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Review Article

Asmaa M. El-Shafey*

Carbon dots: Discovery, structure, fluorescent properties, and applications

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Abstract: Nanotechnology has become one of the most important topics since the beginning of the twenty-first century in numerous fields including drug synthesis and delivery, environmental protection, electronics manufacture, and astronomy due to their nanoscale particles and their properties. The traditional semi-quantum dots are replaced by a new category of fluorescent carbon nanomaterials. Carbon dots (CDs) have been explored in the last few years for their simple synthetic accession, good bio-consonance, and several revelation applications. This review explains the fluorescent properties of CDs in brief, giving also a background on CDs discovery, structure, and composition, as well as on nanocomposites, green synthesis, and their applications. Resources conservation can be achieved by using recycled substances for sustainable development which lead to a new technology. Fluorescent CDs synthesized from food wastes like bananas, orange peel waste, sugarcane bagasse, *Trapa bispinosa* peels, bread, and jaggery have several applications such as sensing, drug delivery, gene transfer, biological imaging, and food safety. In this study, we concentrate on CDs greener methods to prepare effective and biocompatible CDs.

Keywords: C-dots, fluorescence, nanocomposites, green synthesis, bioimaging

1 Introduction

The important environmental matter that occupied our minds is that the world consists of 7.2 billion people and tolerating the waste resulted from food production as well as consumption of this population is considered a big disaster in our universe, a fundamental environmental issue

and huge challenge [1-3]. Agriculture and industry outputs in food production increased by science and technology support, but the statistical studies proved a marked increase in food waste by both the consumer and production sides [4]. It was essential to think of a solution to convert wastes into useful materials to achieve the sustainability principle [5], so scientists worked on nanomaterials synthesis from natural sources and products for biological and environmental sensing applications [6-8]. Xu and colleagues were the first team to discover carbon nanodots (CDs) which are a category of photo luminescent nanoparticles (NPs) [9]. CDs and its derivatives have been used for numerous applications which are considered bioconvenient and eco-friendly as CDs can be prepared from any starting material including carbon; therefore, food wastes were reused because of the majority of by-products of the food; food wastes from vegetables, livestock, fruits, and food industry enhance the progress and sustainability of nanotechnology [10-14]. Figure 1 explains sustainable development of CDs technology [15].

New category of fluorescent carbon nanomaterials has size below 10 nm, which means it possesses ultrafine size which is referred to as carbon quantum dots (CDs). The separation and purification of single-walled carbon nanotubes were the main reason for its first discovery in 2004 [9,16]. CDs are perfect for bioimaging [17–19] by shifting the emission wavelength to the red region as CDs emit blue fluorescence under UV excitation and they also have different properties such as excitation-/pH-dependent property, chemical stability, and resistance to photobleaching as well as up-conversion fluorescence so that the CD synthesis and its numerous applications have been studied (Figure 2) [20].

Renewable resources and small molecules are used as precursors with the aid of heating, microwave irradiation, and ultrasound. Depending on the fluorescence changes, different sensing protocols will be achieved especially plenty of sensing strategies have been developed by variations in the CD synthesis methods [21–23].

CDs have several applications [24–26], such as CD probes in food safety [27]. Various types of contaminants, involving pesticides [28], veterinary drugs [29],

^{*} Corresponding author: Asmaa M. El-Shafey, Chemistry Department, Faculty of Science and Arts, King Khalid University, Sarat Ebida, Saudi Arabia, e-mail: sommy_28@yahoo.com

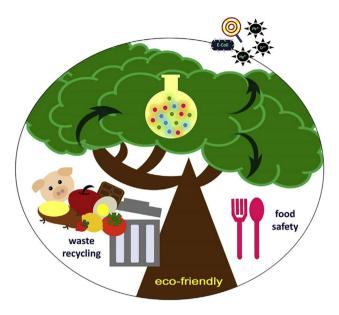


Figure 1: Sustainable development of carbon nanodot technology [15].

bacteria [30], and banned additives [31] can be determined by using CD probes beside food analysis application. Therefore, it is fundamental to gather an inclusive literature review to offer great knowledge about the CD

discovery, structure, and composition as well as about the green synthesis of CDs, its fluorescent properties, and its several applications. We wish that the review presents worth information and easy induction of CD-based sensing.

2 CDs discovery

Surface defects in single- and multiwalled nanotubes were the provenance of fluorescent carbon observation. Luminescence is promoted by the carbon nanotube surface passivation [32,33], which led to the CD discovery as we mentioned before; the first time of discovery in 2004 was fortuitously through electrophoretic single-walled nanotube purification. Under UV light, different colors have been exhibited by the fluorescent material of the fast moving band. The result of characterization indicated that the material was completely metal free and consisted of carboxyl groups, and the composition determined was C 53.93%, H 2.56%, O 40.33%, and N 1.20% as indicated in Figure 3 [9]. Since then, numerous methods of CDs synthesis have been reported by many groups of researchers.

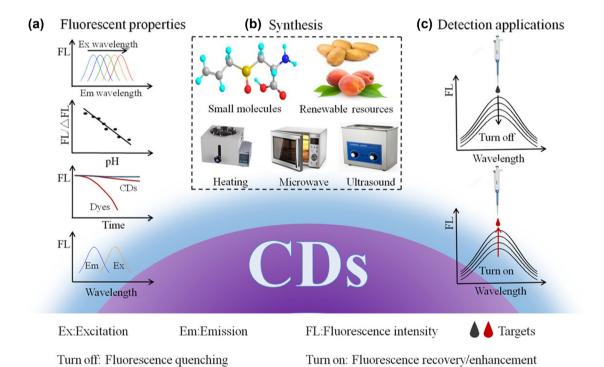


Figure 2: The importance of CDs in fluorescence properties, synthesis, and detection implementations in foods. (a) Fluorescent properties involve excitation-dependent, pH-dependent, antiphotobleaching, and up-conversion fluorescence. (b) Synthetic methods include hydrothermal/solvothermal, microwave-assisted, and ultrasonic-assisted methods. (c) The principles of CD-based biosensor can be divided into two types: fluorescence quenching and fluorescence recovery [20].

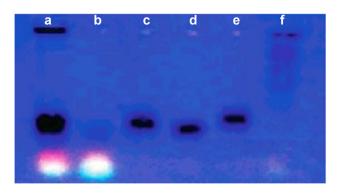


Figure 3: Electrophoretic profile in 1% agarose gel under 365 nm UV light: (a) crude SWNT suspension, (b) fluorescent carbon, (c) short tubular carbon, (d and e) further separation of (c), and (f) cut SWNTs [9].

2.1 Capillary zone electrophoresis (CZE)

CZE with featured separation qualification possesses an important speed separation approach for NPs due to their different electrophoretic mobilities (on the basis of charge/size ratio) through an electrolyte solution contained in a fused silica capillary under the impact of an electric field. The CZE utilization in conjunction with a diode array detector (230 nm) for CD separation attained from the flame of an oil lamp. Buffer composition effect tested on the negatively charged CD electropheric pattern by using various pattern types, pH, and concentrations as capillary electrophoresis (CE) is a perfect method convenient to implement favorable separation qualification, the advantages provided by the theoretical plates as high number obtained with CE for CD separation are overshadowed by its UV detection systems, and low sensitivity due to the sample volumes of small injection. To overcome the low-sensitivity limitation of the CDs' analysis accuracy, CDs can be separated by the utilization of CZE apparatus coupled with a diode array detector and a laser-induced fluorescence (LIF) detector. The CD absorption electrograms were obtained at 250 nm and the fluorescence was observed at an excitation wavelength $(\lambda_{\rm ex}/\lambda_{\rm em})$ of 488/550 nm. The separation was achieved using 40.0 cm capillary (50 μm i.d. and 325 μm o.d.) with 30 mM sodium acetate-acetic acid (NaAc-HAc) at pH 3.6 as run buffer and applied voltage of 15 kV. Figure 4 described that the UV (curve (a) and LIF (curve (b)) detection combination detects boosting as well as complex CD mixture supplemental information. For example, positively charged, neutral, and negatively charged CDs gained with two detection processes are the same. While peaks (1) and (2) show intense UV and fluorescence signals, peak (3) exhibits strong absorption but weak emission signals. To explore

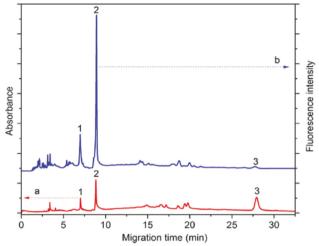


Figure 4: Capillary electrophoretic CD separation monitored at (a) absorption wavelength of 250 nm and (b) laser-induced fluorescence at $\lambda_{\rm ex}/\lambda_{\rm em}$ of 488/550 nm. Peaks (1), (2), and (3) are positively charged, neutral, and negatively charged CDs, respectively [34].

the reaction time-associated kinetics of CD consistence and identify the functional groups-associated charge states, the CE method was used. Applied CE coupled with UV detection (250 nm) and LIF detection ($\lambda_{ex}/\lambda_{em}$ 488/550 nm) for the separation of hollow CD separation synthesized from glacial acetic acid and diphosphorus pentoxide without external heating. The separation was acheived utilizing 40.0 cm capillary (50 µm i.d. and 325 µm o.d.) with 10 mM sodium dodecyl sulfate and 30 mM phosphate (pH 9.0) as run buffer at an applied voltage of 15 kV. By utilizing UV-Vis spectroscopy, photoluminescence (PL) spectroscopy, and transmission electron microscopy (TEM), the separated fractions were also gathered and analyzed. The CZE is confirmly utilized in identifying the different charge states of CDs species existed in a complex CDs mixture by using individual CE-fractionated. CDs fractions enough amounts collection owing to the low sample injection volume of CZE is timeconsumption. The neutral CD species eluted out as one single strong peak, but the charged CDs can only be separated by CE. Isoelectric focusing (IEF) is used for isoelectric point detection of macromolecules and bioparticles. The different sizes of colloidal NPs can selectively be separated by size from their mixture in a homemade miniscale IEF unit as it is a fast, sensitive, and low-cost-effective method [34].

3 CD structure and composition

CDs obtained from different structures may be either graphitic or amorphous. The size of CDs can be tuned by

different nanocomposites. CDs have been studied as a material with a multitude of applications. In terms of analytical attributes (Figure 5), transmission electron microscopy (TEM) and X-ray diffraction (XRD) are commonly utilized to characterize CDs as new material. IR spectroscopy and elementary analysis help in recognizing the surface functional groups which mark CDs' affinity properties and signify chemical mechanisms to be utilized in further expansion. Another attribute of CDs is absorption in UV-Vis spectrum to assess the excitation band for fluorescence spectrum and observe the change in surface properties of CDs because of interaction with another material or derivatization by subordinating fluctuations in the fluorescence intensity of C-dots. CDs' fluorescence emission is used for targets observed under microscope as it makes NPs as labelfree probes [11,12].

The reported average size of CDs of less than 10 nm facilitated its use in biological applications. Fluorescence emission can be enhanced by surface passivation of CDs. When CDs were coated by the hydroxyl groups, we noticed the following observations:

- 1. The reported CD diameter was 3.1 \pm 0.5 nm.
- 2. TEM showed that the particles were uniformly dispersed and spherical in shape as shown in Figure 6.
- 3. The quantum yield (QY) increased and was about 5.5%.
- 4. Better fluorescence emission and the particles were photostable [35,36].

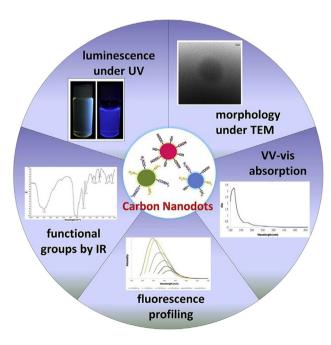
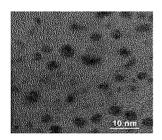


Figure 5: Characterization of C-dots: spectra and images [15].



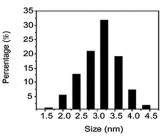


Figure 6: TEM image (left) and size distribution (right) of C-dots [36].

That means if the size of the particle remains under 10 nm even after surface passivation to promote optical properties, CDs would have the best performance in cell imaging.

