Research Article Open Access

Roberto Martínez*, Gladys J. Nieves Zamudio, Gustavo Pretelin-Castillo, Rubén O. Torres-Ochoa, José L. Medina-Franco, Clara I. Espitia Pinzón, Mayra Silva Miranda, Eugenio Hernández and Blanca Alanís-Garza

Synthesis and antitubercular activity of new *N*-[5-(4-chlorophenyl)-1,3,4-oxadiazol-2-yl]- (nitroheteroaryl)carboxamides

https://doi.org/10.1515/hc-2019-0007 Received October 30, 2018; accepted November 19, 2018.

Abstract: Nitro-substituted heteroaromatic carboxamides **1a-e** were synthesized and tested against three *Mycobacterium tuberculosis* cell lines. The activities can be explained in terms of the distribution of the electronic density across the nitro-substituted heteroaromatic ring attached to the amide group. **1,3,5-Oxadiazole** derivatives **1c-e** are candidates for the development of novel antitubercular agents. Ongoing studies are focused on exploring the mechanism by which these compounds inhibit *M. tuberculosis* cell growth.

* Corresponding author: Roberto Martínez, Instituto de Química, Universidad Nacional Autónoma de México, Circuito Exterior, Ciudad Universitaria, 04510, Cd. México, México, e-mail: robmar@unam.mx

Gladys J. Nieves Zamudio, Gustavo Pretelin-Castillo and Rubén O. Torres-Ochoa, Instituto de Química, Universidad Nacional Autónoma de México, Circuito Exterior, Ciudad Universitaria, 04510, Cd. México, México

José L. Medina-Franco, Departamento de Farmacia, Facultad de Química, Universidad Nacional Autónoma de México, Avenida Universidad 3000, 04510 Cd. México, México

Clara I. Espitia Pinzón, Instituto de Investigaciones Biomédicas, Universidad Nacional Autónoma de México, Ciudad Universitaria, 04510, Cd. México, México

Mayra Silva Miranda, Catedrática CONACYT adscrita al Insituto de Investigaciones Biomédicas, Universidad Nacional Autónoma de México, Ciudad Universitaria, 04510, Cd. México, México

Eugenio Hernández, Facultad de Ciencias Químicas, Universidad Autónoma de Nuevo León, Pedro de Alba s/n, Ciudad Universitaria, 66400 San Nicolás de los Garza, Nuevo León, México

Blanca Alanís-Garza, Departamento de Química Analítica, Facultad de Medicina, Universidad Autónoma de Nuevo León, Madero s/n Col. Mitras Centro. Monterrey, N. L. México C. P. 64460

Keywords: antitubercular agents; carboxamides; 1,3,4-oxadiazoles; synthesis.

Introduction

Tuberculosis (TB) is a one of the top causes of death worldwide, and in 2017 the World Health Organization reported 10 million new cases of TB, 1.6 million deaths due to TB and 0.3 million deaths resulting from co-infections by HIV [1]. Although the rate of decline in TB was 3.9 % from 2015 to 2017, there were 457,560 new cases of multidrug-resistant TB (MDR-TB) and 558,000 people with rifampicin-resistant TB (RR-TB). Therefore, TB poses a challenge to researchers searching for potent drugs that can control the growth of the bacillus Mycobacterium tuberculosis while minimizing side effects or the development of drug resistance [2]. Excellent reviews have been published covering the literature between 2000 and 2015 about compounds with activity against M. tuberculosis [3-5]. Notably, Kumar and co-workers grouped anti-TB compounds into 62 different molecular frameworks. In their most recent compilation (2005-2015), they only included compounds exhibiting minimum inhibitory concentration (MIC) values similar to or higher than those of standard drugs used in bioassays [6], with 12 different oxadiazoles meeting this criterion. The 1,3,4-oxadiazole ring system indeed exerts a wide variety of pharmacological activities such as anti-inflammatory [7], antiviral [8], antineoplastic [9], FAK inhibitory [10], adulticidal [11] and anti-Alzheimer properties [12]. A review of the literature up to 2017 includes 440 citations of 1,3,4-oxadiazoles with biological activity but only 34 of these citations describe 1,3,4-oxadiazoles with anti-TB activity [13]. A 5-nitrofuranyl scaffold is present in many derivatives with diverse biological activities including anti-inflammatory [14, 15], antibacterial [16], antileishmanial [17] and antitubercular agents [18-21]. It has been proposed that the

³ Open Access. © 2019 Roberto Martínez et al., published by De Gruyter. © This work is licensed under the Creative Commons Attribution alone 4.0 License.

presence of the nitro group increases the ability of a molecule to form hydrogen bonds with receptor [22].

