Research Article Open Access

Sai Reddy Doda, * [a] Avula Raghavendar, [b] Sharath Babu Haridasyam, [b] Chandra Shekar Putta, [c] Bhattu Kanakadurga rao, [c] and Sudhakar Kadari * [c]

Asymmetric total synthesis of filamentous fungi related resorcylic acid lactones 7-epi-zeaenol and zeaenol

https://doi.org/10.1515/hc-2019-0015 Received October 30, 2018; accepted April 10, 2019.

Abstract: An efficient, short and, a convenient asymmetric total synthesis of filamentous fungi related resorcylic acid lactones 7-epi-zeaenol (2) and zeaenol (1) have been achieved in 7 and 9 linear steps with the high overall yield of 32% and 21% respectively, from the known intermediate **13.** Mitsunobu inversion, De Brabander's protocol for macrolactonisation, Heck cross-coupling, diastereoselective alkyne aldehyde coupling and Ohira–Bestmann alkynylation are the key reactions.

Keywords: Macrolactonisation, Natural products, Asymmetric synthesis, Heck cross coupling, C-C bond formation.

Introduction

Filamentous fungi possess an enormously rich source of biologically active natural products and have immense applications in various fields. Modern synthetic chemistry has enabled the identification, isolation, and manufacturing many of the promising biologically active filamentous fungi related natural products [1-3]. Included in this natural product family, highly oxygenated natural

products zeaenol and 7-epi-zeaenol (Figure 1) are isolated by Oberlies and co-workers [4,5] from a filamentous fungus, MSX 63935, collected from the leaf litter in Nigeria, along with other natural products. These resorcylic acid lactones (RALs) exhibit potent antibacterial activity as well as mitochondrial transmembrane potential activity. Of them, zeaenol (1) and 7-epi-zeaeneol (2) showed cytotoxic activity against human tumor cell lines [6,7] with nearly same IC_{50} values (IC_{50} values >50 μ M) and inhibition activity towards NF-kB. The first total synthesis of zeaenol along with its analog cochliomycin A was reported by Nanda et al [8] using late stage ring closing metathesis (RCM) strategy. Later, several synthetic procedures [9-13] for the total synthesis of zeaenol and 7-epi-zeaenol have been developed using diverse methods. In continuation of our efforts on the total synthesis of biologically active lactonic natural products [14-21], herein we described a first asymmetric total synthesis of zeaenol and 7-epi-zeaenol.

Figure 1. Structures of filamentous fungi derived resorcylic acid lactones (1-6).

e-mail: sudhakarou@osmania.ac.in

^{*} Corresponding author: Sai Reddy Doda, ^aNatural Products Chemistry Division, CSIR – IICT, Hyderabad 500007; India, e-mail: dsaireddyiict@gmail.com

^bDepartment of Chemistry GITAM University Hyderabad Campus, Rudrarar Village, Patancheru Mandal, Medak Dist, Telangana State - 502329, India

^cDepartment of Chemistry Osmania University, Hyderabad-500007 India.

³ Open Access. © 2019 S. R. Doda et al., published by De Gruyter. This work is licensed under the Creative Commons Attribution alone 4.0 License.

Result and discussion

The retrosynthetic analysis of zeaenol and 7-epi-zeaenol is depicted in Scheme 1. The target molecules 1 and 2 could be derived from the common fragment 7 following De Brabander's lactonization, which could be obtained by the aromatic fragment 8 and aliphatic fragment 9, using Heck cross coupling. The advanced fragment 9 could be traced back from the two individual fragments, aldehyde 10 and alkyne 11. The aldehyde 10 was commenced with D-mannitol.

Scheme 1. Retrosynthetic analysis of zeaenol and 7-epi-zeaenol

With the retrosynthetic blueprint in mind, our initial focus was on the synthesis of the alkyne fragment **11** from known aldehyde **12**. The known aldehyde **12** [22] was subjected to Ohira–Bestmann's protocol [23] with dimethyl(diazo-2-oxopropyl) phosphonate to give the propargyl alcohol **11** in 75% yield.

OTBS OME OTBS
$$N_2$$
 n_2 n_2 n_3 n_4 n_4 n_5 n_5 n_6 $n_$

Scheme 2. Reagents and conditions. (a) K2CO2 MeOH, RT, 8 h, 75%.

As planned, with the requisite alkyne fragment 11 and known aromatic fragment 8 [30], the synthesis of natural product 7-*epi*-zeaenol (2) was accomplished from a known intermediate 13, prepared from commercially available D-mannitol following a known protocol [24]. The dihydroxy functionality in 13 was protected as its benzyl, using benzyl bromide and NaH to afford compound 14 in 81% yield. The TBS group of 14 was deprotected smoothly using

camphorsulfonic acid (CSA) in MeOH vielding a primary alcohol, which upon further treatment with Dess-Martin periodinane reagent furnished aldehyde 10 in 83% yield in two steps. Furthermore, the aldehyde 10 was coupled with alkyne 11 in an asymmetric manner following the known protocol [25-28] using Et,Zn, Ti(OiPr), and (R)-BINOL to obtain chromatographically inseparable propargyl alcohol mixture 15. The resulting propargyl alcohol was then reduced with LiAlH, to trans allyl alcohol 16 (83% yield in two steps), which was then separable by column chromatography (dr:92:08 by HPLC). The secondary allyl alcohol obtained in the LiAlH, reduction was protected as its MOM-ether using MOM-Cl and DIPEA. The resulting MOM ether was subjected to Heck cross coupling [29] with the known aromatic fragment 8 [30] using Pd(OAc), catalyst and K₂CO₂ in DMF which resulted in the trans olefin 17 as a sole product with 88% yield in two steps. Now, the TBS deprotection of 17 was achieved smoothly with CSA in 89% yield. The macrolactonization of 7 was achieved under De Brabander's conditions [31] with NaH in 85% yield. Finally, the benzyl and MOM groups were deprotected in one-pot, by treatment of macrolactone 18 using excess TiCl, to afford the natural product, 7-epi-zeaenol, in 88% yield.