Another group of CDs prepared from bananas as a starting material had reported a mean size of 3 nm; is spherical in shape; and includes carbon, oxygen, and potassium, though the percentage of potassium is very less and the majority are carbon and oxygen. The interlayer spacing of CDs that involved oxygen-containing groups with sp³ carbon and hydroxyl connected carbon groups was established to be 0.42 nm which is higher than the graphitic interlayer spacing of 0.33 nm [37], which denotes that CDs are crystalline in nature in comparison to graphite as indicated in Figure 7. The third group of CDs synthesized from food caramels and orange peel waste is amorphous in nature and has no crystalline nature as the XRD displayed a broad amorphous peak [38]. The fourth group of CDs prepared from graphite had a diamond-like structure. The percentage of squares of the ring radius was reported to be 3:8:11:16:19, where the rings correspond to the planes {111}, {220}, {311}, {400}, and {331} that belong to diamond [39]. The fifth group of CDs derived from multiwalled carbon nanotubes by electrochemical oxidation also showed graphitic nature. The high-resolution TEM displayed the lattice spacing to be 3.3°A, which is close to the (002) facet of graphite. The sixth group of CDs attained from candle soot is high in oxygen content and different functional groups such as hydroxyl, carbonyl, carboxyl, and epoxy groups were attached to CDs [40]. The reported structure of soot was C 91.69%, H 1.75%, N 0.12%, and O (calculated) 44.66% [41]. The size distribution of CDs was narrow, i.e., between 2 and 7 nm. CDs have a larger area due to their smaller size, which led to greater absorption to boost CDs' applicability to optoabsorption. CDs also have the susceptibility to allow drug connection to the

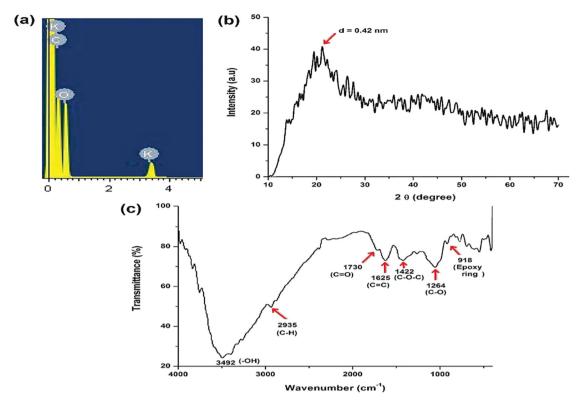


Figure 7: (a) EDX spectrum, (b) XRD pattern, and (c) FTIR spectrum of the carbon dots [37].

surface for drug delivery systems. Finally, the seventh group of CDs, CD/monoclinic bismuth vanadate ($BiVO_4$) nanocomposites, formed two various structures by attaching to nanospherical and nanoplatelet-shaped $BiVO_4$. The complex size increased to 350–400 nm for nanospheres and 500 nm for nanoplatelets [42].

4 Fluorescent properties

CDs' essential optical property is fluorescence emission. However, the mechanism of fluorescence emergence still remains detectable. There are several reported mechanism, including quantum size effect [16,43], surface defect states [44,54], molecular, molecule like states [44–46]. The surface defect states allude to the formed surface defects by oxidation whereas the quantum size effect points to the size effect on radiative recombination of electron–hole pair [47,48]. The CD fluorescence radix in some cases is due to the molecular and molecular-like states, especially for the citric acid-derived CDs [46]. These mechanisms could give explanation for the different fluorescent properties involving excitation- and pH-dependent properties.

4.1 Excitation-/pH-dependent property

By adjusting the excitation wavelength, different fluorescence emissions can be achieved especially when the excitation wavelength changed from 340 to 480 nm, Liu and coworkers observed that the maximum emission wavelength increased from 450 to 550 nm [49] due to the distribution of CDs and the surface states [50–53]. CD fluorescence is affected by pH [54] due to functional group protonation and deprotonation on CDs' surface [55–57]. One of the fluorescence pH sensors has been manufactured by displayed good linearity between fluorescent intensity and pH in the range of 3.5–10.00 [56]. Multicolor imaging, pH sensing, and the fluorescence mechanism are controlled by excitation-/pH-dependent properties which play the main role [58–60].

4.2 Chemical stability, antibleaching, and photobleaching properties

Fluorescence sensing/imaging in detection applications needs stable fluorescence signal and long emission lifetimes. CDs emitted strong fluorescence in aqueous

solution for 6 h [61,62] or even 1 year [63] as CDs are stable when maintained in aqueous solution and high-salinity solutions for long period. By dissolving in 4 NaCl solution as well as in high concentrations (400 μ M) of H₂O₂ solution, Liu's group noticed a perfect signal of oxidation impedance when the prepared CDs were quite stable [64]. CDs usually display excellent impedance to photobleaching as CDs have the ability to resist changes in pH over the wide range of 3–12 [65] that is, photobleaching impedance is more important than chemical stability due to the long observation times reported in other essays. For instance, various CDs fluorescent intensity reduced slightly after UV irradiation for 12 h [66] and 24 h [67].

4.3 Up-conversion fluorescence

The emission wavelength is shorter than the excitation wavelength which is referred to as up-conversion fluorescence. Decrease in the background autofluorescence is due to the longer excitation wavelengths and various labeling with different emission wavelengths' sand photon tissue permeation improvement as a result of up-conversion fluorescence, which makes it an essential role in imaging [68]. CDs prepared by ultrasonic treatment has the property of up-conversion fluorescence [62,69,70]. CDs prepared from glucose and ammonium hydroxide displayed an up-conversion emission in the range of 300–600 nm with excitation at 650–1,000 nm [62]. Two photon absorption may generate up-conversion fluorescence [71].

4.4 Long wavelength attributes

When the CDs reported were illuminated with UV light, it emits blue fluorescence [21,72,73]. The biological analysis utilizes blue-emitting CDs which is not veiled since biotissues and cells also emit blue fluorescence, which will lead to spectral interference, so the perfect solution to this problem was to use long wavelength emission to conquer this matter. Numerous treatments such as surface modification [74], size control [75], and pH control [60] were used to prepare CDs with longer wavelength fluorescence. The introduction of heteroatoms such as nitrogen [76] and oxygen [71] led to the red shift in the emission wavelength in spite of the fluorescence from blue to green/yellow. By forming a large conjugated π

system, the CDs with red fluorescence have been easily prepared. Preparation of red-fluorescencing CDs with high QY can be executed by using p-phenylenediamine [77] and 1,3-di-hydroxynaphthalene [78] and using the threefold symmetric phloroglucinol as precursor, Yuan et al. [79] worked on expanding the conjugated π system under controlled conditions to prepare highly fluorescent CDs from blue to red to decrease the spectral interference fluorescent signals and protect tissues from potential damage in addition to CDs with longer emission wavelengths' preparation shares in the explanation of the fluorescence mechanism.

4.5 Toxicity

Material toxicity is a significant measure of the material's potential manipulate environment and its denizens in various applications. In the last few years, the toxicity of CDs was discovered. 3-(4,5-Dimethyl thiazol-2-yl)-2,5 diphenyltetrazoliumbromide (MTT) assay is a common in vitro toxicity analysis and is assessed by multiple groups. Zhang et al. explored whether the assigned CDs in HeLa cells would have frivolous toxicity in the concentration range of 50-400 µg/mL [80-82]. Sahu and coworkers found that CDs were acceptable at high dose (200 µg/mL) for prolonged incubation time (72 h) and nontoxicity to L929 cell lines when the CD concentration is elevated as 1 mg/mL [83]. Zhou et al. show that the viabilities in MCF-7 cells minimized by 20% [84]. CD toxicity is connected with the surface passivating agents [85] and modulated materials, such as polyethylene glycol (PEG) [80] and polyacrylic acid [86]. The attained results indicated that these CDs are still characterized by the low toxicity in vitro. Yang and coworkers estimated the toxicity of CDs in male CD-1 mice even at elevated dosage of 40 mg/kg (CD/body weight) according to the serum biochemistry assays and histopathological analyses [87]. Also, in vitro toxicity of CDs and by taking their surface charge into consideration, mouse fibroblasts NH/3T3 cells [88] showed that CDs with negative charge due to the carboxylic groups arrested the G2/M phase of the cell cycle. It is clear that CDs' fabricated reproduction led to higher active oxygen species index without emanating the cell nucleus. Polyethylene glycol (PEG)-modified CDs having neutral charge did not stimulate any abnormalities in cell morphology, neither in intracellular trafficking nor in cell up to a dose of 300 mg/mL. If CDs were coated by polyethylenimine (PEI) with positive charge, they will be the most toxic, being capable of

getting in the nucleus and boosting changes in the GO/G1 phase of the cell cycle at concentration as low as 100 mg/mL. Cytotoxicity profiling CDs from fresh tender ginger juice quelled the growth of human hepatocellularcarcinoma cells with an IC₅₀ of 0.35 mg/mL and accompanying 18-fold rise in cellular ROS production [89]. Furthermore, CDs arrested the growth of liver tumors in a BALB/c mouse paradigm, and these CDs were intoxic to MCF-10A cells (noncancerous mammary epithelial cells), FL83B cells (normal liver cells), a 549-cell (human lung cancer), MDA-MB-231 cells (human breast cancer), and HeLa cells (human cervical cancer) [90]. CDs synthesized from spent green tea were water soluble and arrested MCF-7 (IC₅₀ = $0.155 \,\text{mg/mL}$) and MDA-MB-231 ($IC_{50} = 0.072 \text{ mg/mL}$) cell growth with lower toxicity to HeLa, LLC-PKI (renal proximal tubular), and MCF-10A cells. ROS generation occurred in the inhibitory activity. CDs had a little impact on the thrombin activity by fibrinogen conversion to fibrin, thus posing no hazard on blood coagulation. These results showed that CDs with low toxicity can be considered the gold standard in the context of sustainability. According to the CDs' impact in air, rats, and fish [91], many scientists evaluated the low [92] environmental CD toxicity when the CD concentration is not so high.

5 CD nanocomposites

CDs offer an appropriate photo charge carrier flow as it both accepts electron and transports electron, thus boosting the photocatalytic characterization. Semiconductor carbon nanomaterials' preparation led to enhanced catalytic qualification. A lot of reports discussed CDs photocatalytic nature without complex formation as CDs were prepared by using alkali-assisted electrochemical method. C-dot irradiation by near-infrared (NIR) light was used to test the photocatalytic activity led to benzyl alcohol oxidation to form benzaldehyde, which means the photocatalytic activity of CDs has the ability to transform alcohols to their corresponding aldehyde [93]. When ZnO is excited by UV light, electron-hole pair results plus initiation of hydroxyl formation in water [94] making it a stable visible-light-driven photocatalyst [36]. Carbon NPs' union with ZnO aids in enhancing the optical properties. A report explained the solution dispersion method of CDs loaded with ZnO and also naphthol blue black (NBB) azodye was tested using ZnO doped on CDs, as seen in Figure 8. Complete degradation of CDs occurs after 45 min whereas using pure CDs led to the degradation of 4.4% NBB was within the same period [40]. This reflects the fact that photocatalytic nature of CDs makes it useful for harmful dye degradation.

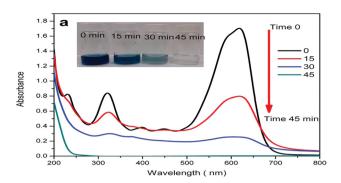


Figure 8: Absorption intensity differentiations of NBB azodye at different irradiation intervals in the existence of CDs/ZnO [40].