We have recently reported the synthesis of caulerpin, a bis-indole alkaloid from the marine alga Caulerpa sp. that displays excellent biological activity [23]. Nevertheless, the conventional synthetic approach to medicinal chemistry is a time-consuming, complicated and expensive process that produces candidate compounds with low diversity. Although this approach is still valuable, it is unable to fulfill the increasing demand for new drugs. Therefore, experimental drug discovery is currently being aided with chemo-informatics approaches that are frequently used to identify active compounds, select candidates, and optimize leads. Chemo-informatic approaches has been promising over the past few years in finding pharmacologically active compounds across a broad range of therapeutic areas [24]. Katsuno and co-workers published a comprehensive summary of the criteria to consider when developing new drugs against infectious diseases such as TB [25], and in our work we consider criteria such as selectivity

index (SI>10), MIC (lower than 10 µM) and activity against drug-resistant strains, to guide our screening of new compounds having potential use as anti-TB drugs. In the present study, we report a new compound active against M. tuberculosis, selected by carrying out a structural analysis of a major public database of compounds reported to have TB activity. We then used the bioisosterism concept [26] to generate a list of other possible compounds (Figure 1) which we subsequently synthesized and tested as inhibitors of the growth of *M. tuberculosis*.

Results and discussion

We conducted a chemotype analysis of compounds in PubChem [27] with reported activity against M. tuberculosis. To this end, the chemotype methodology developed by Johnson and Xu [28] to identify promising molecular scaffolds as starting points for optimization was used. In the approach of Johnson and Xu, each chemotype is assigned

$$\begin{array}{c} N-N \\ N-N \\ N+N \\$$

Figure 1 Designed compounds based on isosteric modification of lead compound 1a.

a unique alphanumeric identifier of four characters [29]. The same approach was previously published to identify promising scaffolds for anti-AIDS activity [30]. Based on the substructure search, the molecular scaffold of compound **1a** (compound identifier, CID in PubChem 4225334) was selected (chemotype identifier BKQ4R). We also compared the newly designed compounds with the reference molecule **1a**. The high structure similarity suggested that their biological activity would be also comparable (Figure 1). This assumption was made considering that here are no activity cliffs, specifically, no similar compounds with very different biological activity [31]. Physical and spectroscopic data for compound **1a** have not yet been published.

The preparation of compounds **1a-e** started with esterification of 4-chlorobenzoic acid (**2**, Scheme 1), with MeOH in the presence of sulfuric acid to provide methyl 4-chlorobenzoate (**3**). Next, the 4-chlorophenylhydrazide **4** was prepared from **3** [32]. Treatment of **4** with cyanogen bromide (CNBr) in MeOH afforded 5-(4-chlorophenyl)-1,3,4-oxadiazol-2-amine (**5**) in 62% yield [33]. The coupling reaction between **5** and crude acyl chlorides **6a-e** was achieved in the presence of sodium hydride in dry THF to afford the desired products **1a-e** in 30-62% yield [34].

The activities of compounds **1a-e** as inhibitors of the growth of the *M. tuberculosis* strains H37Rv and 209 were evaluated using 250 μ g/mL-0.98 μ g/mL serial two-fold dilutions. Rifampin (RIF) is usually used for the treatment

of mycobacterial infections, including TB, and the anti-TB activities of the synthesized compounds **1a-e** were compared with the activity of RIF. As shown in Table 1, RIF shows the MIC_{100} value of 0.06 µg/mL and is hence more active than compounds **1a-e**. Of compounds **1a-e**, **1a** shows the most activity, with the MIC_{100} value of 7.8 µg/mL (23.45 µM), followed by **1b**, **1d** and **1e**, each of them with MIC_{100} of 15.6 µg/mL, followed by **1c**. According to the criteria for

Table 1 Anti-M. tuberculosis effects and cytotoxicity levels of compounds 1a-e.

Compound	R	MIC ₁₀₀ (µg/mL)	MBC (µg/mL)	IC ₅₀ , in Vero cells (µg/mL)	Selectivity index (SI)
1a	5-NO ₂ C ₄ H ₂ O	7.80	2.00	106.0	13.59
1b	$5\text{-NO}_2C_4H_2S$	15.60	15.60	36.0	4.61
1c	$5\text{-NO}_2C_4H_3N$	31.25	31.25	68.3	2.18
1d	$3-NO_2C_6H_4$	15.60	3.90	61.1	3.92
1e	5-C ₆ H ₅	15.60	31.25	44.1	2.83
Rifampin	-	0.06	ND	>1000	>16,666

M. tuberculosis H37Rv ATCC 27294 reference strain: M. tuberculosis. MIC: minimal inhibition concentration determined by using REMA. IC_{50} : inhibitory concentration at 50% determined by carrying out an MTT assay in Vero cells. MBC: minimal bactericidal activity. $SI = IC_{50}/MIC_{100}$.