Synthesis of another natural product, zeaenol, commenced with compound **18**, beaing treated with TMSCl in methanol to obtain allyl alcohol **19** in 86% yield. The resulting chiral alcohol **19** was then inverted under Mitsunobu conditions [32,33] to afford alcohol **20** in 81% yield.

Scheme 3. Reagents and conditions. (a) BnBr, NaH, TBAI, THF 0 °C-rt 8 h, 81%; (b) CSA, MeOH 0 °C-rt, 12h; (c) DMP,CH₂Cl₂, rt, 3 h, 83% over two steps; (d) **11**, Et₂Zn, Ti(O'Pr)₄, (R)-BINOL, THF, rt, 12 h; (e)) LiAlH₄, THF 0 °C rt 12 h, 83% over two steps; (f) MOMCI, DIPEA, DMF, 0 °C-rt 12 h; (g) **8**, Pd(OAc)₂, Bu₄NBr, K₂CO₃, PPh₃, DMF, 80 °C, 5h, 88% over two steps (h) CSA, MeOH, rt, 1 h, 89%; (i) NaH, THF, rt, 4 h, 85%; (j) TiCl₃, DCM, 0 °C-rt, 2 h, 88%.

Scheme 4. Reagents and conditions :(a) TMSCl, MeOH, 12 h, 86%; (b) (i) 4-nitrobenzoic acid, Ph₃P, DEAD, rt 12 h; (ii) K_2CO_3 , MeOH, 50 °C, 10 h, 81%. over two steps; (c) TiCl₄, CH,Cl₇, 2 h,0 °C, 83%.

Similarly, upon treatment of alcohol **19** using excess TiCl₄, the natural product zeaenol (**1**) was obtained in 83% yield.

The spectroscopic (¹H and ¹³C NMR) and analytical data were in good agreement with the values reported values reported for the natural products [4,5].

Conclusion

In summary, we have reported a short and an efficient asymmetric total synthesis of filamentous fungi related resorcylic acid lactones 7-epi-zeaenol (2) and zeaenol (1) in 7 and 9 linear steps from an inexpensive known intermediate. The key steps involved in this synthesis are the Mitsunobu inversion, De Brabander's protocol for macrolactonisation, Heck cross-coupling, asymmetric alkyne aldehyde coupling, and Ohira–Bestmann alkynylation. The current methodology provides a new synthetic route with high yield and can be used further for the synthesis of various natural products of RAL's family.

Experimental Section

General remarks and methods: All reactions were performed under inert atmosphere. All glassware apparatus used for reactions were perfectly oven/flame dried. Anhydrous solvents were distilled prior to use: THF from Na and benzophenone; CH₂Cl₂, DMF from CaH₂; MeOH from Mg cake. Other commercial reagents were used without purification. Column chromatography was carried out by using silica gel (60–120 mesh) unless otherwise mentioned. Analytical thin layer chromatography (TLC) was performed on silica gel 60 F254 pre-coated plates (250 μm thickness). Optical rotation values were recorded on

Horiba sepa 300 polarimeter given in 10^{-1} degcm²g⁻¹. HRMS spectra were obtained using a TOF spectrometer. ¹H NMR and ¹³C NMR spectra were recorded using Bruker AVANCE 300 MHz, Varian INOVA 400 MHz, and Bruker AVANCE 500 MHz spectrometers in CDCl₃ solution unless otherwise mentioned. The chemical shifts values are in ppm downfield from tetramethylsilane (TMS) and coupling constants (*J*) were reported in hertz (Hz). The following abbreviations are used to designate signal multiplicity: s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet, br = broad.

(S)-tert-Butyldimethyl(pent-4-vn-2-vloxy)silane (11): To a stirred solution of aldehyde 13 (1.0 g, 5.3 mmol) and K₂CO₂ (2.2 g, 15.9 mmol) in methanol (30 mL), added a solution of dimethyl(diazo-2-oxopropyl) phosphonate (606 mg, 3.15 mmol) in methanol (15 mL) at room temperature and stirred for 8 hr at room temperature. Then the reaction mixture was filtered through celite and extracted with ethyl acetate (3 x 30 mL). The combined organics were washed with brine and dried over anhydrous Na₂SO₄. The solvent was removed under vacuum and the crude mass was purified by silica gel column chromatography (ethyl acetate/hexane = 1:20) to afford 11 (0.78 g, 75%) as a colorless liquid. $[\alpha]D^{25} + 0.67$ (c 2.5, CHCl₂). H NMR (300 MHz, CDCl₂) 8 3.96 (m, 1H), 2.36 (ddd, J = 16.6, 5.6, 2.7 Hz, 1H), 2.24 (ddd, J = 16.6, 7.1, 2.7 Hz, 1H),1.98 (t, J = 2.7 Hz, 1H), 1.23 (d, J = 6.1 Hz, 3H), 0.89 (s, 9H),0.08,(s, 3H) 0.07 (s, 3H) ppm; 13 C NMR (75 MHz, CDCl₂): δ 81.9, 69.7, 67.5, 29.4, 25.8, 23.2, 18.1, -4.7, -4.8 ppm; HRMS (ESI): calcd for for $C_{11}H_{22}ONaSi [M + Na] + 221.1333$, found: 221.1337.

