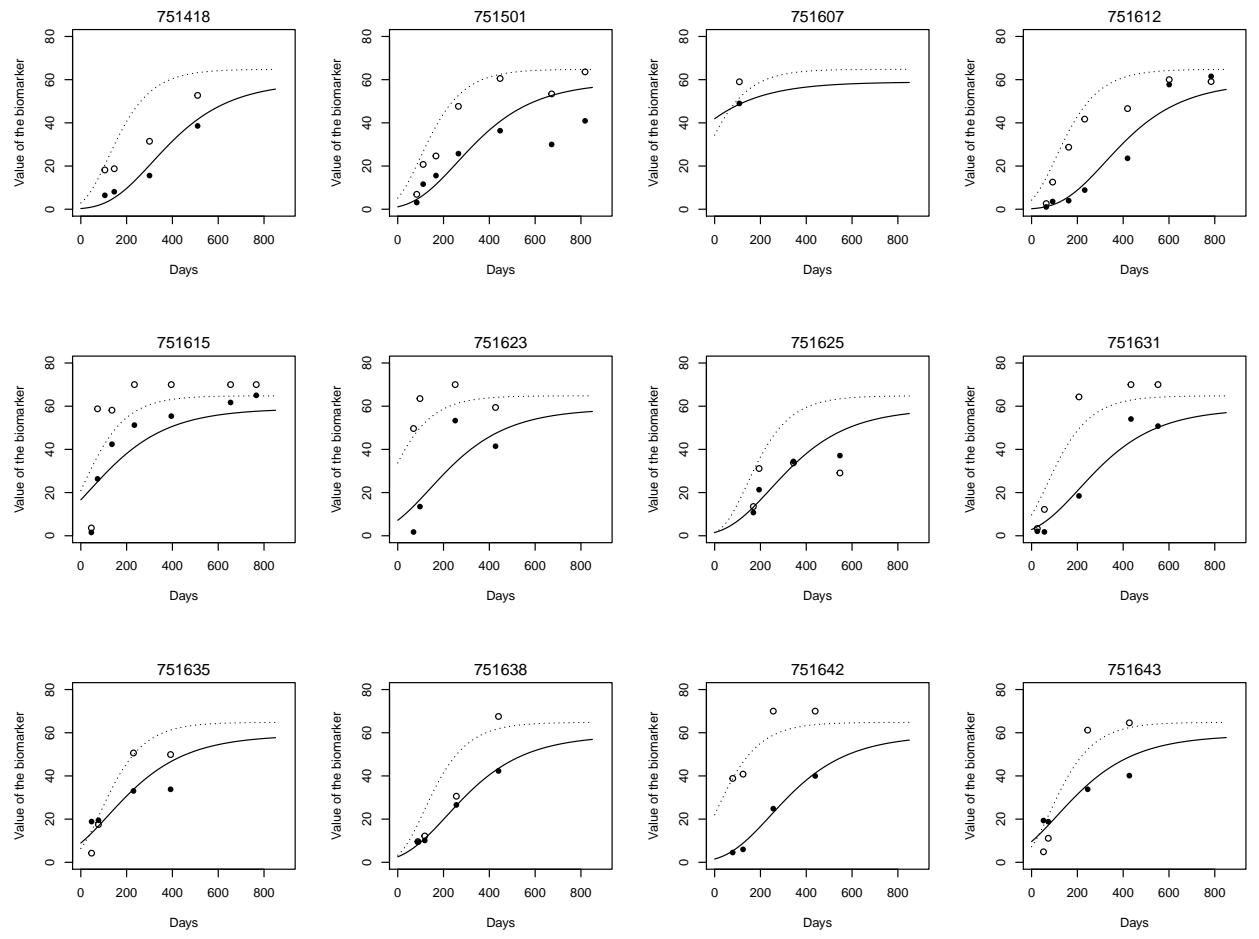


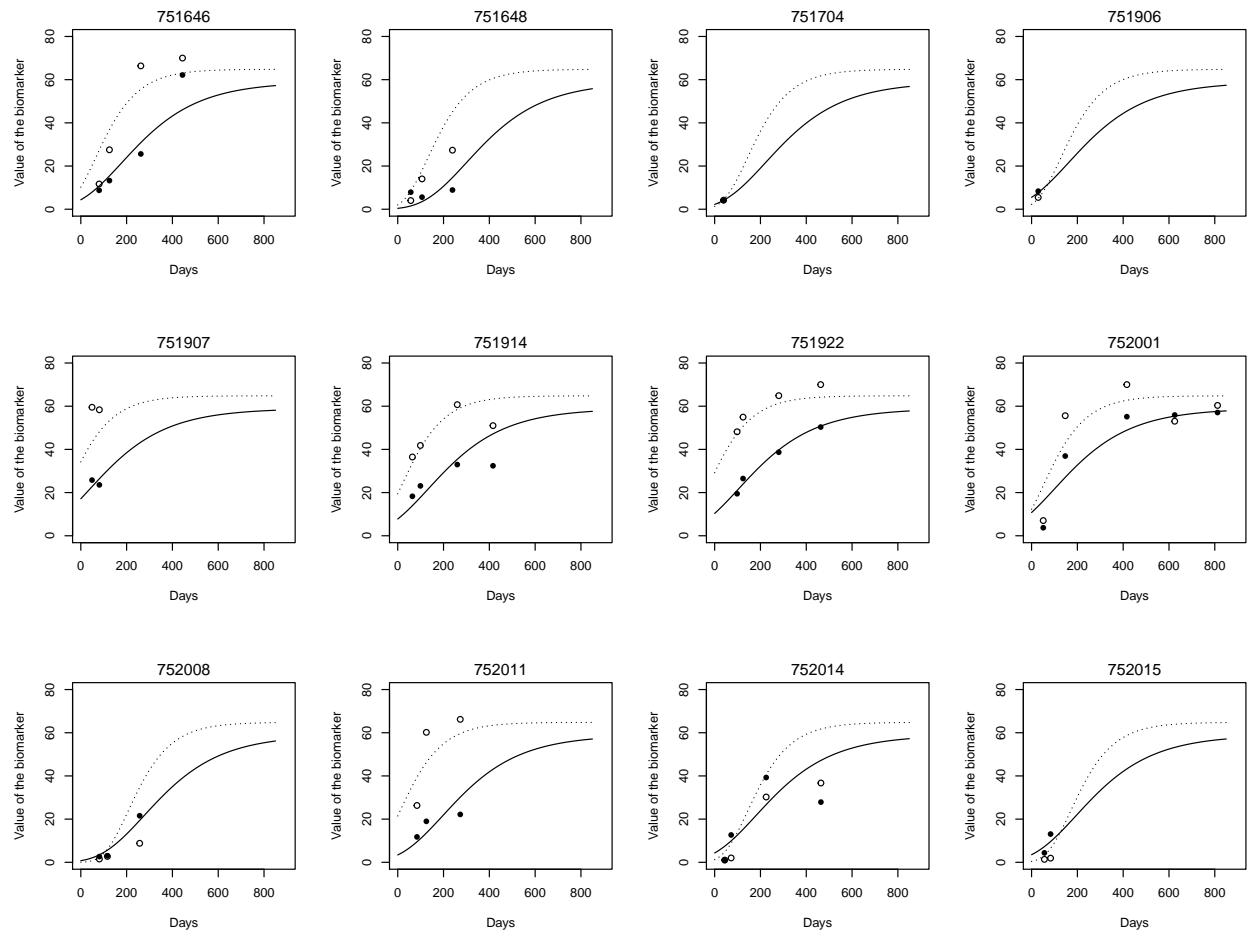


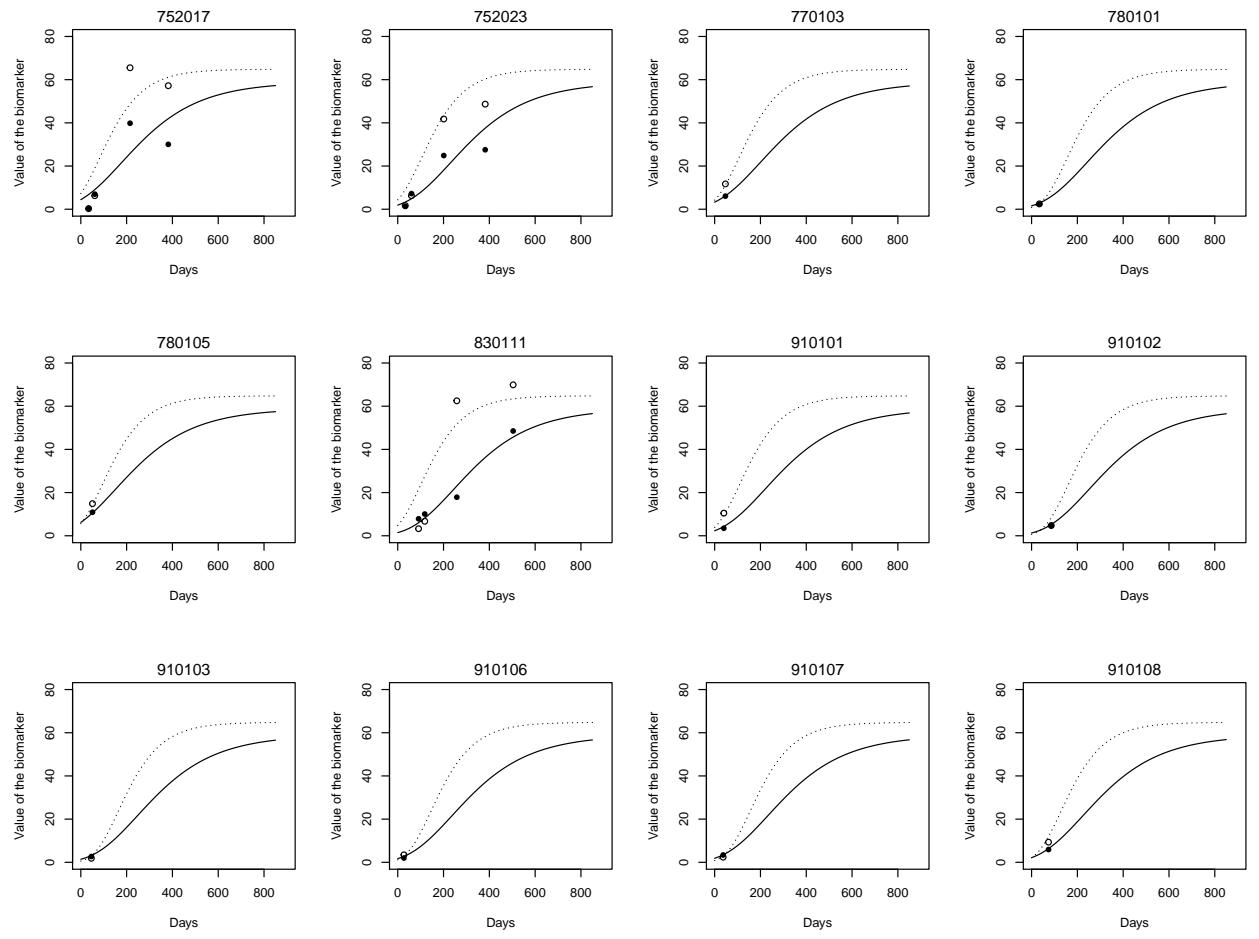


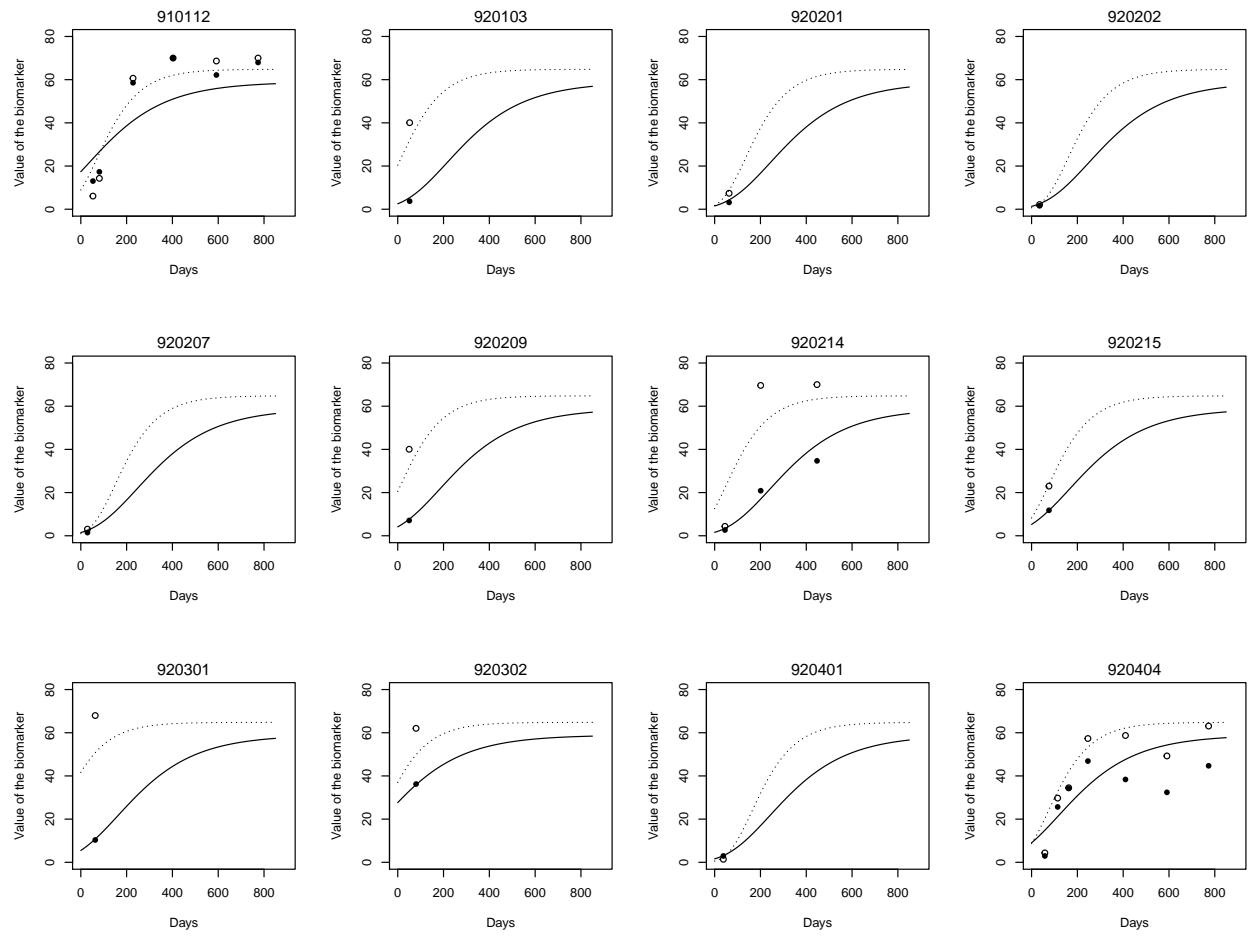