Scheme 1 Synthetic route to compounds 1a-e. Reagents and conditions: (i) MeOH, H₂SO₄, reflux; (ii) NH₂NH₂, MeOH; (iii) CNBr, MeOH; (iv) NaH, anhydrous THF, reflux.

anti-TB hits and leads, the MIC₁₀₀ value of 1a is not competent under replicating growth conditions (< 10 µM) but compounds 1b-e demonstrate a greater SI and, therefore, can be considered as anti-TB leads. Minimal microbicidal activity measurements showed 1a has the highest bactericidal activity, followed by 1d and 1b, and 1e, with 1c being the least active. The values of minimum bactericidal activity show that 1a also has the highest bactericidal activity followed by 1d and 1b, with compounds 1c and 1e being the least active [35].

Another criterion used to determine whether a compound can be considered as an early lead for an anti-TB drug is *in vitro* activity against *M. tuberculosis* strains that are resistant to a single TB drug, such as isoniazid or RIF, indicating a new mechanism of action. We also evaluated the compounds with non-virulent bacteria M. tuberculosis H37Ra. Non-virulent bacteria are susceptible to lower concentrations of drugs than virulent strains. Specifically, the activities of the synthesized compounds 1a-1e as inhibitors of the growth of bacteria of the M. tuberculosis strains H37Ra and 209 (RIF resistant-strain) using 250 μg/mL- 0.98 μg/mL serial two-fold dilutions were evaluated (Table 2). Also listed in Table 2 are the MIC_{100} values of RIF. All compounds are active against M. tuberculosis H37Ra (non-pathogenic) and 209 (resistant strain) with compound 1a being the most active. This result shows that compound 1a is an anti-TB hit with a new mechanism of action that should be explored.

It can be seen that bioisosteric modifications of the 5-nitrofuran moiety of compound 1a to give compounds 1b-e preserves anti-TB activity, albeit with inhibitory activities less than that of the lead compound 1a. The data for compounds 1b-e suggest that the anti-TB effect depends

Table 2 REMA-determined MIC₁₀₀ values of 1a-e against virulent, nonvirulent and RIF-resistant M. tuberculosis bacteria.

Compound	R	MIC ₁₀₀ (µg/mL) in H37Rv ATCC 27294	MIC ₁₀₀ (µg/mL) in H37Ra	MIC ₁₀₀ (µg/mL) in Mtb-209 (resistant)
1a	5-NO2C4H2O	7.80	1-2.00	7.8
1b	5-NO2C4H2S	15.60	15.60	15.60
1c	5-NO2C4H3N	31.25	7.8	7.8
1d	3-NO2C6H4	15.60	31.30	15.60
1e	5-C6H5	15.60	62.50	31.25
Rifampin	-	0.06	0.008	>64

M. tuberculosis H37Rv ATCC 27294 reference strain: Mtb. M. tuberculosis H37Ra: non-virulent strain. Mtb-209: RIF-resistant clinical isolate of M. tuberculosis.

on the aromaticity of the ring attached to the amide group, particularly if this ring has low aromaticity (phenyl > thiophene > pyrrole > furan) [36]. The relatively low anti-TB activity of the 5-nitropyrrole compound 1c, relative to the activities of 1b, 1d and 1e, may be due to the pyrrole-NH group of 1c forming a hydrogen bonding interaction with the active site.

Conclusions

Compounds 1a-e were synthesized using two convergent pathways with the hydrazide 2 as a common intermediate. The anti-TB activities of compounds 1a-e were evaluated using three Mycobacterium tuberculosis cell lines. The results suggest that the anti-TB activity of compounds 1a-e can be explained in terms of the distribution of electronic density across the ring attached to the amide group. Our ongoing studies are focused on exploring the mechanism by which these new compounds inhibit M. tuberculosis cell growth.