(((2R,3S)-2,3-Bis(benzyloxy)hex-5-en-1-yl)oxy)(tert-butyl)dimethylsilane (14): To a suspension of NaH (60% in mineral oil, 3.9 g,97.8 mmol) in dry THF (60 mL), added a solution of alcohol 13 (7.5 g, 32.6 mmol) in THF (50 mL) at 0 °C. After 30 min, benzyl bromide (10.0 mL, 81.5 mmol) was added slowly at 0 °C. The resulting mixture was further stirred at room temperature for 8 h and guenched with water at 0 °C and extracted with ethyl acetate (2 x 200 mL). The combined organic layers were dried over anhydrous Na,SO, and solvent was removed under reduced pressure. The crude product was purified by silica gel column chromatography (ethyl acetate/hexane = 1:20) to give **14** (11.2 g, 81%) as a colorless liquid. $[\alpha]_{D}^{25}$ -4.3 (c 1.0, CHCl₂); ¹H NMR (300 MHz, CDCl₂) δ 7.42 - 7.15 (m, 10H), 5.83 (ddt, J = 17.1, 10.0, 7.0 Hz, 1H), 5.16 – 4.95 (m, 2H), 4.73 - 4.55 (m, 2H), 4.53 (s, 2H), 3.75 (qd, J = 10.8)4.8 Hz, 2H), 3.61 (q, J = 5.5 Hz, 1H), 3.53 (q, J = 4.9 Hz, 1H), 2.38 (t, J = 6.3 Hz, 2H), 0.85 (s, 10H), 0.17 – 0.19 (m, 6H); ¹³C NMR (75 MHz, CDCl₂) δ 138.9, 138.8, 135.5, 128.4, 128.1, 128.0, 127.9, 127.6, 127.6, 117.1, 80.9, 78.6, 72.9, 72.4, 62.9, 35.1,

26.1, 18.4, -5.2, -5.3; HRMS (ESI): calcd. for $C_{16}H_{10}O_{2}SiN$ [M + H]+427.2663 found: 427.2667.

(4S,5R,6R,10S,E)-4,5-Bis(benzyloxy)-10-((tertbutyldimethylsilyl)oxy)undeca-1,7-dien-6-ol (16): Et,Zn (17.6 mL, 17.6 mmol, 1.0 M solution) was added to a solution of alkyne 11 (3.5 g, 17.6 mmol) in toluene (20 mL) then the mixture was stirred under reflux. After 1 h the mixture was cooled to room temperature, thereafter (R)-BINOL (0.44 g, 1.6 mmol), THF (40 mL), and Ti(OiPr), (2.6 mL, 8.8 mmol) were added to the solution. After 1 h, aldehyde 10 (2.5 g, 8.0 mmol) was added under an argon atmosphere, and the mixture was stirred for overnight at room temperature. After completion of the reaction (monitored by TLC), the reaction mixture was quenched with 1.0 M tartaric acid (50 mL) and extracted with ethyl acetate (3 x 50 mL). The combined organic extracts were washed with brine and dried over Na, SO,. The solvent was removed under vacuum, which after purification by flash chromatography on silica gel (5% EtOAc/ hexanes to 20% EtOAc/ hexanes gradient), afforded mixture of propargylic alcohols 15 used in the next step without further characterization. LiAlH, (1.46 g, 12.0 mmol) was added to a stirred solution of above propargylic alcohol (8.0 mmol) in THF (30 mL) at 0 °C. The resulting solution was then stirred at RT for 12 hr. After completion of the reaction (indicated by TLC), it was cooled to 0 °C and quenched with saturated solution of Na₂SO₄ (50 mL). The solids were filtered through Celite pad and the filtrate was concentrated under reduced pressure. The crude product was purified by silica gel column chromatography (5-10% ethyl acetate/ hexane) furnished the desired alcohol 16 (3.4 g, 83%) as a color less liquid; $[\alpha]_D^{25}$ 11.3 (c 1.6, CHCl₃); ¹H NMR (300 MHz, CDCl₂) δ 7.44 – 7.31 (m, 10H), 6.01 – 5.68 (m, 2H), 5.59 (ddt, J = 15.5, 6.2, 1.3 Hz, 1H), 5.31 - 5.05 (m, 3H), 4.75 (d, J)= 11.3 Hz, 1H), 4.70 - 4.62 (m, 2H), 4.57 (d, J = 11.3 Hz, 1H),4.32 (q, J = 5.6 Hz, 1H), 3.92 - 3.77 (m, 1H), 3.77 - 3.69 (m, 1H)1H), 3.54 (dd, J = 5.8, 4.2 Hz, 1H), 3.00 (d, J = 6.3 Hz, 1H), $2.51 \text{ (tdd, } J = 7.2, 2.9, 1.4 \text{ Hz, } 2H), } 2.36 - 2.08 \text{ (m, } 2H), } 1.14 \text{ (d, }$ J = 6.1 Hz, 3H), 0.92 (d, J = 2.4 Hz, 9H), 0.08 (d, J = 1.7 Hz, 6H); ¹³C NMR (75 MHz, CDCl₂) δ 138.4, 138.0, 134.7, 131.2, 130.1, 128.5, 128.3, 128.2, 128.0, 127.8, 127.7 127.6, 127.5, 126.0, 117.4, 82.1, 79.6, 73.8, 73.7, 71.2, 68.5, 42.8, 34.8, 25.9, 23.4, 18.1, -4.5, -4.6; HRMS (ESI): calcd. for $C_{31}H_{47}O_{4}Si [M + H]^{+}$ 511.3239 found: 511.3246.