Waste water treatment can be executed by using CDs with other photocatalytic materials. Excellent photocatalytic characterization of silver orthophosphate (Ag₃PO₄) was explained by many reports. It can be used to degrade organic contaminants and water oxidation in spite of less tendency to be water soluble. CDs/Ag/Ag₃PO₄ and CDs/Ag₃PO₄ photocatalytic systems were prepared as mentioned by different studies by simple dispersion of CH₃COOAg and polyvinyl pyrrolidone in a CD solution, and Na₂HPO₄ was added drop wise, and the former was stirred at 25°C for 4 h in dark and the resultant material was dried in an oven at 50°C for 12 h [95]. After complex formation, characterization was examined by various techniques - as seen in Figure 9, SEM image detected the rhombic dodecahedral morphology of the complex with a size of 800-900 nm; and the UV-Vis spectroscopy explained that the complex absorbs in the range of 530-1,000 nm led to photocatalytic activity estimation and also its essential role in absorbing sunlight. Under visible light irradiation, methyl orange (MO) photocatalytic activity was shown by CDs/Ag/Ag₃PO₄ complex as MO dye was completely degraded within 10 min, on the contrary to CDs/Ag₃PO₄ complex which needed at least 25 min for degradation while pure Ag₃PO₄ requires 55 min to degrade, which means the prepared complex has the highest photostability and catalytic activity in addition to up-conversion PL exhibition. Photostability improvement, photocatalytic activity, and photocorrosion dodge of metal complex perform as an electron reservoir and take advantage of the entire spectrum of sunlight. By mixing the as-prepared mSiO₂ with glycerol and PEG-NH₂ at 230°C for 30 min, a C-dots@miSiO₂-PEG nanocomposite resulted, which was tested for drug delivery as it offered enough space for storing drug molecules, thereby performing as an efficient carrier. For example, controlled drug delivery was prepared using the water-soluble anticancer drug doxorubicin (DOX) was loaded into the

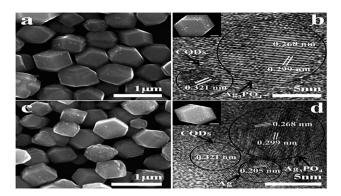


Figure 9: (a) SEM and (b) HRTEM images of CDs/Ag₃PO₄ complex photocatalyst. (c) SEM and (d) HRTEM images of CDs/Ag/Ag₃PO₄ complex photocatalyst [22].

nanocomposite by reducing the drug pre-release before drug absorption by the target cell [96]. BiVO₄, a semiconductor material, was used to improve the photocatalytic activity of complex formation with CDs. Degradation of methyl blue dye under visible light at room temperature was used to examine the photocatalytic ability of this complex [42]. Besides, semiconductor complexes considered an important example of photocatalytic activity of CD complexes, especially formation of semiconductor/CD complexes gives the maximum light energy [97]. The essential requirement for photocatalytic systems is improving the power conversion efficiency. Rhodamine B (RhB)/TiO2 complex was doped on CDs to give an example of CD complex synthesis to perform a semiconductor photoelectric conversion system that led to improved photocurrent density. By CDs' addition to the aqueous RhB solution, there is an increase in the UV-Vis absorption. Due to the presence of oxygen-bearing functional groups, the doped CDs were of greater solubility, and UV-Vis absorption becomes greater when CD was <10 nm. As CDs perform as an electron transfer intermediate, it enhances the charge recombination reduction which means complex system achieved PL quenching. All the previous results led to the knowledge that by using such CDs/dye/semiconductor complex systems, high-efficiency photoelectric systems can be achieved [98]. MO dye degradation can be achieved by N-doped CDs. Harmful chemicals present in water degradation can be achieved by photocatalytic activity of such complexes [99]. CD nanocomposites formed with different materials can be used in many fields such as optoelectronics, drug delivery, waste water treatment, etc. All the previous studies about CD nanocomposites give an idea about the novel composites with NP development to be used in numerous applications.

6 CD green synthesis

New research discussed green synthesis and found good salience. Preparation of value-added products from biodegradable waste is an important topic in the green chemistry field. The affirmation on green chemistry to decrease the use of high-end chemicals due to the toxicity concerns. The materials used in the green synthesis should be eco-friendly and cheap so they are produced from plants to be used as the starting materials. Green synthesis has many advantages, that is, it is biocompatible, non-time-consuming, less toxic, economical, and requires lower temperatures, so it is considered an interesting topic for researchers. Carbon can be extracted by green synthesis as it is available in all organic materials. One of the reports discussed CDs' green synthesis from orange peels by hydrothermal treatment [40], as it is a cheap renewable method to prepare chemical compounds from eco-friendly starting material. Hydrothermal synthesis uses high-pressure and high-temperature conditions, leading to perfect morphology of the structures and considered economic method because of using water as a solvent. First, washing the peels and then exposing it to sunlight for drying followed by oven drying at 150°C for 10 h followed by the addition of H₂SO₄ to wash the carbonized peels rinsed with water, soaked with sodium hypochlorite solution for 4 h, and after that washed with water until the pH of the washed water reached 7; the preheated oxidized orange peels were hydrothermally treated in a Teflon lined autoclave at 180°C for 12 h and then dichloromethane was added to the resulting solution and the separating solution centrifuged at 5,000 rpm for 20 min to obtain a brown solution of an aqueous suspension of CDs as shown in Figure 10. The QY was 12.7% and the resulted particles were 2-7 nm in diameter. Another report displayed a different method for hydrothermal synthesis of CDs from sugarcane bagasse, which contains large amount of carbon and so it is considered a renewable resource as CDs can be easily produced. Bagasse should be soaked in concentrated sulfuric acid for 24 h at room temperature and then washed it in hot water and alcohol until the filtrate become neutral, dried in an oven overnight to obtain carbonaceous blocks, mixed it with NaOH solution, heated for 1h, and then filtered for elimination of large particles. The solution obtained showed weak fluorescence due to the darkness of the filtrate. Alkaline hydrothermal treatment was implemented on the obtained solution, which gave a bright yellow solution. The QY was about 4.7% [100].



Figure 10: Formation of C-dots by using waste orange peels' hydrothermal treatment [40].

Another group prepared biocompatible CDs from aqueous extracts of Trapa bispinosa peels which is considered simple, rapid, and effective method. First, T. bispinosa peels were washed in water, soaked in cold water for 30 min, the peels were crushed in distilled water, centrifuged, and suspended in NaOH solution to produce a clear yellow CD suspension [101]. CDs were prepared by centrifugal separation from sugarcane juice; a facile method to prepare crystalline CDs from highly alkaline sugarcane juice. A few milliliters of NaOH solution were added drop wise into the sugarcane juice, which is stirred until a reddish brown solution is obtained, centrifuged at 5,000 rpm for 15 min, and checked under the UV light to have a dark green fluorescence. Sucrose density gradient centrifugation was executed to separate CDs from the mixture. As banana juice contains glucose, sucrose, fructose, and ascorbic acid, some recent study synthesized CDs from banana juice. Banana juice was extracted by crushing banana with a small amount of water, the pulp-free juice was extracted, mixed with ethanol, heated in a glass bottle closed with a cotton cork, then kept in an oven at 150°C for 4 h. When the solution cooled to room temperature, it turned into a dark brown product, which was then mixed with water and filtered to separate the residue. The aqueous solution was mixed with ethanol and centrifuged at 3,000 rpm for 15 min at room temperature, and the ethanol was evaporated completely to obtain the fluorescent CDs whose yield was 600 mg [37]. According to all the previous reports, CDs' green synthesis is of high prominence in the present-day script. These methods have many advantages as it uses very cheap starting materials so it considered cost-effective in addition to high yield of CDs [102]. All the previous methods in the abovementioned reports facilitate the discovery, and many other research of CDs synthesis explained the simple and costeffective methods. The CDs obtained from eco-friendly

starting materials are nontoxic and biocompatible and have fluorescence properties to be applicable in bioimaging technique.

7 CDs synthesis from commercially available food products

The commercial food products consumed in our dayto-day life can be discussed in many recent research. Amorphous CD synthesis is discussed in many recent research. Amorphous CDs are present in different caramel-containing food products like jaggery, sugar, bread, biscuits, and corn flakes; and CDs were extracted from the browner part of the bread, caramelized sugar, and jaggery. The materials were of caramel color under white light and blue fluorescence under UV light as shown in Figure 11. The lowest QY was from jaggery observed at 0.55% and the highest bone was from bread observed at 1.2%. CD morphology was spherical according to TEM images. Bread-extracted CDs possess the highest particle size while those extracted from sugar caramel have the lowest one. The amorphous nature of CDs was obvious by XRD analysis. Due to the presence of carboxyl and alcohol functional groups, the synthesized CDs were hydrophilic in nature [38]. All the pervious perceptions led to the affirmation of CDs' existence in these food products. Coffee grounds can be heated, dried, and separated to synthesize CDs. CDs' preparation from coffee grounds is a simple method as it includes the following four stages: dehydration, polymerization, carbonization, and passivation. As a result of dehydration, polymerization and nucleation occurred in addition to carbonization due to heating. Nuclei growth occurred by solute diffusion toward particle surfaces. Prepared CDs were characterized by using different techniques. The average diameter of the particles was ±2 nm according to TEM results. The existence of carbon atoms was confirmed by XRD as well as EDX data as showed in Figure 12. The amorphous nature of CDs was detected by Raman analysis. The fluorescence emission spectra exhibited a broad range from blue (400 nm) to red (600 nm). The emission peak position shifted to longer wavelengths when the excitation wavelength was increased and the intensity decreased. When excited at 365 nm, the strongest PL of 3.8% occurred at 400 nm and the CDs' fluorescence could only be noticed when they were surface passivated [103]. CD synthesis aids in the management of wastes as the mentioned in the previous reports. CDs can

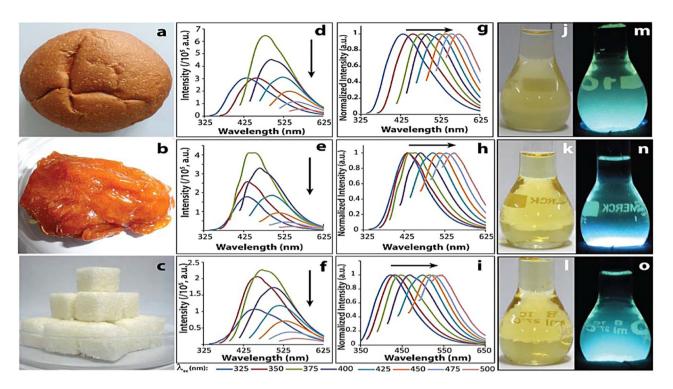


Figure 11: (a-c) Photographs of commercial bread, jaggery, and sugar. (d-i) Excitation-wavelength-dependent emission spectra of CDs from bread, jaggery, and sugar caramel. (j-l) Photographs of CDs dispersion from bread, jaggery, and sugar caramel observed under white light, and (m-o) the same under UV light [38].

be produced from many carbohydrate-rich food products. Waste management participation and green chemistry help in promoting scientific research. Recent improvement in CD synthesis was achieved using greener methods. Green chemistry use compared to the physical methods facilitates the process and offers an eco-friendly synthesis.

8 Applications of C-dots

8.1 Sensing

The most important CD application is the sensing field. On the basis of the fluorescence properties and the surface of CDs' functional groups, such as $\mathrm{Hg^{2+}}$ detection and biological thiols (Figure 13a) [104] the electrons and the holes of CDs can be restructured and the fluorescence of CDs can be quenched by $\mathrm{Hg^{2+}}$. The fluorescence of CDs can be restored as $\mathrm{Hg^{2+}}$ has strong coordination ability with thiols and recognize the "turn-off" detection of $\mathrm{Hg^{2+}}$ and "turn-on" detection of thiols. By using fluorescence quenching from the coordination between CDs' amino

group and Cu^{2+} , Dong et al. [105,106] synthesized a branched PEI-modified CDs for Cu^{2+} detection in a river samples with a 6-nM detection limit of 9 (Figure 13b). Quenching effect of Cu^{2+} on the fluorescence CDs enhances image detection of cellular Cu^{2+} , and probing the Cu^{2+} position in cells (Figure 13c) was attained by using AE-TPEA-CDs-CdSe/ZnS nanocomposite [107]. Cellular Cu^{2+} was detected by Vedamalai et al. [108] who used synthesized Cu^{2+} -sensitive CDs. Acetyl choline (ACh), which transformed to choline by acetylcholinesterase by using CDs reduced graphite oxide (CDs@RGO) composites whereas choline could beget H_2O_2 in the existence of choline oxidase (Figure 13d) [109]. The quenching C-dots@RGO fluorescence

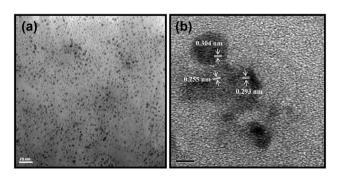


Figure 12: (a) TEM and (b) HRTEM images of CDs [103].

ability of H₂O₂ assists the ACh quantitative detection with 30 PM detection limit. Biological active substances were detected by CDs-Ag/CDs-Au nanocomposite which has another trial in biosensing field [110-112]. The detection limit of glutathinone by using fluorometric/colorimetric bimodal sensor based on CDs-Au composites was 50 nM [111]. In colorimetric method, the detection limit was 0.18 and 1.6 µM for H₂O₂ and glucose, respectively [110]. A boric acid-modified CDs were prepared by Zhang et al. [113] for the glucose detection with the limit of 0.03 nM, taking into consideration the strong affinity between glucose and boric acid (Figure 13e). Glucose was used as a precursor to synthesize a CD via neutralizing the heat process [114]. Because the adjacent hydroxyl groups from glucose remained on the CDs' surface, these CDs exhibited a perfect affinity to boric acid, so CDs had an ability to detect glycoprotein. Chemiluminescence (CL) and electrochemiluminescence (ECL) biosensors were well constructed by using CDs. Lin et al. invented a CD-based CL sensor for nitride detection as they discovered that CDs had CL signal in the presence of nitrous acid peroxide (Figure 13f) [115]. To track Cu²⁺ in the mice brain, Shao et al. [116] utilized CDs and Tri prime and amplification (TPEA), which showed electrochemical response. Cd2+ detection with a

limit of 13 nM was achieved by Li et al. [117] who constructed a graphene nanodots-based ECL sensor (Figure 13g).