Experimental

Melting points were measured in open capillaries using a Mel-Temp apparatus. 1H-NMR spectra were recorded in DMSO- d_{ζ} on a Jeol Eclipse 300 spectrometer (300 MHz). ¹³C-NMR spectra were recorded at 75 MHz under otherwise similar conditions. IR spectra were obtained in KBr pellets using a Magna-IR spectrometer. Mass spectra were recorded on a Jeol JEM-AX505HA spectrometer with electronic impact (EI) ionization at 70 eV for low-resolution measurements and a Jeol AccuTOF DART and Jeol JMS700 (FAB+) instruments, for high-resolution measurements. Flash column chromatography was carried out on silica gel 60 (230-400 mesh ASTM) from Macherey-Nagel GmbH. Progress of the reactions was monitored by using TLC. The compounds were visualized using a dual short-wavelength/longwavelength UV lamp or staining with an ethanol solution of potassium permanganate, vanillin or p-anisylaldehyde. All reagents were used as purchased from Aldrich.

Synthesis of methyl 4-chlorobenzoate (3)

A solution of 4-chlorobenzoic acid (2, 33.07 mmol) and H₂SO₄ (1 mL) in anhydrous methanol (20 mL) was heated under reflux for 20 h. After consumption of starting material (monitored using TLC), the mixture was neutralized with a saturated NaHCO₃ solution (2 x 25 mL) and extracted with AcOEt (3 x 100 mL). The organic extracts were combined, dried over anhydrous Na₂SO₄, filtered and concentrated under reduced pressure to give 3 as a yellow oil (yield 80 %). The physical and spectral data of 3 were in accordance with previously reported data [37].

Synthesis of 4-chlorobenzohydrazide (4)

A solution of ester 3 (26.36 mmol) and hydrazine (61.85 mmol) in anhydrous methanol (20 mL) was heated under reflux for 12 h. After consumption of the starting material (as monitored using TLC), the reaction mixture was cooled, and the resulting solid was filtered using suction. The crude product was crystallized from hexane to give 4 as a white crystalline solid; yield 82%; mp 162-164°C. The physical and spectral data of 4 were in accordance with previously reported data [37].

Synthesis of 5-(4-chlorophenyl)-1,3,4-oxadiazol-2-amine (5)

A solution of 4 (5.86 mmol) and CNBr (10.65 mmol) in anhydrous methanol was heated under reflux for 5 h. After consumption of the starting material (as monitored using TLC), the mixture was neutralized with a saturated NaHCO₂ solution (2 x 25 mL) and the resulting solid was filtered using suction. The crude product was crystallized from methanol to give 5 as a white solid; yield 62%; mp 242-244°C. The physical and spectral data of 5 were in accordance with previously reported data [37].

General procedure for synthesis of compounds 1a-e

A solution of 5-(4-chlorophenyl)-1,3,4-oxadiazol-2-amine (5, 0.51 mmol) in anhydrous THF (15 mL) was treated with NaH (1.53 mmol), and the resulting mixture was cooled to 0°C under a nitrogen atmosphere before addition of acyl chloride **6a-e** (0.51 mmol). The mixture was stirred at room temperature for 15 h, quenched with a saturated NaHCO₃ solution (30 mL) and extracted with AcOEt (3 x 50 mL). The organic extracts were combined, dried over anhydrous Na,SO,, filtered, and concentrated under reduced pressure. The residue was purified using flash column chromatography on silica gel eluting with hexanes/EtOAc (6:4) to furnish the 1,3,4-oxadiazole 1a-e.

N-(5-(4-Chlorophenyl)-1,3,4-oxadiazol-2-yl)-5-nitrofuran-2-carboxamide (1a)

Yield 30% of a yellow solid; mp 250-252°C; IR: 3139, 1607 cm⁻¹; ¹H NMR (DMF- d_2): δ 7.40 (1H, d, J = 4 Hz), 7.64 (2H, d, J = 8 Hz), 7.74 (1H, d, J = 4 Hz), 7.94 (2H, d, J = 8 Hz); ¹³C NMR $(DMF-d_7)$: δ 113.8, 116.5, 123.6, 127.1, 127.8, 129.8, 136.5, 152.2, 157.8, 162.6, 168.3. HRMS (DART). Calcd for C₁H₀ClN₄O₄ $[M+1]^+$: m/z 335.0183. Found: m/z 335.0186.

N-(5-(4-Chlorophenyl)-1,3,4-oxadiazol-2-yl)-5-nitrothiophene-2-carboxamide (1b)

Yield 38% of a yellow solid; mp 260-262°C; IR: 3103, 1714 cm⁻¹; ¹H NMR (DMSO- d_6) δ : 7.68 (2H, d, J = 8 Hz), 7.95 (3H, m), 8.15 (1H, d, J = 4 Hz). ¹³C NMR (DMSO- d_c): δ 124.5, 128.9, 129.4, 130.0, 130.8, 131.4, 137.6, 143.9, 154.2, 159.8, 165.4. HRMS (DART). Calcd for $C_{13}H_7ClN_4O_4S$, $[M]^+$: m/z 350.9954. Found: m/z 350.9957.