5-[(1E,4S,5S,6R,7E,10S)-4,5-bis(benzyloxy)-10-((tert-butyldimethylsilyl)oxy)-6-(methoxymethoxy) undeca-1,7-dien-1-yl)-7-methoxy-2,2-dimethyl-4Hbenzo[d][1,3]dioxin-4-one (17): To a stirred solution of compound 16 (0.5 g, 0,95 mmol)) in DMF (10 mL) was added diisopropyl ethylamine (0.31 mL, 1.9 mmol) followed by methoxymethyl chloride (0.11 mL, 1.4 mmol) at 0

°C. The resultant mixture was stirred at room temperature for additional 12 h. After completion of the reaction (monitored by TLC), the reaction mixture was quenched with water (20 mL) and extracted with ethyl acetate (3 x 30 mL). The combined organic layer was washed with water (2 x 50 mL) and dried over anhydrous Na₃SO₄ then removed under reduced pressure. The crude mass was purified by silica gel column chromatography (ethyl acetate/hexane = 1:19) to afford the MOM ether which was immediately used for the next reaction without further characterization.

To a stirred solution of aryl triflate 8 (0.507 g, 1.4 mmol), and the above MOM protected compound (0.95 mmol) in DMF (8 mL), Pd(OAc), (21 mg, 0.095 mmol), Bu, NBr (120 mg, 0.95 mmol), K, CO, (262 mg, 1.9 mmol) and PPh₃ (12 mg, 0.047 mmol) were added then the mixture was degassed three times with argon under vacuum and the resultant solution was stirred at 80 °C for 5 h. After completion of the reaction (monitored by TLC), the reaction mixture was filtered through Celite and washed with water (10 mL), then extracted with ethyl acetate (3 x 20 mL). The combined organics were washed with water and dried over Na, SO,. The solvent was removed, and the crude mass was purified by silica gel column chromatography (ethyl acetate/hexane = 1:20) to afford compound 17 (0.66 g, 88%) as a colorless oil. $[\alpha]_{D}^{25}$ –27.1 (*c* 1.6, CHCl₃); H NMR (400 MHz, CDCl₂) δ 7.56 (d, J = 15.7 Hz, 1H), 7.42 – 7.20 (m, 10H), 6.72 (d, J = 2.5 Hz, 1H), 6.35 (d, J = 2.5 Hz, 1H), 6.34 – 6.25 (m, 1H), 5.69 (ddd, J = 15.5, 7.7, 6.1 Hz, 1H), 5.53 (dd, J= 15.9, 8.2 Hz, 1H), 4.87 (d, J = 11.3 Hz, 1H), 4.78 – 4.72 (m, 2H), 4.64 (d, J = 11.4 Hz, 1H), 4.59 - 4.49 (m, 2H), 4.35 (dd, J = 8.2, 3.8 Hz, 1H, 3.83 (s, 3H), 3.79 – 3.68 (m, 2H), 3.37 (s, 3H), 2.75 (t, J = 6.9 Hz, 2H), 2.22 (tq, J = 14.0, 7.2 Hz, 2H), 1.72 (s, 6H), 1.28 (s, 6H), 1.12 (d, J = 6.1 Hz, 3H), 0.89 (s, 9H),0.06 (s, 6H); ¹³C NMR (75 MHz, CDCl₂) δ 164.7, 160.2, 158.7, 144.1, 138.8, 138.4, 133.0, 131.6, 130.2, 129.7, 128.5, 128.3, 128.2, 128.1, 128.0, 127.7, 127.5, 127.5, 108.3, 104.9, 103.5, 100.1, 93.4, 81.6, 78.6, 77.5, 74.0, 71.6, 68.3, 55.6, 55.6, 42.9, 33.5, 29.7, 25.9, 25.7, 25.6, 23.3, 18.1, -4.5, -4.7; HRMS (ESI): calcd. for $C_{44}H_{41}O_0NSi [M + H]^+$ 761.4080, found: 761.4083.

5-[(1E,4S,5S,6R,7E,10S)-4,5-Bis(benzyloxy)-10-hydroxy-6-(methoxymethoxy)undeca-1,7-dien-1-yl)-7methoxy-2,2-dimethyl-4H-benzo[d][1,3]dioxin-4-one (7): CSA (30.5 mg, 0.13 mmol) was added to a stirred solution of 17 (0.45 g, 0.1.3 mmol)) in 10 mL methanol and stirred for 1 hr at room temperature. After completion of the reaction (monitored by TLC), the solvent was removed, and the residue was dissolved in a saturated solution of NaHCO₂ (20 mL) and then extracted with ethyl acetate (3 x 25 mL). The organic layer was dried over anhydrous Na₃SO₄ and concentrated the crude mass by silica gel column chromatography (ethyl acetate/hexane = 2:3) and