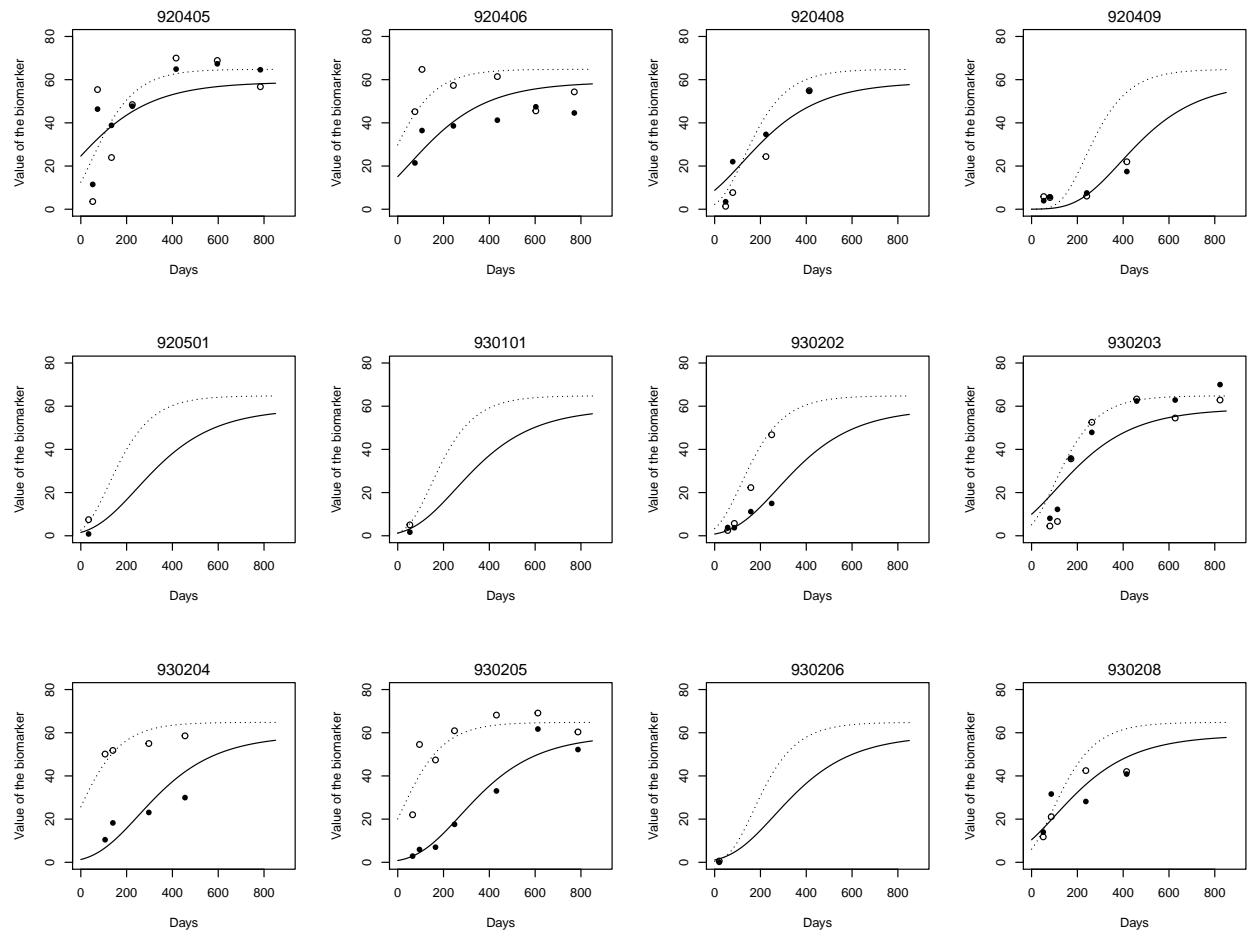


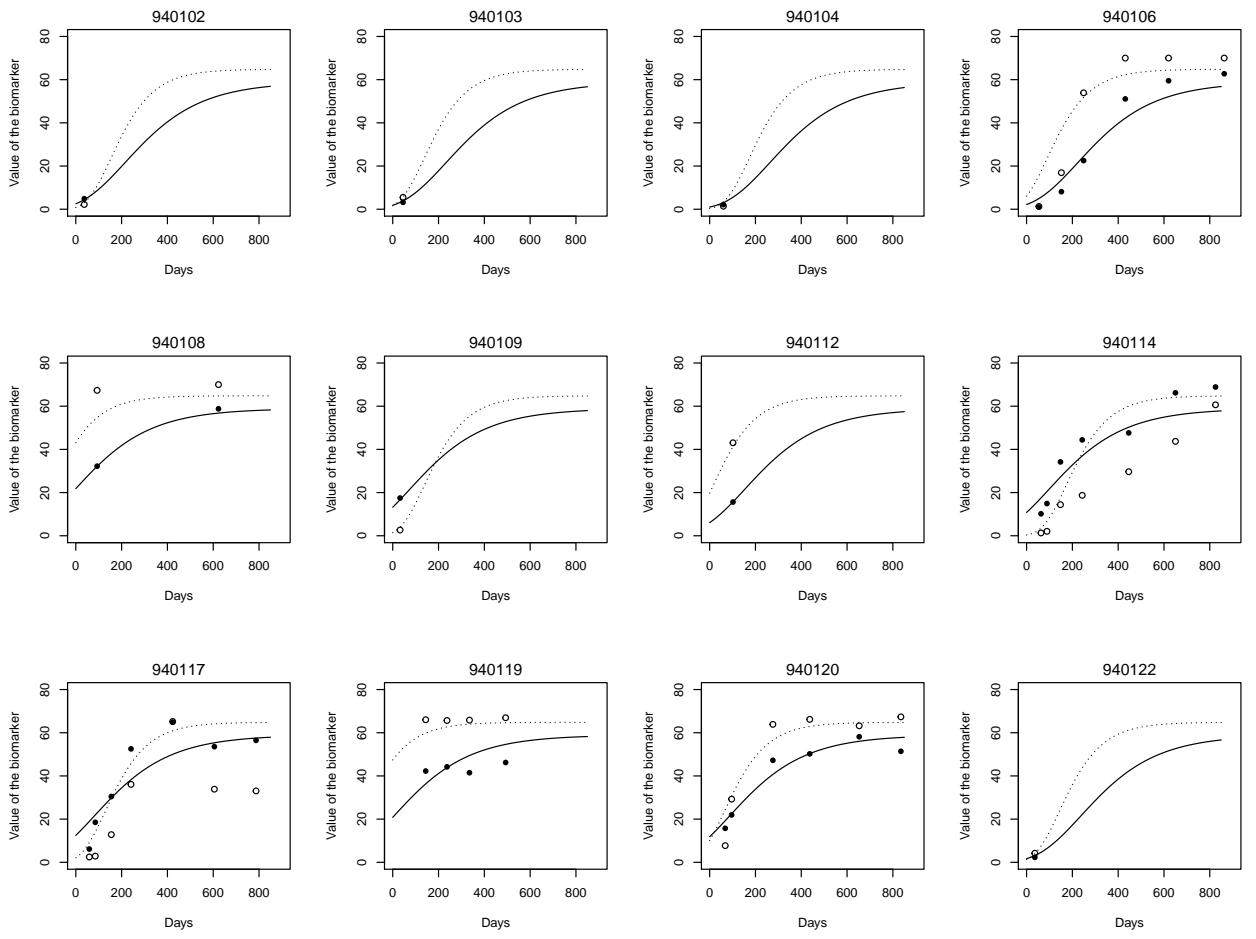


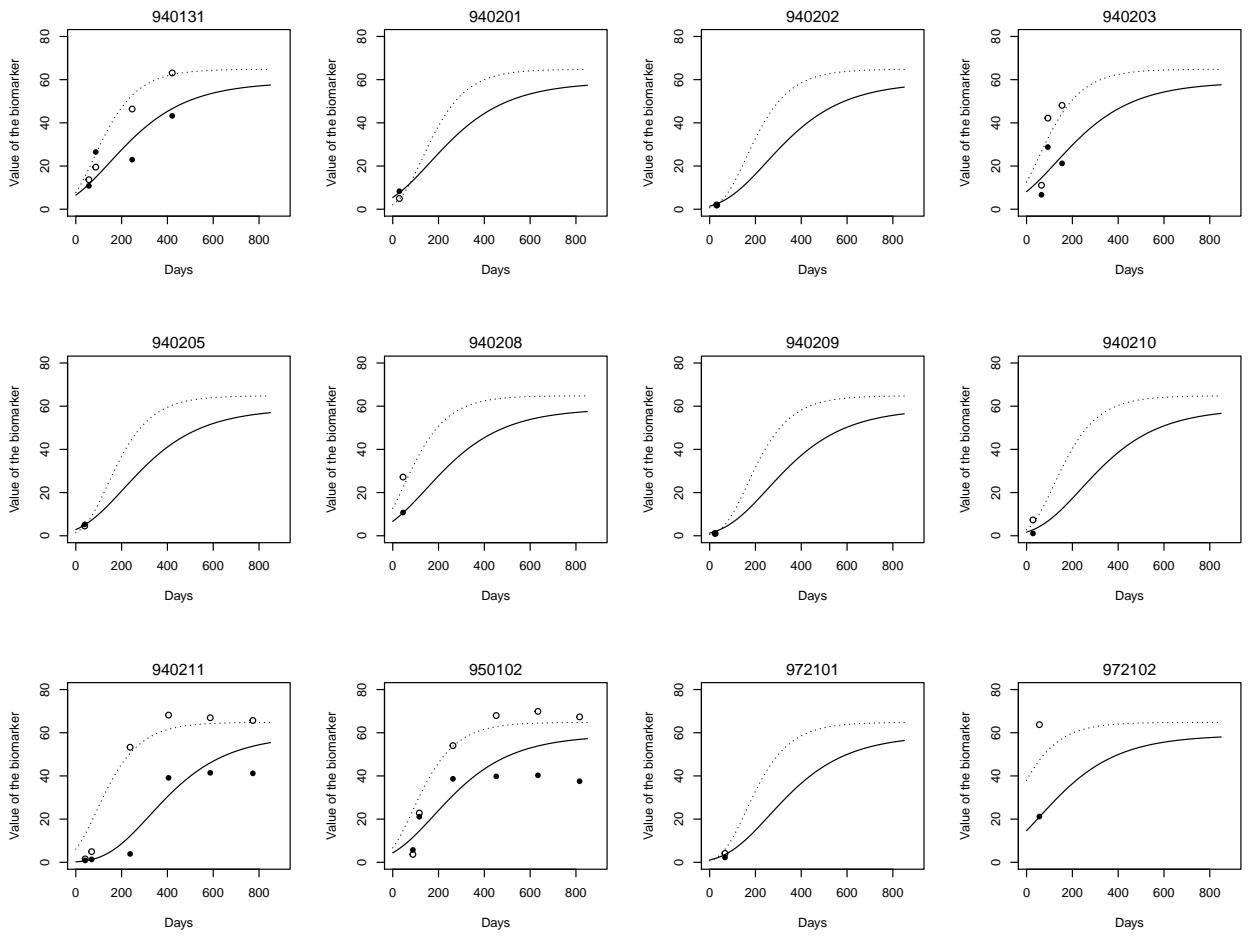


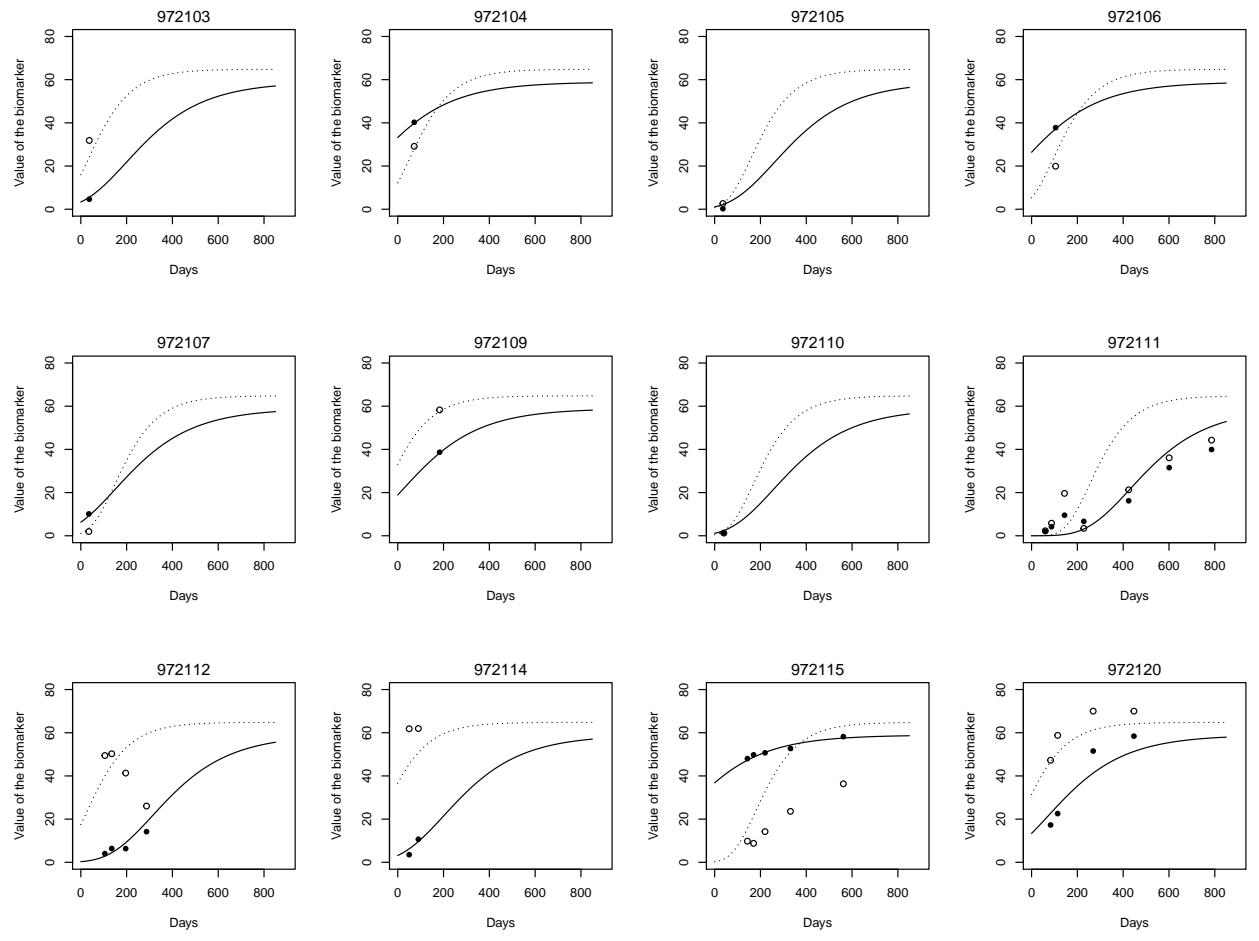