N-(5-(4-Chlorophenyl)-1,3,4-oxadiazol-2-yl)-5-nitro-1H-pyrrole-2-carboxamide (1c)

Yield 30% of a yellow solid; mp 278-280°C; IR: 3176, 1635 cm⁻¹; ¹H NMR (DMSO- d_c) δ : 6.98 (1H, d, J = 4 Hz), 7.07 (1H, d, J = 4 Hz), 7.67 (2H, d, J = 8.5 Hz), 7.95 (2H, d, J = 8.5 Hz); ¹³C NMR (DMSO-*d_c*): δ 111.5, 114.6, 122.9, 128.2, 130.1, 132.2, 136.7, 143.4, 158.8, 159.5, 160.0. HRMS (DART). Calcd for $C_{13}H_{0}ClN_{5}O_{4}$, [M+1]+: m/z 334.0343. Found: m/z 334.0347.

N-(5-(4-Chlorophenyl)-1,3,4-oxadiazol-2-yl)-3-nitrobenzamide (1d)

Yield 59% of a white solid; mp 264-266°C; IR: 3176, 1635 cm⁻¹; ¹H NMR (DMSO- d_c): δ 8.88 (1H, s), 8.49 (1H, d, J = 7.5Hz), 8.39 (1H, d, J = 7.5 Hz), 7.95 (2H, d, J = 8 Hz), 7.78 (1H, m), 7.65 (2H, d, J = 8 Hz); ¹³C NMR (DMSO- d_c): δ 122.1, 123.2, 123.7, 127.4, 128.0, 129.7, 130.5, 130.6, 134.8, 135.4, 136.7, 147.9, 165.6. HRMS (DART). Calcd for $C_{15}H_{10}ClN_{\mu}O_{\mu}$, $[M+1]^+$: m/z345.0390. Found: *m/z* 345.0394.

N-(5-(4-Chlorophenyl)-1,3,4-oxadiazol-2-yl)benzamide (1e)

Yield 78% of a white solid; mp 224-226°C; IR: 3050, 1712 cm⁻¹; ¹H NMR (DMSO- d_z) δ : 7.65 – 7.51 (3H, m), 7.69 (2H, d, J = 8.5 Hz), 7.98 (2H, d, J = 8.5 Hz), 8.04 (2H, d, J = 7 Hz), 12.2 (1H, br); 13 C NMR (DMSO- d_c): δ 122.2, 127.8, 128.3, 128.6, 129.6, 132.2, 133.0, 136.4, 158.0, 160.4, 164.9. HRMS (DART). Calcd for $C_{15}H_{11}ClN_3O_2$, $[M+1]^+$: m/z 300.0540. Found: m/z300.0540.

In vitro antimycobacterial assay

Stock solutions of all compounds were prepared in 100% dimethyl sulfoxide (DMSO) at 10 µg/mL. For the REMA assay, compounds were diluted in 7H9 medium without tyloxapol. For reference drugs, stocks of 64 µg/mL were prepared and filtered through a membrane with pore diameters of 0.22 mm (Millipore; Darmstadt, Germany). All working solutions were kept refrigerated at -20°C until they were evaluated.

The cytotoxicity assay

The assay was carried out using Vero cell lines (kidney of African green monkey) from ATCC. The cells were cultured in RPMI 1640 medium supplemented with 10% FBS and nonessential amino acids.

Cytotoxicity assay

Ten-thousand Vero cells were placed in a 96 well-plate and incubated for 24 h in 100 µL of RPMI medium. After incubation, the plate was washed and new fresh medium with the compound at a various concentration was added. Each tested compound was incubated for 48h at 37°C in a 5% CO_3 atmosphere. Then, a volume of 10 μL of MTT (5 µg/mL in sterile PBS) was added to each well and incubation was continued for another 4 h. The medium was removed and a volume of 100 µL of DMSO was added to solubilize the formazan. Absorbance was determined at a wavelength of 570 nm and cytotoxicity was calculated as % toxicity = (1-(ABS problem/ABS control))*100. Controls were cells without treatment [38].

M. tuberculosis culture conditions

M. tuberculosis, H37Rv, H37Ra and 209 strains were cultivated in a 7H9-glycerol-10% ADC-0.01% tyloxapol medium at 37°C until an O.D. of 0.4 at a wavelength of 600 nm was reached. Working bacteria-solutions were obtained by dilution 1:25 in 7H9-ADC 10%.