obtained **7** (340 mg, 89%) as a colorless viscous liquid. $[\alpha]$ _D²⁵ –17.6 (*c* 1.0, CHCl₃); ¹H NMR (500 MHz, CDCl₂) δ 7.52 (d, *J* = 15.8 Hz, 1H), 7.41 - 7.22 (m, 10H), 6.70 (d, J = 2.7 Hz, 1H), 6.33 (d, J = 2.5 Hz, 1H), 6.25 (dt, J = 15.1, 7.1 Hz, 1H), 5.62 (dd, J = 15.1, 7.1 Hz, 1H), 5.62J = 6.1, 3.7 Hz, 2H, 4.84 (d, J = 11.4 Hz, 1H), 4.77 - 4.70 (m,2H), 4.63 (d, J = 11.6 Hz, 1H), 4.58 (q, J = 5.9, 5.5 Hz, 1H), 4.55 - 4.48 (m, 1H), 3.82 (s, 3H), 3.76 (m, 1H), 3.68 (d, J = 5.3Hz, 2H), 3.36 (s, 3H), 2.72 (q, J = 7.5 Hz, 2H), 2.31 - 2.09 (m, 2H), 1.70 (s, 6H), 1.17 (d, J = 6.4 Hz, 3H).; ¹³C NMR (75 MHz, CDCl₂) 8 164.7, 160.0, 158.7, 144.0, 138.6, 138.3, 131.9, 131.4, 130.3, 130.3, 128.3, 128.3, 128.2, 128.1, 128.0, 127.7, 127.6, 127.5, 108.3, 104.9, 100.4, 100.1, 93.9, 81.4, 78.5, 78.3, 73.9, 71.6, 67.0, 55.6, 42.1, 33.5, 25.7, 25.6, 22.8; HRMS (ESI): calcd. for $C_{20}H_{47}O_{0}$ [M + H]+647.3215, found: 647.3211.

(3S,5E,7R,8S,9S,11E)-8,9-Bis(benzyloxy)-16hydroxy-14-methoxy-7-(methoxymethoxy)-3methyl-3,4,7,8,9,10-hexahydro-1H-benzo[c][1] oxacyclotetradecin-1-one (18): To a suspension of NaH (0.15 g, 3.7 mmol) washed with hexane twice to remove mineral oil and dried in dry THF (10 mL) was added a solution of alcohol 7 (0.3 g, 0.46 mmol) in THF (5 mL) at 0 °C under argon and the suspension was stirred for 5 h at room temperature. After completion of the reaction (monitored by TLC), the reaction mixture was quenched with ice pieces at 0 °C and extracted with ethyl acetate (3 x 10 mL). The combined organic layer was dried over anhydrous Na₂SO₄ and solvent was removed under reduced pressure and purified by silica gel column chromatography (ethyl acetate/hexane = 1:9) to afford 18 (0.27 mg, 85%) as a colorless liquid. [α]_D²⁵ –55.8 (*c* 1.2, CHCl₂) ¹H NMR (500 MHz, CDCl₂) δ 7.31 – 7.17 (m, 10H), 7.09 (d, J = 15.4 Hz, 1H), 6.38 (d, J = 2.5 Hz, 1H), 6.35 (d, J = 2.5 Hz, 1H), 5.89 (s, 1H),5.79 (dd, J = 15.6, 9.3 Hz, 1H), 5.61 (ddd, J = 15.6, 8.1, 3.5 Hz,1H), 5.16 - 5.07 (m, 1H), 4.76 (d, J = 33.0 Hz, 2H), 4.62 (t, J= 10.3 Hz, 1H), 4.47 (dd, J = 22.5, 8.9 Hz, 3H), 4.01 (m, 1H), 3.92 (m, 3H), 3.81 (s, 3H), 3.32 (s, 3H), 2.70 (dt, J = 18.5, 9.3)Hz, 1H), 2.65 - 2.53 (m, 3H), 2.44 - 2.27 (m, 1H), 1.42 (d, J =6.0 Hz, 3H); ¹³C NMR (126 MHz, CDCl₂) δ 171.7, 165.0, 164.0, 143.6, 139.0, 133.3, 132.3, 128.4, 128.2, 127.8, 127.6, 127.6, 127.3, 107.4, 99.8, 92.8, 83.6, 81.3, 78.5, 73.4, 73.2, 71.9, 55.4, 55.3, 38.7, 35.0, 29.7, 20.5; HRMS (ESI): calcd. for C₃₅H₄₄O₈ [M + H] +589.2878, found: 589.2881.

7-epi-zeaenol (2): TiCl_a (1.1 mL, 1.1 mmol, 1 M in CH₂Cl₂) was added to a stirred solution of **18** (35 mg, 0.06 mmol) in CH₂Cl₂ (5 mL) at 0 °C, and the mixture was stirred for 1 hr for 30 min at 0 °C. After completion of the reaction (monitored by TLC), the reaction mixture was quenched with a saturated solution of NaHCO₂ (10 mL), extracted with CH₂Cl₂ (3 x 25 mL) and washed with brine (20 mL). The organic layer was dried over anhydrous Na₂SO₄ and concentrated to afford a yellowish liquid, which was purified

by silica gel column chromatography (acetone/hexane 1:1) to obtain compound 2 (17.3 mg, 88%) as a white powder. $[\alpha]_{D}^{25}$ –91 (c 1.5, MeOH); ¹H NMR (500 MHz, DMSO-d_s) δ 10.57 (s, 1H), 6.62 (d, J = 15.5 Hz, 1H), 6.43 (d, J = 2.28 Hz, 1H), 6.30 (d, I = 2.27 Hz, 1H), 6.1 (m, 1H), 5.66-5.53 (m, 2H), 5.19 (m, 1H), 4.82 (s, 1H), 4.80 (s, 1H), 4.50 (d, J = 4.1 Hz, 1H), 4.05 (m, 1H), 3.75 (s, 3H), 3.60 (m, 1H), 3.35 (m, 1H), 2.48-2.32 (m, 3H), 2.17 (m, 1H), 1.33 (d, J = 5.9 Hz, 3H) ppm; ¹³C NMR (126 MHz, DMSO) δ 169.0, 161.7, 159.0, 139.6, 133.1, 132.5, 128.6, 127.0, 110.3, 102.7, 99.9, 77.5, 74.7, 72.9, 71.8, 55.2, 38.6, 36.8, 29.6, 20.4. HRMS (ESI): calcd. for C₁₀H₂₆O₂Na [M + Na]+387.1415, found: 387.1419.