8.2 Drug delivery and gene transfer

One of the interesting fields of CDs application is drug delivery and gene transfer. Recognition of cancer cells can be achieved by using folic acid-modified CDs by amide condensation reaction which offered a good method to improve cell screening and disease diagnosis [118]. PEI-modified CDs have a positive surface potential, so it could absorb the negative DNA to transfer gene [119]. Liu et al. [119] discovered that CDs not only had fluorescence characterization to display the plasmid DNA distribution during the transfer process but also offered detailed information for plasmid DNA physiological function research. After transfection for 3 h, CD DNA composites could enter the cell. During the transport process, different wavelength excitation CDs preserved multicolor fluorescence property.

Lai et al. [120] synthesized a PEG-modified CDs and fulfilled DOX loading and delivery. DOX in cells' release

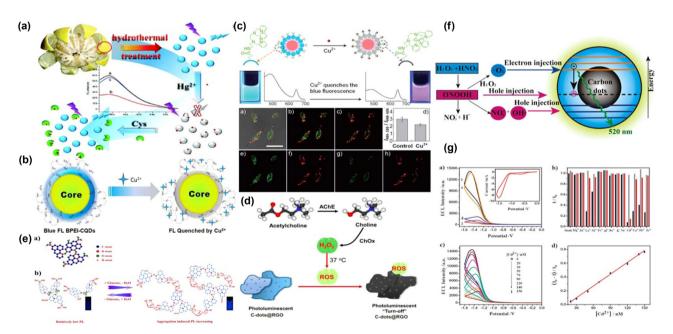


Figure 13: Bio-/chemsensing applications of carbon dots. (a) Schematic representation of an Hg²⁺/cys (cysteine) biosensor based on CDs prepared from pomelo peel [104]. (b) Schematic representation of a Cu²⁺ biosensor based on CDs-BPEI (branched polyethylenimine) [105]. (c) Fabrication of a ratiometric fluorescent sensor based on a CDs-CdSe/ZnS hybrid material for detection of copper ions, and *in vivo* imaging and monitoring of cellular copper ions [107]. (d) Schematic representation of detection of ACh by C-dots@RGO in a TURN-OFF Strategy [109]. (e) Schematic representation of boron-doped graphene quantum dots, and mechanism for glucose sensing by boron-doped graphene quantum dots [113]. (f) Mechanism for nitrite sensing based on CL of CDs induced by peroxynitrous acid [115]. (g) ECL sensing for Cd²⁺ based on graphene quantum dots [117].

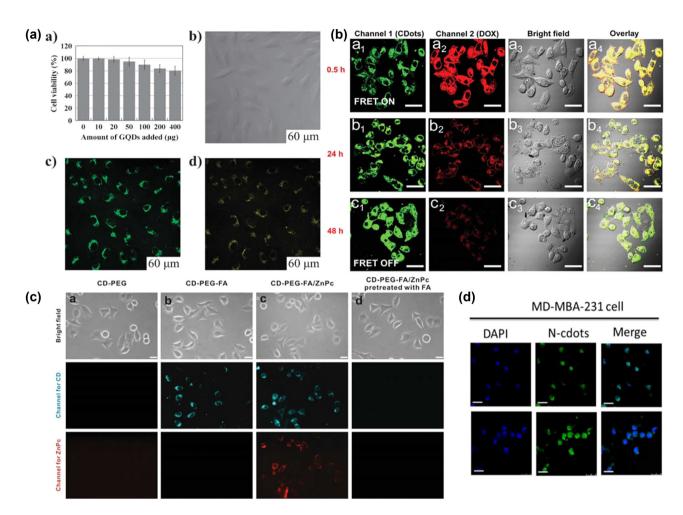


Figure 14: (a) Multicolor images of cells treated with CDs [16]. (b) Fluorescence images of cells treated with CDs-DOX at 0.5, 24, and 48 h [133]. (c) Fluorescence images of (a) CDs-PEG, (b) CDs-PEG-FA, (c) CDs-PEG-FA/ZnPc, and (d) CDs-PEG-FA/ZnPc pretreated with FA [123]. (d) Fluorescence images of MD-MBA-231 cells pretreated with "biomolecule-mimicking" CDs [134].

process is exhibited by fluorescence images. CDs acquired cytoplasm green fluorescence. At the time of treatment, DOX releases into the nucleus after entering the cells, causing the appearance of the red fluorescence. To observe drug delivery, Chowdhuri et al. [121] joined CDs with metal organic frameworks. Wang et al. [122] used the mixture of chitosan, PEG, and CDs to attain a NIR light/pH dual-responsive hybrid gel. All the previously mentioned research indicate CDs' application in drug delivery and gene transfer and corroborate CDs' clinical applications.

8.3 Biological imaging

8.3.1 In vitro imaging

In vitro imaging is executed to offer plentiful information on the imaging ability, distribution and cytotoxicity of probes in cells. CDs have many effective uses in cell

transfection such as HeLa [123-129], human neutral stem cells [130] 4-T1 [131], NIH-3T3 [132], A549 [108,133], HepG-2 [126], etc. Through endocytosis, CDs enter into the cells and concentrate in the cytoplasm [126]. CDs may enter cell nucleus but only a few reports were discussed this way. Zhu et al. [16] successfully used CDs in cell imaging as they synthesized fluorescent CDs with low cytotoxicity in solvothermal method (Figure 14a). Nitrogen-doped CDs displayed an excitation which was dependent on fluorescence emission for multicolor fluorescence imaging [126] by carbonization extraction. Multifunctional tumor targeting probe preparation can be prepared by CDs' surface modification. Tang et al. [133] discussed cancer cell recognition, drug transport, and fluorescence imaging by using CDs with folic acids and DOX (Figure 14b). Bhunia et al. [129] explained surface modification of CDs with folic acid by preparing a series of CDs with different fluorescence and observed target recognition. To make the tumor abilities obvious and

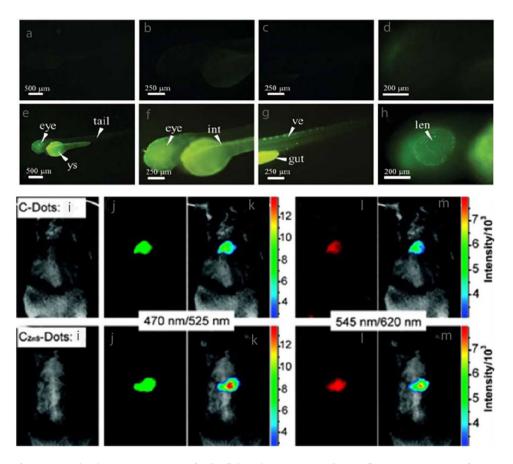


Figure 15: (a-h) Fluorescence image of zebrafish with CDs [139] and (i-m) fluorescence image of rat injected with CDs [87].

execute photothermal therapy (Figure 14c), Choi et al. [123] used CDs' altered with zinc phthalocyanine and folic acid. Dopamine-neutralization heat treatment [134] was used to synthesize a biomolecule mimicking CDs. The dopamine-mimicking CDs could "hoax" nuclear membranes to fulfill nuclear localization and imaging [134] because the functional groups of dopamine were maintained in this CDs (Figure 14d). Elimination of autofluorescence interference can be achieved by CDs' two-photon, cell imaging upon NIR light excitation (800-900 nm) [134]. Under the 880 nm laser excitation, Yang et al. [87] attained green fluorescence images. Cell nucleus twophoton imaging was fulfilled by Zhang et al. [135] who utilized C₃N₄ nanodot as a probe. For living cells and tissue imaging, Kong et al. [136] synthesized pH-sensitive CDs because of its two-photon fluorescence property.

8.3.2 In vivo imaging

Many essential medicine science such as disease progression, improved mechanism, and pattern formation were achieved using zebrafish due to its well-defined

developmental stages and optical imaging amenability [137]. The fluorescence imaging model was the reason to choose zebrafish; CDs were found to heap up in eyes and yolk sac selectively (Figure 15) [114,138,139]. CDs were preserved over 60 h, so they are suitable for observing the long-term zebrafish developmental stages. PEGpassivated CDs and Zns-doped CDs (CZns-dots-PEG) [80] were used for vivo imaging in spite of their blue and green emission, which were not suitable for in vivo imaging. Under different excitations, no apparent toxicity was noticed in tissues and organs [80], green and red emission fluorescence images were obtained. Using CD probes by administering interdermal injection, the lymph of mice was labeled. The metabolism speed of the nonmodified CDs was faster than the metabolism speed of PEG-modified CDs. CD fluorescence was seen at the stomach past intravenous injection and heaped up in the bladder after 1 h. Four h post injection, the fluorescence signal decreased and heaped up in the kidney, explaining that CDs are excreted from mice by urine [80]. Under different excitations (455-704 nm), Tao et al. [140] obtained CD in vivo images. When CDs were used in vivo imaging [141], CDs could enter into brain and appeared blue.

8.4 CDs food safety applications

Chemical contaminations such as metal ions, anions, pesticides, veterinary drugs, and banned additive are the main reasons for many health problems. This demands very effective and actual methods for guaranteeing food safety. In the food quality and safety area, a number of sophisticated CD-based biosensors are required. The main concepts of these CDs-based biosensors can be divided into four kinds:

- 1. Direct fluorescence quenching (turn-off),
- 2. Direct fluorescence boost (turn-on),
- 3. Fluorescence quenching again (off-on), and
- 4. Fluorescence quenching again (off-in-off).

The fluorescence quenching mechanisms implicate:

- 1. Dynamic quenching,
- 2. Static quenching,
- 3. Fluorescence resonance energy transfer (FRET), and
- 4. Inner filter effect (IFE) [142].

CDs' excited states return to ground state by collision with quencher and produces energy transfer/charge transfer (ET/CT), which is known as dynamic quenching while the static quenching results when CDs form nonfluorescent ground state complex with quencher. FRET and IFE are on the contrary mechanisms that demand two essential factors, namely, spectrum overlap and the distance between CDs and quencher (receptors). At less than 10 nm distance, FRET occurs while at more than 20 nm distance, IFE takes place. Some targets' quench lead to improve the CDs' fluorescence immediately, permitting merely sensors' design for some targets (metal ions, veterinary drugs, bacteria, and some functional components in foods) [29,56,143,144]. Restoring CDs' fluorescence after supplementing the target into quencher-containing fluorescence quenching system is known as fluorescence quenching (off-on-off) strategy, which depends on quencher, restorer, and target. Where the fluorescence recovers due to the interaction between quencher and restorer and then quench again; to enable the interaction of target and restorer, the "off-on" and "offon-off" strategies can be utilized to determine other targets (such as anions, pesticides, veterinary drugs, and some functional components in foods) [21,23].

synthesis and gave a brief summary of the latest research development of CDs in fluorescent properties and their applications that can give explanation of the optical properties among the fluorescent properties. It is obvious that understanding these properties has a long way to go. We focused on the CDs' greener methods which are more economical, less complicated, and less consuming, have lower toxicity, better compatibility, nondestructive nature, and improved optical properties. High QY of CDs by using green synthesis methods is not difficult any more. CDs' research emphasizes on the development of optical properties and discovery of red fluorescence of CDs. Moreover, functional modifications play another essential role in CD improvement: CD fluorescence focuses on blue-green emission and few red-emitting CDs were discussed [145]. Fluorescent CDs can be as follows:

- CDs' optical properties could be enhanced by tuning carbon sources.
- Red-emitting CDs can be obtained from phenylenediamines as carbon source as Jiang et al. discussed [146]. Polythiophene as a carbon source exhibited CDs' fluorescence emission shifting to 650–700 nm as Ge et al. explained [147].
- 3. CDs' fluorescence may be improved by using appropriate passivated agents.
- 4. The bandgap spacing of CDs' adjusting fluorescence yield and emission range can be done by introducing heteroatoms.
- 5. Functional molecules can be used to modify carboxyl and amino groups on the surface of CDs by amide condensation [39].
- 6. CDs' blue shift emission and fluorescence quenching can be achieved by fluorescence resonance energy transfer between CDs and modifications which must be taken into consideration through the following points:
 - a. The distance between CDs and modifications could be increased by modified method choice such as choosing appropriate bridging content (such as silicon beads, inorganic clay, etc.) led to resonance energy transfer reduction.
 - Building novel CDs for multimodal imaging can be executed by multimodal functionalization and multimodal imaging factor introduction.