Antimicrobial susceptibility test using the resazurin microtiter assay (REMA)

The assay used here was previously described by Collins and Franzablau [39]. Briefly, the outer wells of a 96-well plate were each filled with 200 µL of sterile PBS to prevent dehydration from occurring during the long-duration (8-day) incubation. RIF was used as a reference drug $(16 - 0.001 \,\mu\text{g/mL} \text{ serial two-fold dilutions})$ in each plate, and controls of DMSO, DMSO+Mtb, medium, media+Mtb, and compound only to validate the plate were also included. Compounds were evaluated at various concentrations from 0.98 μ g/mL to 250 μ g/mL, and in triplicate in independent assay experiments. Plates were incubated for six days; then, 30 µL of 0.01% resazurin (weight/

volume) (Sigma Aldrich) were added to each well and the plates were incubated for two more days. Visual inspection was used to determine the colors of the contents of each well, with blue interpreted as no growth, pink as growth, and MIC as the last concentration in which blue color were observed. The minimum inhibitory concentration (MIC) is defined as the minimum concentration of tested compound to which there was not higher shift from resazurin (blue) to resorufin (pink) than that generated by control of a 1:100 dilution of the bacterial inoculum [36].

Minimal bactericidal concentration

The MBC values were determined for the well (s)+compound that did produce a shift in color and were then re-incubated in fresh medium. A volume of 5 µL of this well (s) was added to 195 µL of fresh medium and incubated for carrying out the REMA assay as described above. MBC corresponds to the minimum concentration of tested compound that does not produce a shift in cultures reincubated in fresh medium [36].

Selectivity indexes

The **SI** values were obtained using the formula $SI = IC_{50}$ in Vero cells and the MIC₁₀₀ values were determined using REMA.

Acknowledgements: Financial support from the DGAPA and UNAM (projects PAPIIT IN208015 and NUATEI-IIB-UNAM) is gratefully acknowledged. We also thank R. Patiño, A. Peña, E. Huerta, B. Quiroz, L. Velasco, J. Pérez, and E. Segura Salinas for technical support. Gladys J. Nieves Zamudio (273584) and Gustavo Pretelín Castillo (308250) are recipients of CONACYT Graduate Scholarships.

References

- [1] http://www.who.int/en/news-room/fact-sheets/detail/ tuberculosis> cited 16 February, 2018
- [2] Wong, E.; Cohen, K.; Bishai, W. Rising to the challenge: new therapies for tuberculosis. Trends Microbiol. 2013, 21, 493-501.
- [3] Nayyar, A.; Jain, R. Recent advances in new structural classes of anti-tuberculosis agents. Curr. Med. Chem. 2005, 12, 1873-1886.
- [4] Rawat, B.; Rawat, D.S. Antituberculosis drug research: a critical overview. Med. Res. Rev. 2013, 33, 693-764.
- [5] Negi, A. S.; Kumar, J. K.; Luqman, S.; Saikia, D.; Khanuja, S. P. S. Antitubercular potential of plants: A brief account of some important molecules. Med. Res. Rev. 2010, 30, 603-645.