(3S,5E,7R,8R,9S,11E)-8,9-Bis(benzyloxy)-7,16-dihydroxy-14-methoxy-3-methyl-3,4,7,8,9,10-hexahydro-1H-benzo[c][1]oxacyclotetradecin-1-one (19): TMSCl (0.056 mL, 0.45 mmol) was added to stirred solution of 18 (0.175 mg, 0.297 mmol) in MeOH (5 mL) and allowed to stir for 2 hrs at room temperature. After completion of the reaction as indicated by TLC, the methanol was removed and the residue was dissolved in a saturated solution of NaHCO₂ (10 mL) then extracted with ethyl acetate (3 x 15 mL), dried over anhydrous Na₂SO₄ and concentrated under reduced pressure and then purified by silica gel column chromatography (ethyl acetate/hexane = 1:3) to give **19** (134 mg, 86%) as a colorless liquid. $[\alpha]_n^{25}$ -76.0 (c 1.1, CHCl₃) ¹H NMR (500 MHz, C₃D₆O-d₆): 7.48-7.16 (m, 11H), 6.46 (s, 1H), 6.39 (s, 1H), 5.99 (m, 1H), 5.89 (m, 1H), 5.72 (m, 1H), 5.21 (m, 1H), 4.87-4.61 (m, 4H), 4.57 (m, 1H), 4.27 (m, 1H), 3.91-3.82 (m, 4H), 2.79 (m, 1H), 2.64-2.57 (m, 3H), 1.43 (d, J = 5.9 Hz, 3H) ppm; ¹³C NMR (126 MHz, CDCl₂) 8 171.6, 165.1, 164.2, 143.6, 138.9, 128.1, 128.0, 127.7, 127.5, 127.2, 106.8, 104.0, 99.7, 83.3, 78.3, 73.2, 55.0, 38.2, 19.5.HRMS (ESI): calcd. for $C_{22}H_{27}O_{7}$ [M + H]⁺ 545.2534, found: 545.2541.

(3S,5E,7S,8R,9S,11E)-8,9-Bis(benzyloxy)-7,16-dihydroxy-14-methoxy-3-methyl-3,4,7,8,9,10-hexahydro-1*H*-benzo[c][1]oxacvclotetradecin-1-one (20): A solution of alcohol **19** (0.12 g, 0.22 mmol), p-nitrobenzoic acid (73 mg, 0.44 mmol), and TPP (86 mg, 0.33 mmol) in THF (15 mL) at 0 °C were treated with diisopropyl azodicarboxylate (66.6 g, 0.33 mmol) and the contents were stirred at 0 °C for 1 hr and then at RT for 12 h. After completion of the reaction, the reaction mixture was concentrated, and the resulting crude material was dissolved in ethyl acetate (60 mL), and subsequently washed with the aqueous NaHCO₃ (30 mL), water (50 mL), dried over Na, SO, and concentrated under vacuum. Purification of the residue by silica gel column chromatography (25% EtOAc in hexanes) yielded an ester which is not stable and immediately used in the next step. To a solution of above ester (0.22 mmol) in MeOH (5 mL) was added KOH (24 mg, 0.44 mmol) and the reaction

mixture is stirred for 10 hours at RT. After completion of the reaction, as indicated by TLC the solvent was removed under vacuum and the crude material was diluted with water (10 mL), then extracted with ethyl acetate (3 x 20 mL) the combined organic layer was dried over Na₂SO₄ and concentrated under the vacuum. Purification of the residue by silica gel column chromatography gave compound 20 (96 mg, 81%) as a colorless liquid. [α]_D²⁵ –61.3 (c 0.5, CHCl₂); ¹H NMR (500 MHz, $C_3D_6O-d_6$): δ 7.50-7.13 (m, 11H), 6.47 (s, 1H), 6.40 (s, 1H), 6.10 (m, 1H), 6.00 (m, 1H), 5.81 (m, 1H), 5.31 (m, 1H), 4.97-4.45 (m, 4H), 4.34 (m, 1H), 3.95 (m, 1H), 3.90 (s, 3H), 3.66 (m, 1H), 2.65-2.47 (m, 4H), 1.44 (d, J = 6.2Hz, 3H); ¹³C NMR (176 MHz, CDCl₂) δ 171.63, 165.3, 164.2, 143.8, 139.7, 139.2, 132.2, 132.0, 131.5, 128.2, 127.8, 127.7, 127.2, 126.7, 106.8, 103.8, 99.6, 83.0, 82.0, 73.3, 72.5, 71.0, 55.0, 37.2, 34.1, 18.8; HRMS (ESI): calcd. for $C_{33}H_{37}O_{7}[M + H]+545.2534$, found: 545.2531.