9 Conclusion

This review discussed the discovery, structure, and composition of CDs as well as CDs' nanocomposites and green

10 Example

One pot hydrothermal treatment of gadopentetic acid mixture, trihydroxy methyl aminomethane, and betaine

[148] to attain Gd-doped CDs with a size of 3–4 nm was explained by Bourlinos et al. [148,149].

Heteroatom doping [150], large conjugated system formation [78], and change the reaction medium [151] have been executed to enhance QY or red-shift degree. Due to the surface groups (such as carboxylic acids and hydrolysis), most of the synthesized CDs are water soluble. Further surface functionalization of CDs can be achieved by doping with heteroatoms (such as N and S) conferring specific chemical reactivity to CDs permitting CDs to be specific probes/bioconjugates in detection application which is very critical toward surface functionalization [54,152]. Surface groups' significance in linkage with targets is to enhance the CD-based sensors selectivity in food safety applications [153-159]. Toxic elements' detection by CDs depends on energy-transfer capability. CDs can be used in biological sample analyses due to its biocompatible nature. There is still a lot of areas for discovering CDs' functionality with various elements. In the near future, CDs will lighten human lives by slapping the way for more advanced techniques.

11 Future perspectives

11.1 Green chemistry as an essential approach to attain heteromeric superstructures

11.1.1 CDs eco-friendly and sustainable synthesis

Green chemistry is a linkage of important concepts that minimizes or eliminates the implementation or generation of hazardous substances in the design, manufacture, and chemical product implementations. Green chemistry involves using none or less dangerous chemical synthesis, applying safer and nontoxic chemicals, solvents, and processes [160]; for example, biowastes can be utilized as sustainable and cheap carbon sources for CD synthesis. Spherical water-soluble CDs (about 1-3 nm) have been prepared from lemon peel waste, which is the prevailing cost-effective hydrothermal strategy, and the ensuring stable CDs were discovered to be oxygen rich in surface functionalities and displayed the water-soluble and unique PL properties with QY of about 14%. The synthesized CDs were used to design an economic, green, and highly sensitive fluorescent probe for Cr⁶⁺ ions' detection limit of about 73 nM. Water-soluble CDs considered simple, fast sensitive, and selective Cr⁶⁺ detection is suitable technique in operations of water purification.

TiO₂ - water - soluble CDs composite was inspected utilizing methylene blue dve as a model pollutant because CDs have been immobilized over electrospan. TiO₂ nanofibers and the emanating photocatalytic activity being about 2.5 times more than that of titanium dioxide nanofibers [161]. Water-soluble fluorescent carbon quantum dots (~260-400 nm) were reported via a simple one-step hydrothermal treatment utilizing Tamarindus *indica* leaves. These bioconvenient CDs can be potentially executed in sensing, bioimaging, disease diagnostics, and other analytical implementations [162]. Another fluorescent CD synthesis facile approach requires the hydrothermal treatment of pineapples (Ananas comosus) and calamansi (Citrofortunella microcarpa) wastes [163]. Escherichia coli, fluoresced with CDs as operative probe, has been used for bioimaging applications. Generation of hydrogen bonds between the bacterial cells and CDs eases the quantum dots' attachment onto the bacteria [163]. Greener ozone oxidation of ignite coal was utilized for well-dispersed CDs was utilized for well-dispersed CDs (~2-4 nm) which is copious, cost-effective, and involves oxygen-rich functional groups with suitable water-solubility, optical characteristics, and yield reaching 35% [164]. Synthesized quantum dots utilized for Fe³⁺ detection with remarkable sensitivity and selectivity as well as desirable anti-interference had quenching effects but the CDs' intensity showed clear linear responses to the Fe^{3+} concentrations (10–150 μ mol L^{-1}).

With the detection limit of 0.26 μ mol L⁻¹ [164], in one pot by using cydonia oblonga powder as carbon precursors via microwave irradiation which shows maximum emission at 450 nm if excited at 350 nm with a QY of 85% highly stable and luminescent multi-color CDs would generate. Also CDs were prepared through microwave heating technique in 30 min in comparison to those produced hydrothermally in a Teflon-linen stainless steel body autoclave at 200°C [165]. Fluorescent CDs' greener synthesis by using citrus lemon juice (~2-10 nm) was achieved utilizing hydrothermal approach [166] which showed high PL intensity was obtained at pH of 6 in cell imaging [166]. CDs can be utilized as a fluorescent probe. Amorphous fluorescent CDs can be obtained from orange peel waste by utilizing hydrothermal carbonization approach at mild temperature [40]. Bright green luminescent graphitic carbon nitride doped with oxygen and sulfur were prepared using microwave treatment of citric acid and thiourea. They exhibited excitation wavelength and pH-dependent luminescence behaviors in the visible range. Moreover, the QY of 31.67% shows a strong impedance to the interference of high ionic strength environment, and perfect biocompatability as estimated by the cell viability assay [167]. Prunus avium fruit

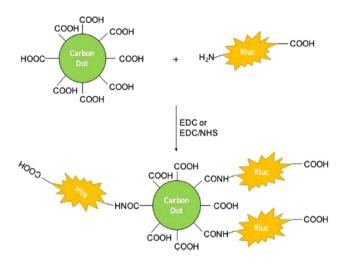


Figure 16: EDC- or EDC/NHS-mediated conjunction of bioluminescent proteins, *Renilla luciferase* (Rluc) to CDs. Both conjunction couples the free surface carboxylic acids on CDs with the amine group on the N-terminal of Rluc. The close proximity between the two molecules allows for bioluminescence resonance energy transfer [186].

extracts were used for fabricating fluorescent nitrogen-doped CDs (~7 nm) which exhibited blue fluorescent characteristics fluorescence. The green CDs were utilized as fluorescent probes for Fe³⁺ ions' biological detection in water with selectivity and sensitivity and for bioimaging of the tested cells [168]. The simple microwave process was entered for fluorescent nitrogen-doped CDs which were hydrothermally synthesized by applying L-ascorbic acid (as the carbon precursor) and β -aniline (perform as nitrogen dopant) with distinct blue fluorescence and low cytotoxicity. These CDs can be utilized as staining probes for confocal cellular imaging [169].

11.1.2 CDs self-illumination by bioluminescence resonance energy transfer

The CDs of the emerging fluorescent NPs (FNPs) display excitation-dependent emission and chemical stability as their identical multicolor PL have magnetized inclusive attention due to the various implementations in photovoltaic devices, bioimaging, etc. [170,171]. Various agents influence CDs' fluorescence:

- 1. Quantum size effect results from CDs nanoscale size.
- 2. Surface oxidation degree.
- 3. Surface functional group like carbonyl and imines [172].

To begin the FNPs excitation, external high-power light source is essential which may generate photocytotoxicity and tissue damage [173]. There are different

mechanisms in which fluorescence is executed without external high-power source, for instance, CL resonance energy transfer (CRET) and bioluminescence resonance energy transfer (BRET) as they include a nonradiative energy transfer between a light energy donor and a luminescence acceptor. The donor molecular emission enables covering the acceptor molecular excitation or absorption when the distance between donor and acceptor molecules is close, i.e., <10 nm, so these luminescence resonance energy transfers can occur [174-176]. In order to excite the CRET acceptor molecule, CL reactions in the main reactions in which CRET is relied. While emitting photon to release energy [177], CL reaction causes an electronically excited product followed by electron dropping to ground state [177]. Modern limitations appeared when this successfully removes the external light source requirement. Metallic ions or catalyst enzymes are usually used as catalysts [178] to lower the initial CL reaction activation energy and also cofactors may also be essential for the initial reaction to take place. Biocompatibility may be jeopardized with the use of metallic ions, moreover, the required catalyst and cofactors may not be compatible with microenviroment. Natural bioluminescent molecules exist in firefly, jellyfish, and other organisms, as the light energy donor [179] unlike CRET and BERT. For instance, Renilla luciferase (Rluc) enzymatic reaction with its substrate, coelenterazine, can generate luminescence at 480 nm [180]. BERT is more suitable compared to CRET as catalysts and cofactors are not required for the reaction to happen. In bioconjugation of bioluminescent proteins onto nanostructures, e.g. gold NPs, semiconductors' quantum dots have been used in biosensors and bioimaging without the external power supply [181–184]. Compared to the BRET pair containing the acceptor made of organic dye and/or other nanostructures, CDs could be particularly of interest in using for BRET pair, and modify functional groups for further bioconjugation. CDs clearly show better bioconvenient ability than other fluorescence nanostructures, e.g., quantum dots. Some studies debated CDs' self-illuminescence proteins. Herein, explanation of a method for self-illuminated CD synthesis by using BERT technique. Figure 16 explained the bioconjugation of Rluc onto CDs' strategies through carboxyl-to-amine crosslinking. Two essential water-soluble carbodiimide crosslinkers, N'-(3-dimethyl aminopropyl)-N-ethylcarbodiimide and EDC associated with N-hydroxysuccinimide, were investigated. Other different factors including the CD ratio and Rluc and the bioconjunction reaction time are investigated for the successful execution of BERT [185].

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References

- [1] Xie Y, Filchakova O, Yang Q, Yesbolatov Y, Tursynkhan D, Kassymbek A, et al. Inhibition of cancer cell proliferation by carbon dots derived from date pits at low-dose. ChemistrySelect. 2017;2:4079-83. doi: 10.1002/ slct.201700575.
- [2] Baig MB, Al-Zahrani KH, Schneider F, Straquadine GS, Mourad M. Food waste posing a serious threat to sustainability in the Kingdom of Saudi Arabia a systematic review. Saudi J Biol Sci. 2019;26(7):1743–52. doi: 10.1016/j.sjbs.2018.06.004.
- [3] De Menna F, Dietershagen J, Loubiere M, Vittuari M. Life cycle costing of food waste: a review of methodological approaches. J Waste Manag. 2018;73:1–13. doi: 10.1016/ j.wasman.2017.12.032.
- [4] Galanakis CM. Food waste recovery: prospects and opportunities. Sustainable food systems from agriculture to industry, improving production and processing. Cambridge: Academic Press; 2018. p. 1–442. Paperback ISBN: 9780128119358, eBook ISBN: 9780128119617.
- [5] Rafiq S, Kaul R, Sofi SA, Bashir N, Nazir F, Nayik GA. Citrus peel as a source of functional ingredient: a review. J Saudi Soc Agric Sci. 2016;17(4):351–8. doi: 10.1016/j.jssas.2016.07.006.
- [6] Hu Q, Gong X, Liu L, Choi MMF. Characterization and analytical separation of fluorescent carbon nanodots.
 J Nanomater. 2017;2017:1–23. doi: 10.1155/2017/1804178.
 Article ID 1804178.
- [7] Mishra RK, Ha SK, Verma K, Tiwari SK. Recent progress in selected bionanomaterials and their engineering applications: an overview. J Sci Adv Mater Devices. 2018;3(3):263–88. doi: 10.1016/j.jsamd.2018.05.003.
- [8] Safarik I, Baldikova E, Prochazkova J, Safarikova M, Pospiskova K. Magnetically modified agricultural and food waste: preparation and application. J Agric Food Chem. 2018;66(11):2538–52. doi: 10.1021/acs.jafc.7b06105.
- [9] Xu X, Ray R, Gu Y, Ploehn HJ, Gearheart L, Rakeret K, et al. Electrophoretic analysis and purification of fluorescent single-walled carbon nanotube fragments. J Am Chem Soc. 2004;126(40):12736-7. PMID: 15469243. doi: 10.1021/ ja040082h.