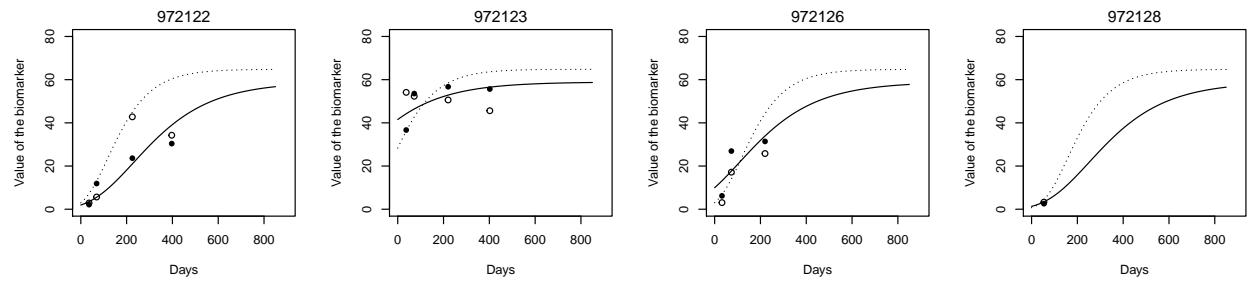












Values of the TM and V3 biomarkers for the four individuals who did not converge:

id	TM	V3	tps
220103	29.44	68.64	92
220103	36.20	69.38	126
220103	42.74	70.00	174
220103	41.26	70.00	266
220103	51.56	70.00	447
220103	51.64	58.48	629
220103	57.02	63.34	814

id	TM	V3	tps
751406	51.08	66.94	103
751406	54.24	70.00	133
751406	48.24	66.38	194
751406	53.32	70.00	271
751406	57.52	70.00	452
751406	63.74	70.00	634

id	TM	V3	tps
751617	49.10	70.00	115
751617	54.42	68.40	150
751617	55.44	70.00	262
751617	62.04	69.62	416
751617	70.00	70.00	878

id	TM	V3	tps
751903	42.14	67.14	77
751903	50.14	61.72	99
751903	63.60	70.00	165
751903	67.96	70.00	250
751903	70.00	70.00	432

Posterior density and posterior expectation value (vertical dotted line) of the infection time for all the individuals of the PRIMO ANRS-C06 cohort. The vertical line represents the date of infection of subjects in the PRIMO ANRS-C06 cohort