zeaenol (1): TiCl₄ (1.1 mL, 1.1 mmol, 1M in CH,Cl₅) was added to a stirred solution of 20 (30 mg, 0.05 mmol) in CH₂Cl₂ (5 mL) at 0 °C and the mixture was stirred for 1.45 hr at 0 °C. After completion of the reaction as indicated by TLC, the reaction mixture was guenched with a saturated solution of NaHCO₃ (10 mL), extracted with CH₂Cl₃ (3 x 20 mL) and washed with brine (10 mL), dried over anhydrous Na, SO, and concentrated. The crude mass was that purified by silica gel column chromatography (5% methanol in chloroform) gave zeaenol (1) (16.6 mg, 83%) as a white powder. $\left[\alpha\right]_{D}^{25}$ -87 (c 0.9, MeOH); ¹H NMR (500 MHz, CDCl₂): δ 11.72 (m, 1H), 7.15 (d, J = 15.3 Hz, 1H), 6.47 (d, J = 2.3, 1H), 6.42 (d, J = 2.3 Hz, 1H), 6.08 - 5.95 (m, 1H),5.90 - 5.80 (m, 1H), 5.74 (dd, J = 15.5, 7.4 Hz, 1H), 5.36 (d, J= 3.6 Hz, 1H), 4.30 (d, J = 7.3 Hz, 1H), 4.01 (t, J = 6.8 Hz, 1H),3.62 (d, J = 7.7 Hz, 1H), 2.61 - 2.49 (m, 3H), 2.49 - 2.41 (m, 3H)1H), 1.49 (d, J = 6.2 Hz, 3H).); ¹³C NMR (125 MHz, CDCl₂): δ ¹³C NMR (126 MHz, None) δ 171.2, 165.3, 164.0, 142.9, 133.7, 131.5, 129.2, 128.5, 107.6, 103.9, 100.1, 73.1, 71.96, 71.5, 55.4, 37.8, 36.0, 29.3, 19.7; HRMS (ESI): calcd. for $C_{10}H_{24}O_7Na$ [M + Na]+387.1415, found: 387.1421

Acknowledgements: KSR thanks the DST-SERB (Young scientist File: CS-138/2012), New Delhi.

References

- [1] Bhadury. P.; Mohammad, B. T.; Wright, P.C. The current status of natural products from marine fungi and their potential as anti-infective agents. J. ind. Microbiol. Biotechnol. 2006, 33, 325.
- [2] Hoffmeister, D.; Kellerb, N. P. Natural products of filamentous fungi: enzymes, genes, and their regulation. Nat. Prod. Rep., 2007,24, 393-416.

- [3] Fabrizio.; Foster, G. D.; Bailey, A. M. Natural products from filamentous fungi and production by heterologous expression. Appl Microbiol Biotechnol. 2017, 101, 493-500.
- [4] Ayers, S.; Graf, T. N.; Adcock, A. F.; Kroll, D. J.; Matthew, S.; Carcache deBlanco, E. J.; Shen, Qi.; Swanson, S. M.; Wani, M. C.; Pearce, C. J.; Oberlies. N. H. Resorcylic Acid Lactones with Cytotoxic and NF-kB Inhibitory Activities and Their Structure-Activity Relationships. J. Nat. Prod. 2011, 74, 1126-1131.
- Sugawara, F.; Kim, K. W.; Kobayashi, K.; Yoshida, J. S.; Murfushi, N.; Takahashi N.; Strobel, G. A. Zearalenone derivatives produced by the fungus Drechslera portulacae. Phytochemistry 1992, 31,1987-1990.
- Alali, F. Q.; El-Elimat, T.; Li, C.; Qandil, A.; Alkofahi, A.; Tawaha, K.; Burgess, J. P.; Nakanishi, Y.; Kroll, D. J.; Navarro, H. A.; Falkinham, I. O.: Wani M. C.: Oberlies, N. H. New Colchicinoids from a Native Jordanian Meadow Saffron, Colchicum brachyphyllum: Isolation of the First Naturally Occurring Dextrorotatory Colchicinoid. J. Nat. Prod. 2005, 68, 173-178.
- [7] Li, C.; Lee, D.; Graf, T. N.; Phifer, S. S.; Nakanishi, Y.; Riswan, S.; Setyowati, F. M.; Saribi, A. M.; Soejarto, D. D.; Farnsworth, N. R.; Falkinham, J. O.; Kroll, D. J.; Kinghorn, A. D.; Wani, M. C.; Oberlies, N. H. Bioactive Constituents of the Stem Bark of Mitrephora glabra. J. Nat. Prod. 2009, 72, 1949-1953.
- Jana, N.; Nanda, S. Asymmetric Total Syntheses of Cochliomycin A and Zeaenol. Eur. J. Org. Chem. 2012, 4313-4320.
- [9] Gao, Y.; Liu, J.; Wang, L.; Xiao, M.; Du, Y. Total Syntheses of Cochliomycin B and Zeaenol. Eur. J. Org. Chem. 2014, 2092-2098.
- [10] Mohapatra, D. K.; Reddy, D. S.; Mallampudi, N. A.; Gaddam, J.; Polepalli, S.; Jainb, N.; Yadav, J. S. The protecting-group directed diastereoselective Nozaki-Hiyama-Kishi (NHK) reaction: total synthesis and biological evaluation of zeaenol, 7-epi-zeaenol and its analogues. Org. Biomol. Chem. 2014, 12, 9683-9695.
- [11] Pal, P.; Chakraborty, J.; Mali, A.; Nanda, S. Asymmetric total synthesis of paecilomycin F, cochliomycin C, zeaenol, 5-bromozeaenol and 3,5-dibromo-zeaenol by Heck coupling and late stage macrolactonization approach. Tetrahedron 2016, 72, 2336-2348.
- Kumar, R. N.; Meshram, H.M. Convergent and stereoselective synthesis of (-)-zeaenol. Tetrahedron 2015, 71, 5669-5677.
- [13] Bolte, B.; Basutto, J. A.; Bryan, C. S.; Garson, M. J.; Banwell, M. G.; Ward. J. S. Modular Total Syntheses of the Marine-Derived Resorcylic Acid Lactones Cochliomycins A and B Using a Late-Stage Nozaki-Hiyama-Kishi Macrocyclization Reaction. J. Org. Chem. 2015, 80, 460-470.
- [14] Sabitha, G.; Sudhakar, K.; Sriinvas, Ch.; Yadav, J. S. Stereoselective Synthesis of a Mevinic Acid Analogue. Synthesis 2007, 5, 705-708;
- [15] Mohapatra, D. K.; Das, P. P.; Reddy, D. S.; Yadav, J. S. First total syntheses and absolute configuration of rugulactone and 6 (R)-(4'-oxopent-2'-enyl)-5, 6-dihydro-2H-pyran-2-one. Tetrahedron Lett. 2009, 50, 5941-5944.
- [16] Reddy, D. S.; Kutateladze, A. G.; Structure Revision of an Acorane Sesquiterpene Cordycepol A. Org. Lett. 2016, 18, 4860-4863.
- [17] Reddy, D. S.; Kutateladze, A. G.; Computational structure revision of a longipinane derivative meridane. Tetrahedron Lett. 2016, 57, 4727-4729.