- [10] Jiang C, Wu H, Song X, Ma X, Wang J, Tan M. Presence of photoluminescent carbon dots in Nescafe® original instant coffee: applications to bioimaging. Talanta. 2014;127:68–74. PMID: 24913858. doi: 10.1016/j.talanta.2014.01.046.
- [11] Sharma V, Tiwari P, Mobin SM. Sustainable carbon-dots: recent advances in green carbon dots for sensing and bioimaging. J Mater Chem B. 2017;5(45):8904–24. doi: 10.1039/ C7TB02484C.
- [12] Zhang X, Jiang M, Niu N, Chen Z, Li S, Liu S, et al. Natural-product derived carbon dots: from natural products to functional materials. ChemSusChem. 2018;11(1):11–24. PMID: 29072348. doi: 10.1002/cssc.201701847.
- [13] Zhang J, Wang H, Xiao Y, Tang J, Liang C, Li F, et al. A simple approach for synthesizing of fluorescent carbon quantum dots from Tofu wastewater. Nanoscale Res Lett. 2017;12(1):611–8. PMID: 29188541, PMCID: PMC5707215. doi: 10.1186/s11671-017-2369-1.
- [14] Zhou Y, Liu Y, Li Y, He Z, Xu Q, Chen Y, et al. Multicolor carbon nanodots from food waste and their heavy metal ion detection application. RSC Adv. 2018;8(42):23657–62. doi: 10.1039/C8RA03272F.
- [15] Huang CC, Hung YS, Weng YM, Chen W, Lai YS. Review: sustainable development of carbon nanodots technology: natural products as a carbon source and applications to food safety. Trends Food Sci Tech. 2019;86:144–52. doi: 10.1016/ j.tifs.2019.02.016.
- [16] Zhu S, Zhang J, Qiao C, Tang S, Li Y, Yuan W, et al. Strongly green-photoluminescent graphene quantum dots for bioimaging applications. Chem Commun. 2011;47:6858–60.
- [17] Ding H, Ji Y, Wei JS, Gao QY, Zhou ZY, Xiong HM. Facile synthesis of red-emitting carbon dots from pulp-free lemon juice for bioimaging. J Mater Chem B. 2017;5(26):5272-7. doi: 10.1039/C7TB01130J.
- [18] Miao X, Yan X, Qu D, Li D, Tao FF, Sun Z. Red emissive sulfur, nitrogen codoped carbon dots and their application in ion detection and theraonostics. ACS Appl Mater Interfaces. 2017;9(29):18549–56. PMID: 28508626. doi: 10.1021/acsami.7b04514.
- [19] Gao W, Song H, Wang X, Liu X, Pang X, Zhou Y, et al. Carbon dots with red emission for sensing of Pt²⁺, Au³⁺, and Pd²⁺ and their bioapplications in vitro and in vivo. ACS Appl Mater Interfaces. 2018;10(1):1147–54. PMID: 29235348. doi: 10.1021/acsami.7b16991.
- [20] Shi X, Wei W, Fu Z, Gao W, Zhang C, Zhao Q, et al. Review on carbon dots in food safety applications. Talanta. 2019;194:809-21. doi: 10.1016/j.talanta.2018.11.005.
- [21] Borse V, Thakur M, Sengupta S, Srivastava R. N-doped multifluorescent carbon dots for 'turn off-on' silver-biothiol dual sensing and mammalian cell imaging application. Sens Actuators B Chem. 2017;248:481–92. doi: 10.1016/ j.snb.2017.03.158.
- [22] Fong JFY, Chin SF, Ng SM. A unique "turn-on" fluorescence signalling strategy for highly specific detection of ascorbic acid using carbon dots as sensing probe. Biosens Bioelectron. 2016;85:844–52. PMID: 27290666. doi: 10.1016/j.bios.2016.05.087.
- Gao Z, Wang L, Su R, Huang R, Qi W, He Z. A carbon dot-based "off-on" fluorescent probe for highly selective and sensitive detection of phytic acid. Biosens Bioelectron. 2015;70:232–8. PMID: 25829220. doi: 10.1016/j.bios.2015.03.043.

[24] Dong Y, Cai J, You X, Chi Y. Sensing applications of luminescent carbon based dots. Analyst. 2015;140(22):7468-86. doi: 10.1039/C5AN01487E.

- [25] Guo Y, Zhang L, Zhang S, Yang Y, Chen X, Zhang M. Fluorescent carbon nanoparticles for the fluorescent detection of metal ions. Biosens Bioelectron. 2015;63:61–71. PMID: 25058940. doi: 10.1016/j.bios.2014.07.018.
- [26] Xiao L, Sun H. Novel properties and applications of carbon nano-dots. Nanoscale Horiz. 2018;3(6):565-97. doi: 10.1039/C8NH00106E.
- [27] Qu JH, Wei Q, Sun DW. Carbon dots: principles and their applications in food quality and safety detection. Crit Rev Food Sci Nutr. 2018;58(14):2466. doi: 10.1080/ 10408398.2018.143771275.
- [28] Hou J, Dong G, Tian Z, Lu J, Wang Q, Ai S, et al. A sensitive fluorescent sensor for selective determination of dichlorvos based on the recovered fluorescence of carbon dots-Cu(II) system. Food Chem. 2016;202:81–7. PMID: 26920268. doi: 10.1016/j.foodchem.2015.11.134.
- [29] Feng Y, Zhong D, Miao H, Yang X. Carbon dots derived from rose flowers for tetracycline sensing. Talanta. 2015;140:128-33. PMID: 26048834. doi: 10.1016/ j.talanta.,2015.03.038.
- [30] Wang N, Wang Y, Guo T, Yang T, Chen M, Wang J. Green preparation of carbon dots with papaya as carbon source for effective fluorescent sensing of iron(III) and Escherichia coli. Biosens Bioelectron. 2016;85:68–75. PMID: 27155118. doi: 10.1016/j.bios.2016.04.089.
- [31] Lei CH, Zhao XE, Jiao SL, He L, Li Y, Zhu SY, et al. A turn-on fluorescent sensor for the detection of melamine based on the anti-quenching ability of Hg²⁺ to carbon nanodots. Anal Methods. 2016;8(22):4438-44. doi: 10.1039/ C6AY01063F.
- [32] Riggs JE, Guo Z, Carroll DL, Sun YP. Strong luminescence of solubilized carbon atoms. J Am Chem Soc. 2000;122(24):5879-80. doi: 10.1021/ja9942282.
- [33] Lin Y, Zhou B, Martin RB, Henbest KB, Harruff BA, Riggs JE, et al. Visible luminescence of carbon nanotubes and dependence on functionalization. J Phys Chem B. 2005;109(31):14779–82. doi: 10.1021/jp053073j.
- [34] Hu Q, Gong X, Liu L, Choi MMF. Review article: characterization and analytical separation of fluorescent carbon nanodots. Hindawi J Nanomater. 2017;2017:1–23. doi: 10.1155/2017/1804178.
- [35] Liu L, Li YF, Zhan L, Liu L, Huang CZ. One-step synthesis of fluorescent hydroxyls-coated carbon dots with hydrothermal reaction and its application to optical sensing of metal ions. Sci China Chem. 2011;54:1342-7. doi: 10.1007/s11426-011-4351-6.
- [36] Maeda K, Takata T, Hara M, Saito N, Inoue Y, Kobayashi H, et al. GaN:ZnO solid solution as a photocatalyst for visible-light-driven overall water splitting. J Am Chem Soc. 2005;127(23):8286–7. doi: 10.1021/ja0518777.
- [37] De B, Karak N. A green and facile approach for the synthesis of water soluble fluorescent carbon dots from banana juice. RSC Adv. 2013;3(22):8286–90. doi: 10.1039/C3RA00088E.
- [38] Sk MP, Jaiswal A, Paul A, Ghosh SS, Chattopadhyay A. Presence of amorphous carbon nanoparticles in food caramels. Sci Rep. 2012;2:383. PMID: 22540029, PMCID: PMC3337553. doi: 10.1038/srep00383.

- [39] Baker SN, Baker GA. Luminescent carbon nanodots: emergent nanolights. Angew Chem Int Ed Engl. 2010;49(38):6726-44. PMID: 20687055. doi: 10.1002/ anie.200906623.
- [40] Prasannan A, Imae T. One-pot synthesis of fluorescent carbon dots from orange waste peels. Ind Eng Chem Res. 2013;52(44):15673-8. doi: 10.1021/ie402421s.
- [41] Liu H, Ye T, Mao C. Fluorescent carbon nanoparticles derived from candle soot. Angew Chem Int Ed Engl. 2007;46(34):6473–5. PMID: 17645271. doi: 10.1039/ b902509i.
- [42] Tang D, Zhang H, Huang H, Liu R, Han Y, Liu Y, et al. Carbon quantum dots enhance the photocatalytic performance of BiVO₄ with different exposed facets. Dalton Trans. 2013;42(18):6285–9. doi: 10.1039/C3DT50567G.
- [43] Wang X, Feng Y, Dong B, Huang J. A mini review on carbon quantum dots: preparation, properties, and electrocatalytic application. Front Chem. 2019;7:671. doi: 10.3389/ fchem.2019.00671.
- [44] Ghosh S, Chizhik AM, Karedla N, Dekaliuk MO, Gregor I, Schuhmann H, et al. Photoluminescence of carbon nanodots: dipole emission centers and electron-phonon coupling. Nano Lett. 2014;14(10):5656. PMID: 25247753. doi: 10.1021/nl502372x.
- [45] Wu M, Wang Y, Wu W, Hu C, Wang X, Zheng J, et al. Preparation of functionalized water-soluble photoluminescent carbon quantum dots from petroleum coke. Carbon. 2014;78:480-9. doi: 10.1016/j.carbon.2014.07.029.
- [46] Gude V, Das A, Chatterjee T, Mandal PK. Molecular origin of photoluminescence of carbon dots: aggregation induced orange-red emission. Phys Chem Chem Phys. 2016;18(40):28274–80. doi: 10.1039/C6CP05321A.
- [47] Song Y, Zhu S, Zhang S, Fu Y, Wang L, Zhao X, et al. Investigation from chemical structure to photoluminescent mechanism: a type of carbon dots from the pyrolysis of citric acid and an amine. J Mater Chem C. 2015;3(23):5976-84. doi: 10.1039/C5TC00813A.
- [48] Schneider J, Reckmeier CJ, Xiong Y, Seckendorff MV, Susha AS, Kasak P, et al. Molecular fluorescence in citric acid-based carbon dots. J Phys Chem C. 2017;121(3):2014-22. doi: 10.1021/acs.jpcc.6b12519.
- [49] Liu C, Zhang P, Tian F, Li W, Li F, Liu W. One-step synthesis of surface passivated carbon nanodots by microwave assisted pyrolysis for enhanced multicolor photoluminescence and bioimaging. J Mater Chem. 2011;21(35):13163-7. doi: 10.1039/C1JM12744F.
- [50] Zheng B, Liu T, Paau MC, Wang M, Liu Y, Liu L, et al. One-pot selective synthesis of water and organic soluble carbon dots with green fluorescent emission. RSC Adv. 2015;5(15):11667-75. doi: 10.1039/C4RA16529B.
- [51] Zhu S, Meng Q, Wang L, Zhang J, Song Y, Jin H, et al. Highly photoluminescent carbon dots for multicolor patterning, sensors, and bioimaging. Angew Chem Int Ed. 2013;52(14):3953–7. doi: 10.1002/anie.201300519.
- [52] Zhang Y, Wang Y, Feng X, Zhang F, Yang Y, Li X. Effect of reaction temperature on structure and fluorescence properties of nitrogen-doped carbon dots. Appl Surf Sci. 2016;387:1236–46. doi: 10.1016/j.apsusc.2016.07.048.
- [53] Bhattacharya A, Chatterjee S, Prajapati R, Mukherjee TK.
 Size-dependent penetration of carbon dots inside the ferritin