- [6] Kumar, V.; Patel, S.; Jain, R. New structural classes of antituberculosis agents. Med. Res. Rev. 2018, 38, 684-740.
- [7] Akhter, M.; Husain, A.; Azad, B.; Ajmal, M. Aroylpropionic acid based 2,5-disubstituted-1,3,4-oxadiazoles: synthesis and their anti-inflammatory and analgesic activities. Eur. J. Med. Chem. 2009, 44, 2372-2378.
- [8] Calcagno, A.; D'Avolio, A.; Bonora, S. Pharmacokinetic and pharmacodynamic evaluation of Raltegravir and experience from clinical trials in HIV-positive patients. Expert Opin. Drug Metab. Toxicol. 2015, 11, 1167-1176.
- [9] Nieddu, V.; Pinna, G.; Marchesi, I.; Sanna, L.; Asproni, B.; Pinna, G. A.; Bagella, L.; Murineddu, G. Synthesis and antineoplastic evaluation of novel unsymmetrical 1,3,4-oxadiazoles. J. Med. Chem. 2016, 59, 10451-10469.
- [10] Sun, J.; Ren, S. Z.; Lu, X. Y.; Li, J. J.; Shen, F. Q.; Xu, C.; Zhu, H. L. Discovery of a series of 1,3,4-oxadiazole-2(3H)-thione derivatives containing piperazine skeleton as potential FAK inhibitors. Bioorg. Med. Chem. 2017, 25, 2593-2600.
- [11] Tok, F.; Kocyigit-Kaymakcioglu, B.; Tabanca, N.; Estep, A. S.; Gross, A. D.; Geldenhuys, W. J.; Becnel, J. J.; Bloomquist, J. R. Synthesis and structure-activity relationships of carbohydrazides and 1,3,4-oxadiazole derivatives bearing an imidazolidine moiety against the yellow fever and dengue vector. Aedes aegypti. Pest. Manag. Sci. 2017, 74, 413-421.
- [12] Mei, W.; Ji, S.; Xiao, W.; Wang, X.; Jiang, C.; Ma, W.; Zhang, H.; Gong, J.; Guo, Y. Synthesis and biological evaluation of benzothiazol-based 1,3,4-oxadiazole derivatives as amyloid β -targeted compounds against Alzheimer's disease. *Monatsh*. Chem. 2017, 148, 1807-1815.
- [13] Search in the SciFinder database with the entry "biological activity of 1,3,4-oxadiazoles", October 10, 2017.
- [14] Kipandula, W.; Young, S.A.; MacNeill, S. A.; Smith, T. K. Screening of the MMV and GSK open access chemical boxes using a viability assay developed against the kinetoplastid Crithidia fasciculata. Mol. Biochem. Parasitol. 2018, 222, 61-69
- [15] Kumar Reddy, A. L.V.; Kathale, N. E. Synthesis, Characterization and antiinflammatory activity of chalcone derivatives linked with Apocynin and 5-nitrofuran moiety. Asian J. Chem. 2018, 30, 312-316.
- [16] Xu, B., Ding, X., Wu, Y.; Cui, L.; Qian, P.; Wang, D.; Zhao, Y. Synthesis and antibacterial activity of oxazolidinone derivatives containing nitro heteroaromatic moiety. Chem. Res. Chin. Univ. 2018 34, 51-56.
- [17] Romero, A. H.; Medina, R.; Alcala, A.; Garcia-Marchan, Y.; Nunez-Duran, J.; Leanez, Jacques; Mijoba, Ali; Ciangherotti, Carlos; Serrano-Martin, Xenon; Lopez, Simon E. Design, synthesis, structure-activity relationship and mechanism of action studies of a series of 4-chloro-1-phthalazinyl hydrazones as a potent agent against Leishmania braziliensis. Eur. J. Med. Chem. 2017, 127, 606-620.
- [18] Asadipour, A.; Edraki, N.; Nakhjiri, M.; Yahya-Meymandi, A.; Alipour, E.; Saniee, P.; Siavoshi, F.; Shafiee, A.; Foroumadi, A. Anti-Helicobacter pylori activity and structure-activity relationship study of 2-alkylthio-5-(nitroaryl)-1,3,4-thiadiazole derivatives. IJPR. 2013, 12, 281-287.
- [19] Krasavin, M.; Lukin, A.; Vedekhina, T.; Manicheva, O.; Dogonadze, M.; Vinogradova, T.; Zabolotnykh, N.; Rogacheva, E.; Kraeva, L.; Yablonsky, P. Conjugation of a 5-nitrofuran-2-oyl moiety to aminoalkylimidazoles produces non-toxic nitrofurans that are efficacious in vitro and in vivo against multidrug-