- [18] Kutateladze, A. G.; Reddy, D. S.; High-Throughput in Silico Structure Validation and Revision of Halogenated Natural Products Is Enabled by Parametric Corrections to DFT-Computed 13C NMR Chemical Shifts and Spin-Spin Coupling Constants. J. Org. Chem. 2017, 82, 3368-3381.
- [19] Mohapatra, D. K.; Reddy, D. S.; Ramaiah, M. J.; Ghosh, S.; Pothula, V.; Lunavath, S.; Thomas, S.; Valli. S. N.; Bhadra, M. P.; Yadav, J. S. Rugulactone derivatives act as inhibitors of NF-κB activation and modulates the transcription of NF-κB dependent genes in MDA-MB-231cells. Bioorg. Med. Chem. Lett Bioorg. Med. Chem. Lett. 2014, 24,1389-1396.
- [20] Mohapatra, D. K.; Reddy, D. S. Reddy, G. S.; Yadav, J. S. Synthesis of the C-8-C-24 Fragment of Maltepolide C by Using a Tandem Dihydroxylation/SN2 Cyclization Sequence. Eur. J. Ora. Chem. 2015, 5266-5274.
- [21] Reddy, D. S.; Mukhina, O. A.; Cronk, W. C.; Kutateladze, A.G. Polyheterocycle-carbohydrate chimeras: photoassisted synthesis of 2,5-epoxybenzoxacines and 2,5-epoxybenzazocine scaffolds and their postphotochemical hydroxylations. Pure Appl. Chem. 2017, 89, 259-268.
- [22] David, L.; Rainer, S. Synthesis of the fungus metabolite cladosin C. Org. Biomol. Chem., 2017,15, 7672-7677.
- [23] Müller, S.; Liepold, B.; Roth, G. J.; H. Bestmann, J. An Improved One-pot Procedure for the Synthesis of Alkynes from Aldehydes. Synlett 1996, 521-522.
- [24] Mohapatra, D. K.; Reddy, DS.; Mallampudi, N. A.; Yadav, J. S. Stereoselective Total Syntheses of Paecilomycins E and F through a Protecting Group Directed Diastereoselective Intermolecular Nozaki-Hiyama-Kishi (NHK) Reaction Eur. J. Org. Chem. 2014, 5023-5032.

- [25] Marshall, J. A.; Bourbeau, M. P. Synthesis of Enantioenriched Propargylic Alcohols Related to Polyketide Natural Products. A Comparison of Methodologies. Org. Lett. 2003, 5, 3197-3199.
- [26] Anand, N. K.; Carreira, E. M. A Simple, Mild, Catalytic, Enantioselective Addition of Terminal Acetylenes to Aldehydes. J. Am. Chem. Soc. 2001, 123, 9687.
- [27] Moore, D.; Pu, L. BINOL-Catalyzed Highly Enantioselective Terminal Alkyne Additions to Aromatic Aldehydes. Org. Lett. 2002, 4, 1855.
- [28] Balsells, J.; Davis, T. J.; Carroll, P.; Walsh, P. J. Insight into the Mechanism of the Asymmetric Addition of Alkyl Groups to Aldehydes Catalyzed by Titanium-BINOLate Specie. J. Am. Chem. Soc. 2002, 124, 10336.
- [29] Mikula, H.; Skrinjar, P.; Sohr, B.; Ellmer, D.; Hametner, C.: Frohlich, I. Total synthesis of masked Alternaria mycotoxins—sulfates and glucosides of alternariol (AOH) and alternariol-9-methyl ether (AME). Tetrahedron 2013, 69, 10322-10330.
- [30] Srihari, P.; Mahankali, B.; Rajendraprasad, K. Stereoselective total synthesis of paecilomycin E. Tetrahedron Lett. 2012, 53,
- [31] Bhattacharjee, A.; Sequil, O. R.; De Brabander, J. K. Synthesis of side chain truncated apicularen A. Tetrahedron Lett. 2000, 41, 8069-8073.
- [32] Mitsunobu, O. The Use of Diethyl Azodicarboxylate and Triphenylphosphine in Synthesis and Transformation of Natural Products. Synthesis 1981, 1.
- [33] Mitsunobu, O.; Jenkins, I. D., in Encyclopedia of Reagents for Organic Synthesis; Paquette, L. A., Ed.; Wiley: New York, 1995; 8,5379.