- nanocages: evidence for the quantum confinement effect in carbon dots. Phys Chem Chem Phys. 2015;17(19):12833-40. doi: 10.1039/C5CP00543D.
- [54] Zuo P, Lu X, Sun Z, Guo Y, He H. A review on syntheses, properties, characterization and bioanalytical applications of fluorescent carbon dots. Microchim Acta. 2016;183:519–42. doi: 10.1007/s00604-015-1705-3.
- [55] Qian Z, Ma J, Shan X, Feng H, Shao L, Chen J. Highly luminescent N-doped carbon quantum dots as an effective multifunctional fluorescence sensing platform. Chem. 2014;20(8):2254-63. PMID: 24449509. doi: 10.1002/chem.201304374.
- [56] Shi L, Li Y, Li X, Zhao B, Wen X, Zhang G, et al. Controllable synthesis of green and blue fluorescent carbon nanodots for pH and Cu⁽²⁺⁾ sensing in living cells. Biosens Bioelectron. 2016;77:598–602. doi: 10.1016/ j.bios.2015.10.031.
- [57] Choudhury SD, Chethodil JM, Gharat PM, Praseetha PK, Pal H. pH-Elicited luminescence functionalities of carbon dots: mechanistic insights. J Phys Chem Lett. 2017;8(7):1389-95. doi: 10.1021/acs.jpclett.7b00153.
- [58] Jin X, Sun X, Chen G, Ding L, Li Y, Liu Z, et al. pH sensitive carbon dots for the visualization of regulation of intracellular pH inside living pathogenic fungal cells. Carbon. 2015;81:388-95. doi: 10.1016/j.carbon.2014.09.071.
- [59] Wang C, Xu Z, Cheng H, Lin H, Humphrey MG, Zhang C. A hydrothermal route to water-stable luminescent carbon dots as nanosensors for pH and temperature. Carbon. 2015;82:87–95. doi: 10.1016/j.carbon.2014.10.035.
- [60] Jia X, Li J, Wang E. One-pot green synthesis of optically pH-sensitive carbon dots with upconversion luminescence. Nanoscale. 2012;4(18):5572-5. doi: 10.1039/C2NR31319G.
- [61] Zen YW, Ma DK, Wang W, Chen JJ, Zhou L, Zheng YZ, et al. N, S co-doped carbon dots with orange luminescence synthesized through polymerization and carbonization reaction of amino acids. Appl Surf Sci. 2015;342:136–43. doi: 10.1016/ j.apsusc.2015.03.029.
- [62] Su W, Wu H, Xu H, Yang Z, Li Y, Li X, et al. Carbon dots: a booming material for biomedical applications. Mater Chem Front. 2020;4(3):821–36. doi: 10.1039/C9QM00658C.
- [63] Pei S, Zhang J, Gao M, Wu D, Yang Y, Liu R. A facile hydrothermal approach towards photoluminescent carbon dots from amino acids. J Colloid Interface Sci. 2015;439:129–33. doi: 10.1016/j.jcis.2014.10.030.
- [64] Liu ML, Chen BB, Liu ZX, Huang CZ. Highly selective and sensitive detection of 2,4,6-trinitrophenol by using newly developed blue-green photoluminescent carbon nanodots. Talanta. 2016;161:875–80. doi: 10.1016/ j.talanta.2016.08.046.
- [65] Wen X, Shi L, Wen G, Li Y, Dong C, Yang J, et al. Green and facile synthesis of nitrogen-doped carbon nanodots for multicolor cellular imaging and Co²⁺ sensing in living cells. Sens Actuators B Chem. 2016;235:179–87. doi: 10.1016/ j.snb.2016.05.066.
- [66] Wang B, Tang W, Lu H, Huang Z. Hydrothermal synthesis of ionic liquid-capped carbon quantum dots with high thermal stability and anion responsiveness. J Mater Sci. 2015;50:5411–8. doi: 10.1007/s10853-015-9085-y.
- [67] Zhuo Y, Miao H, Zhong D, Zhu S, Yang X. One-step synthesis of high quantum yield and excitation-independent emission

- carbon dots for cell imaging. Mater Lett. 2015;139:197-200. doi: 10.1016/j.matlet.2014.10.048.
- [68] Cui Y, Hu Z, Zhang C, Liu X. Simultaneously enhancing upconversion fluorescence and red-shifting down-conversion luminescence of carbon dots by a simple hydrothermal process. J Mater Chem C. 2014;2(40):6947–52. doi: 10.1039/ C4TB01085J.
- [69] Li H, He X, Liu Y, Huang H, Lian S, Lee S, et al. One-step ultrasonic synthesis of water-soluble carbon nanoparticles with excellent photoluminescent properties. Carbon. 2011;49(2):605–9. doi: 10.1016/j.carbon.2010.10.004.
- [70] Li H, He X, Liu Y, Yu H, Kang Z, Lee ST. Synthesis of fluorescent carbon nanoparticles directly from active carbon via a one-step ultrasonic treatment. Mater Res Bull. 2011;46(1):147–51. doi: 10.1016/j.materresbull.2010.10.013.
- [71] Vithalani R, Patal D, Modi CK, Suthar DH. Review: glowing photoluminescene in carbon-based nanodots: current state and future perspectives. J Mater Sci. 2020;55:8769–92. doi: 10.1007/s10853-020-04671-x.
- [72] Chen J, Liu J, Li J, Xu L, Qiao Y. One-pot synthesis of nitrogen and sulfur codoped carbon dots and its application for sensor and multicolor cellular imaging. J Colloid Interface Sci. 2017;485:167–74. doi: 10.1016/j.jcis.2016.09.040.
- [73] Wu ZL, Zhang P, Gao MX, Liu CF, Wang W, Leng F, et al. One-pot hydrothermal synthesis of highly luminescent nitrogen-doped amphoteric carbon dots for bioimaging from Bombyx mori silk natural proteins. J Mater Chem B. 2013;1(22):2868–73. doi: 10.1039/C3TB20418A.
- [74] Zhou J, Lin P, Ma J, Shan X, Feng H, Chen C, et al. Facile synthesis of halogenated carbon quantum dots as an important intermediate for surface modification. RSC Adv. 2013;3(25):9625–8. doi: 10.1039/C3RA41243A.
- [75] Fang Y, Guo S, Li D, Zhu C, Ren W, Dong S, et al. Easy synthesis and imaging applications of cross-linked green fluorescent hollow carbon nanoparticles. ACS Nano. 2012;6(1):400–9. doi: 10.1021/nn2046373.
- [76] Zhang YQ, Ma DK, Zhuang Y, Zhang X, Chen W, Hong LL, et al. One-pot synthesis of N-doped carbon dots with tunable luminescence properties. J Mater Chem. 2012;22(33):16714-8. doi: 10.1039/C2JM32973E.
- [77] Sun Y, Wang X, Wang C, Tong D, Wu Q, Jiang K, et al. Red emitting and highly stable carbon dots with dual response to pH values and ferric ions. Microchim Acta. 2018;185:83. doi: 10.1007/s00604-017-2544-1.
- [78] Wang Z, Yuan F, Li X, Li Y, Zhong H, Fan L, et al. 53% efficient red emissive carbon quantum dots for high color rendering and stable warm white-light-emitting diodes. Adv Mater. 2017;29(37):1702910. doi: 10.1002/adma.201702910.
- [79] Yuan F, Yuan T, Sui L, Wang Z, Xi Z, Li Y, et al. Engineering triangular carbon quantum dots with unprecedented narrow band width emission for multicolored LEDs. Nat Commun. 2018;9:1–11. doi: 10.1038/s41467-018-04635-5. Article no: 2249.
- [80] Yang ST, Cao L, Luo PG, Lu F, Wang X, Wang H, et al. Carbon dots for optical imaging in vivo. J Am Chem Soc. 2009;131(32):11308-9. doi: 10.1021/ja904843x.
- [81] Anand A, Unnikrishnan B, Wei SC, Chou CP, Zhang LZ, Huang CC. Graphene oxide and carbon dots as broad-spectrum antimicrobial agents a minireview. Nanoscale Horiz. 2019;4(1):117–37. doi: 10.1039/C8NH00174J.

- [82] Zhang H, Chen Y, Liang M, Xu L, Qi S, Chen H, et al. Solid-phase synthesis of highly fluorescent nitrogendoped carbon dots for sensitive and selective probing ferric ions in living cells. Anal Chem. 2014;86(19):9846-52. doi: 10.1021/ac502446m.
- [83] Sahu S, Behera B, Maiti T, Mohapatra S. Simple one-step synthesis of highly luminescent carbon dots from orange juice: application as excellent bio-imaging agents. Chem Commun. 2012;48(70):8835-7. doi: 10.1039/C2CC33796G.
- [84] Zhou M, Zhou Z, Gong A, Zhang Y, Li Q. Synthesis of highly photoluminescentcarbon dots via citric acid and tris for iron (III) ions sensors and bioimaging. Talanta. 2015;143:107–13. PMID: 26078136. doi: 10.1016/ j.talanta.2015.04.015.
- [85] Wang Y, Anilkumar P, Cao L, Liu JH, Luo PG, Tackett KN, et al. Carbon dots of different composition and surface functionalization: cytotoxicity issues relevant to fluorescence cell imaging. Exp Biol Med. 2011;236(11):1231–8. doi: 10.1258/ ebm.2011.011132.
- [86] Wang Y, Bao L, Liu Z, Pang DW. Aptamer biosensor based on fluorescence resonance energy transfer from upconverting phosphors to carbon nanoparticles for thrombin detection in human plasma. Anal Chem. 2011;83(11):8130–7. doi: 10.1021/ac201631b.
- [87] Yang ST, Wang X, Wang H, Lu F, Luo PG, Cao L, et al. Carbon dots as nontoxic and high-performance fluorescence imaging agents. J Phys Chem C. 2009;113(42):18110–4. doi: 10.1021/ jp9085969.
- [88] Havrdova M, Hola K, Skopalik J, Tomankova K, Petr M, Cepe K, et al. Toxicity of carbon dots – effect of surface functionalization on the cell viability, reactive oxygen species generation and cell cycle. Carbon. 2016;99:238–48. doi: 10.1016/j.carbon.2015.12.027.
- [89] Hsu PC, Chen PC, Ou CM, Chang HY, Chang HT. Extremely high inhibition activity of photoluminescent carbon nanodots toward cancer cells. J Mater Chem B. 2013;1:1774–81. doi: 10.1039/C3TB00545C.
- [90] Li CL, Ou CM, Huang CC, Wu WC, Chen YP, Lin TE, et al. Carbon dots prepared from ginger exhibiting efficient inhibition of human hepatocellular carcinoma cells. J Mater Chem B. 2014;2(28):4564-71. doi: 10.1039/C4TB00216D.
- [91] Wang ZG, Zhou R, Jiang D, Song JE, Xu Q, Si J, et al. Toxicity of graphene quantum dots in zebrafish embryo. Biomed Environ Sci. 2015;28(5):341. PMID: 26055561. doi: 10.3967/ bes2015.048.
- [92] Borisova T, Dekaliuk M, Pozdnyakova N, Pastukhov A, Dudarenko M, Borysov A, et al. Harmful impact on presynaptic glutamate and GABA transport by carbon dots synthesized from sulfur-containing carbohydrate precursor. Environ Sci Pollut Res. 2017;24(21):17688-700. PMID: 28601995. doi: 10.1007/s11356-017-9414-6.
- [93] Li H, Liu R, Lian S, Liu Y, Huang H, Kang Z. Near-infrared light controlled photocatalytic activity of carbon quantum dots for highly selective oxidation reaction. Nanoscale. 2013;5(8):3289-97. doi: 10.1039/C3NR00092C.
- [94] Comparelli R, Fanizza E, Curri ML, Cozzoli PD, Mascolo G, Agostiano A. UV-induced photocatalytic degradation of azo dyes by organic-capped ZnO nanocrystals immobilized onto substrates. Appl Catal B. 2005;60(1-2):1-11. doi: 10.1016/ j.apcatb.2005.02.013.