- resistant Mycobacterium tuberculosis. Eur. J. Med. Chem. 2018, 157, 1115-1126.
- [20] Chiarelli, I. R.; Mori, M.; Barlocco, D.; Beretta, G.; Gelain, A.; Pini, E.; Porcino, M.; Mori, G.; Stelitano, G.; Costantino, L.; Lapillo, M.; Bonanni, D.; Poli, G.; Tuccinardi, T.; Villa, S.; Meneghetti, F. Discovery and development of novel salicylate synthase (Mbtl) furanic inhibitors as antitubercular agents. Eur. J. Med. Chem. 2018, 155, 754-763,
- [21] Yempalla, K. R.; Munagala, Y. G.; Singh, S.; Magotra, A.; Kumar, S.; Rajput, V. S.; Bharate, S. S.; Tikoo, M.; Singh, G. D.; Khan, I. A.; Vishwakarma, R. A.; Singh, P.P. Nitrofuranyl methyl piperazines as new anti-TB agents: identification, validation, medicinal chemistry, and pk studies. ACS Med. Chem. Lett. **2015**, 6, 1041-1046.
- [22] Szatylowicz H.; Stasyuk O. A.; Fonseca Guerra C.; Krygowski, T. M. Effect of intra- and intermolecular interactions on the properties of para-substituted nitrobenzene derivatives. Crystals 2016, 6, 29.
- [23] Canché-Chay, C.; Gómez-Cansino, R.; Espitia-Pinzón, C.; Torres-Ochoa, R. O.; Martínez, R. Synthesis and anti-tuberculosis activity of the marine natural product caulerpin and its analogues. Mar. Drugs 2014, 12, 1757-1772.
- [24] Gasteiger, J. Chemoinformatics: achievements and challenges, a personal view. Molecules 2016, 21, 151-165.
- [25] Katsuno, K.; Burrows, J. N.; Duncan, K.; van Huijsduijnen, R. H.; Kaneko, T.; Kita, K.; Mowbray, C.E.; Schmatz, D.; Warner, P.; Slingsby, B.T. Hit and lead criteria in drug discovery for infectious diseases of the developing world. Nat. Rev. Drug Discov. 2015, 14, 751-758.
- [26] Moreira Lima, L.; Barreiro, E. Bioisosterism: a useful strategy for molecular modification and drug design. Curr. Med. Chem.
- [27] Kim S.; Thiessen, P. A.; Bolton, E. E.; Chen, J.; Fu, G.; Gindulyte, A.; Han, L.; He, J.; He, S.; Shoemaker, B. A.; Wang, J.; Yu, B.; Zhang, J.; Bryant, S. H. PubChem substance and compound databases. Nucleic Acids Res. 2016, 44, D1202-13.
- [28] Xu Y. J.; Johnson M. Using molecular equivalence numbers to visually explore structural features that distinguish chemical libraries. J. Chem. Inf. Comput. Sci. 2002, 42, 912-926.
- [29] Xu Y. J.; Johnson M.A. Algorithm for naming molecular equivalence classes represented by labeled pseudographs. J. Chem. Inf. Comput. Sci. 2001, 41,181-185.
- [30] Medina-Franco, J. L.; Petit, J.; Maggiora, G. M. Hierarchical strategy for identifying active chemotype classes in compound databases. Chem. Biol. Drug Des. 2006, 67, 395-408.
- [31] Medina-Franco, J. L. Activity cliffs: facts or artifacts? Chem. Biol. Drug Des. 2013, 81, 553-556.
- [32] Jorge, S.D.; Palace-Berl, F.; Masunari, A.; Cechinel, C. A.; Ishii, M.; Pasqualoto, K. F. M; Tavares, L.C. Novel benzofuroxan derivatives against multidrug-resistant Staphylococcus aureus strains: design using Topliss' decision tree, synthesis and biological assay. *Bioorg. Med. Chem.* **2011,** *19,* 5031-5038.
- [33] Patel, N. B.; Patel, J. C. Synthesis and antimicrobial activity of 3-(1,3,4-oxadiazol-2-yl) quinazolin-4(3H)-ones. Sci. Pharm. 2010. 78. 171-193.
- [34] Hou, Z.; Nakanishi, I.; Kinoshita, T.; Takei, Y.; Yasue, M.; Misu, R.; Suzuki, Y.; Nakamura, S.; Kure, T.; Ohno, H.; Murata, K.; Kitaura, K.; Hirasawa, A.; Tsujimoto, G.; Oishi, S.; Fujii, N.

- Structure-based design of novel potent protein kinase CK2 (CK2) inhibitors with phenyl-azole scaffolds. J. Med. Chem. **2012,** 55, 2899-2903.
- [35] Coronado-Aceves, E. W.; Sánchez-Escalante, J. J.; López-Cervantes, J.; Robles-Zepeda, R. E.; Velázquez, C.; Sánchez-Machado, D. I.; Garibay-Escobar, A. Antimycobacterial activity of medicinal plants used by the Mayo people of Sonora, Mexico. J. Ethnopharmacol. 2016, 190, 106-115.
- [36] Balaban, A. T.; Oniciu, D. C.; Katritzky, A. R. Aromaticity as a cornerstone of heterocyclic chemistry. Chem. Rev. 2004, 104, 2777-2812.
- [37] Khan, S. A.; Ahuja, P.; Husain, A. Oxidative cyclization of isoniazid with fluoroquinolones: synthesis, antibacterial and antitubercular activity of new 2,5-disubstituted 1,3,4oxadiazoles. J. Chin. Chem. Soc. 2017, 64, 918-924.].
- [38] Mossman, T. Rapid colorimetric assay for cellular growth and survival: application to proliferation and cytotoxicity assays. J. Immunol. Methods 1983, 65, 55-63.
- [39] Collins, L. A.; Franzblau, S. G. Microplate Alamar Blue assay versus BACTEC 460 system for high-throughput screening of compounds against Mycobacterium tuberculosis and Mycobacterium avium. Antimicrob. Agents Chemother. 1997, 41,1004-1009.