- [95] Zhang H, Huang H, Ming H, Li H, Zhang L, Liu Y, et al. Carbon quantum dots/Ag₃PO₄ complex photocatalysts with enhanced photocatalytic activity and stability under visible light. J Mater Chem. 2012;22(21):10501–6. doi: 10.1039/ C2JM30703K.
- [96] Lai CW, Hsiao YH, Peng YK, Chou PT. Facile synthesis of highly emissive carbon dots from pyrolysis of glycerol; gram scale production of carbon dots/mSiO₂ for cell imaging and drug release. J Mater Chem. 2012;22(29):14403–9. doi: 10.1039/C2JM32206D.
- [97] Li H, He X, Kang Z, Huang H, Liu Y, Liu J, et al. Water-soluble fluorescent carbon quantum dots and photocatalyst design. Angew Chem Int Ed. 2010;49(26):4430-4. doi: 10.1002/ anie.200906154.
- [98] Ma Z, Zhang YL, Wang L, Ming H, Li H, Zhang X, et al. Bioinspired photoelectric conversion system based on carbon-quantum-dot-doped dye-semiconductor complex. ACS Appl Mater Interfaces. 2013;5(11):5080-4. doi: 10.1021/ am400930h.
- [99] Ma Z, Ming H, Huang H, Liu Y, Kang Z. One-step ultrasonic synthesis of fluorescent N-doped carbon dots from glucose and their visible-light sensitive photocatalytic ability. New J Chem. 2012;36(4):861–4. doi: 10.1039/C2NJ20942J.
- [100] Liu XJ, Guo ML, Huang J, Yin XY. Improved fluorescence of carbon dots prepared from bagasse under alkaline hydrothermal conditions. Bio Res. 2013;8(2):2537-46. doi: 10.15376/biores.8.2.2537-2546.
- [101] Mewada A, Pandey S, Shinde S, Mishra N, Oza G, Thakur M, et al. Green synthesis of biocompatible carbon dots using aqueous extract of *Trapa bispinosa* peel. Mater Sci Eng C. 2013;33(5):2914–7. doi: 10.1016/j.msec.2013.03.018.
- [102] Pandey S, Mewada A, Oza G, Thakur M, Mishra N, Sharon M, et al. Synthesis and centrifugal separation of fluorescent carbon dots at room temperature. Nanosci Nanotechnol Lett. 2013;5(7):775–9. doi: 10.1166/nnl.2013.1617.
- [103] Hsu P-C, Shih ZY, Lee C-H, Chang H-T. Synthesis and analytical applications of photoluminescent carbon nanodots. Green Chem. 2012;14(4):917–20. doi: 10.1039/C2GC16451E.
- [104] Lu W, Qin X, Liu S, Chang G, Zhang Y, Luo Y, et al. Economical, green synthesis of fluorescent carbon nanoparticles and their use as probes for sensitive and selective detection of mercury(II) ions. Anal Chem. 2012;84(12):5351–7. doi: 10.1021/ac3007939.
- [105] Dong Y, Wang R, Li G, Chen C, Chi Y, Chen G. Polyamine-functionalized carbon quantum dots as fluorescent probes for selective and sensitive detection of copper ions. Anal Chem. 2012;84(14):6220-4. doi: 10.1021/ac3012126.
- [106] Dong Y, Wang R, Li H, Shao J, Chi Y, Lin X, et al. Polyamine-functionalized carbon quantum dots for chemical sensing. Carbon. 2012;50(8):2810-5. doi: 10.1016/j.carbon.2012.02.046.
- [107] Zhu A, Qu Q, Shao X, Kong B, Tian Y. Carbon-dot-based dualemission nanohybrid produces a ratiometric fluorescent sensor for *in vivo* imaging of cellular copper ions. Angew Chem Int Ed. 2012;51(29):7185–9. doi: 10.1002/ anie.201109089.
- [108] Vedamalai M, Periasamy AP, Wang CW, Tseng YT, Ho LC, Shih CC, et al. Carbon nanodots prepared from o-phenylenediamine for sensing of Cu⁽²⁺⁾ ions in cells. Nanoscale. 2014;6(21):13119-25. doi: 10.1039/C4NR03213F.

- [109] Wang CI, Periasamy AP, Chang HT. Photoluminescent C-dots@RGO probe for sensitive and selective detection of acetylcholine. Anal Chem. 2013;85(6):3263-70. doi: 10.1021/ac303613d.
- [110] Xia C, Hai X, Chen S, Chen X, Wang JH. Chin J Anal Chem. 2016;44(1):41–8.
- [111] Shi Y, Pan Y, Zhang H, Zhang Z, Li MJ, Yi C, et al. A dual-mode nanosensor based on carbon quantum dots and gold nanoparticles for discriminative detection of glutathione in human plasma. Biosens Bioelectron. 2014;56:39–45. doi: 10.1016/j.bios.2013.12.038.
- [112] Wu L, Wang J, Ren J, Li W, Qu X. Highly sensitive electrochemiluminescent cytosensing using carbon nanodot @Ag hybrid material and graphene for dual signal amplification. Chem Commun (Cambridge). 2013;49:5675–7. doi: 10.1039/ C3CC42637H.
- [113] Zhang L, Zhang ZY, Liang RP, Li YH, Qiu JD. Boron-doped graphene quantum dots for selective glucose sensing based on the "abnormal" aggregation-induced photoluminescence enhancement. Anal Chem. 2014;86(9):4423-30. doi: 10.1021/ac500289c.
- [114] Wei XM, Xu Y, Li YH, Yin XB, He XW. Ultrafast synthesis of nitrogen-doped carbon dots via neutralization heat for bioimaging and sensing applications. RSC Adv. 2014;4(84):44504–8. doi: 10.1039/C4RA08523J.
- [115] Lin Z, Xue W, Chen H, Lin JM. Peroxynitrous-acid-induced chemiluminescence of fluorescent carbon dots for nitrite sensing. Anal Chem. 2011;83(21):8245-51. doi: 10.1021/ ac202039h.
- [116] Shao X, Gu H, Wang Z, Chai X, Tian Y, Shi G. Highly selective electrochemical strategy for monitoring of cerebral Cu²⁺ based on a carbon Dot-TPEA hybridized surface. Anal Chem. 2013;85(1):418–25. doi: 10.1021/ac303113n.
- [117] Li LL, Ji J, Fei R, Wang CZ, Lu Q, Zhang JR, et al. A facile microwave avenue to electrochemiluminescent two-color graphene quantum dots. Adv Funct Mater. 2012;22(14):2971–9. doi: 10.1002/adfm.201200166.
- [118] Song Y, Shi W, Chen W, Li X, Ma H. Fluorescent carbon nanodots conjugated with folic acid for distinguishing folate-receptor-positive cancer cells from normal cells. J Mater Chem. 2012;22(25):12568-73. doi: 10.1039/ C2JM31582C.
- [119] Liu C, Zhang P, Zhai X, Tian F, Li W, Yang J, et al. Nano-carrier for gene delivery and bioimaging based on carbon dots with PEI-passivation enhanced fluorescence. Bio Mater. 2012;33(13):3604–13. doi: 10.1016/ j.biomaterials.2012.01.052.
- [120] Lai CW, Hsiao YH, Peng YK, Chou PT. Facile synthesis of highly emissive carbon dots from pyrolysis of glycerol; gram scale production of carbon dots/mSiO₂ for cell imaging and drug release. J Mater Chem. 2012;22(29):14403–9. doi: 10.1039/C2JM32206D.
- [121] Chowdhuri AR, Singh T, Ghosh SK, Sahu SK. Carbon dots embedded magnetic nanoparticles @chitosan @metal organic framework as a nanoprobe for pH sensitive targeted anticancer drug delivery. ACS Appl Mater Interfaces. 2016;8(26):16573–83. doi: 10.1021/acsami.6b03988.
- [122] Wang H, Di J, Sun Y, Fu J, Wei Z, Matsui H, et al. Biocompatible PEG-chitosan@carbon dots hybrid nanogels for two-photon fluorescence imaging, near infrared light/pH dual-

- responsive drug carrier, and synergistic therapy. Adv Funct Mater. 2015;25(34):5537-47. doi: 10.1002/adfm.201501524.
- [123] Choi Y, Kim S, Choi MH, Ryoo SR, Park J, Min DH, et al. Highly biocompatible carbon nanodots for simultaneous bioimaging and targeted photodynamic therapy *in vitro* and *in vivo*. Adv Funct Mater. 2014;24(37):5781–9. doi: 10.1002/adfm.201400961.
- [124] Luo PG, Sahu S, Yang ST, Sonkar SK, Wang J, Wang H, et al. Carbon "quantum" dots for optical bioimaging. J Mater Chem B. 2013;1(16):2116-27. doi: 10.1039/C3TB00018D.
- [125] Ding C, Zhu A, Tian Y. Functional surface engineering of C-dots for fluorescent biosensing and *in vivo* bioimaging. Acc Chem Res. 2014;47(1):20-30. doi: 10.1021/ar400023s.
- [126] Xu Y, Wu M, Liu Y, Feng XZ, Yin XB, He XW, et al. Nitrogen-doped carbon dots: a facile and general preparation method, photoluminescence investigation, and imaging applications. Chem Eur J. 2013;19(7):2276-83. doi: 10.1002/chem.201203641.
- [127] Chandra S, Das P, Bag S, Laha D, Pramanik P. Synthesis, functionalization and bioimaging applications of highly fluorescent carbon nanoparticles. Nanoscale. 2011;3(4):1533-40. PMID: 21293809. doi: 10.1039/ c0nr00735h.
- [128] Chong Y, Ma Y, Shen H, Tu X, Zhou X, Xu J, et al. The *in vitro* and *in vivo* toxicity of graphene quantum dots. Biomaterials. 2014;35(19):5041–8. PMID: 24685264. doi: 10.1016/j.biomaterials.2014.03.021.
- [129] Bhunia SK, Saha A, Maity AR, Ray SC, Jana NR. Carbon nanoparticle-based fluorescent bioimaging probes. Sci Rep. 2013;3:1473. PMID: 23502324, PMCID: PMC3600594. doi: 10.1038/srep01473.
- [130] Shang W, Zhang X, Zhang M, Fan Z, Sun Y, Han M, et al. The uptake mechanism and biocompatibility of graphene quantum dots with human neural stem cells. Nanoscale. 2014;6(11):5799–806. PMID: 24740121. doi: 10.1039/c3nr06433f.
- [131] Wang L, Wang Y, Xu T, Liao H, Yao C, Liu Y, et al. Gram-scale synthesis of single-crystalline graphene quantum dots with superior optical properties. Nat Commun. 2014;28(5):5357. doi: 10.1038/ncomms6357.
- [132] Zhang X, Wang S, Xu L, Feng L, Ji Y, Tao L, et al. Biocompatible polydopamine fluorescent organic nanoparticles: facile preparation and cell imaging. Nanoscale. 2012;4(18):5581–4. PMID: 25348348. doi: 10.1038/ncomms6357.
- [133] Tang J, Kong B, Wu H, Xu M, Wang Y, Wang Y, et al. Carbon nanodots featuring efficient FRET for real-time monitoring of drug delivery and two-photon imaging. Adv Mater. 2013;25(45):6569-74. doi: 10.1002/adma.201303124.
- [134] Kang YF, Fang YW, Li YH, Li W, Yin XB. Nucleus-staining with biomolecule-mimicking nitrogen-doped carbon dots prepared by a fast neutralization heat strategy. Chem Commun. 2015;51(95):16956-9. doi: 10.1039/C5CC06304C.
- Zhang X, Wang H, Wang H, Zhang Q, Xie J, Tian Y, et al. Single-layered graphitic-C₍₃₎N₍₄₎ quantum dots for two-photon fluorescence imaging of cellular nucleus. Adv Mater. 2014;26(26):4438–43. doi: 10.1002/adma.201400111.
- 136] Kong B, Zhu A, Ding C, Zhao X, Li B, Tian Y. Carbon dot-based inorganic-organic nanosystem for two-photon imaging and biosensing of pH variation in living cells and tissues. Adv Mater. 2012;24(43):5844-8. doi: 10.1002/adma.201202599.

- [137] Lieschke GJ, Currie PD. Animal models of human disease: zebrafish swim into view. Nat Rev Genet. 2007;8(5):353-67. PMID: 17440532. doi: 10.1038/nrg2091.
- [138] Kang YF, Li YH, Fang YW, Xu Y, Wei XM, Yin XB. Carbon quantum dots for zebafish fluoresence imaging. Sci Rep. 2015;5:11835. doi: 10.1038/srep11835.
- [139] Xu Y, Li YH, Wang Y, Cui JL, Yin XB, He XW, et al. ¹³C-engineered carbon quantum dots for *in vivo* magnetic resonance and fluorescence dual-response. Analyst. 2014;139(20):5134–9. doi: 10.1039/C4AN01194E.
- [140] Tao H, Yang K, Ma Z, Wan J, Zhang Y, Kang Z, et al. *In vivo* NIR fluorescence imaging, biodistribution, and toxicology of photoluminescent carbon dots produced from carbon nanotubes and graphite. Small. 2012;8(2):281–90. PMID: 22095931. doi: 10.1002/smll.201101706.
- [141] Li N, Liang X, Wang L, Li ZH, Li P, Zhu Y, et al. Biodistribution study of carbogenic dots in cells and *in vivo* for optical imaging. J Nanopart Res. 2012;14(10):1177. doi: 10.1007/ s11051-012-1177-x.
- [142] Zu F, Yan F, Bai Z, Xu J, Wang Y, Huang Y, et al. The quenching of the fluorescence of carbon dots: a review on mechanisms and applications. Microchim Acta. 2017;184(5):1899–914. doi: 10.1007/s00604-017-2318-9.
- [143] Kundu A, Nandi S, Das P, Nandi AK. Facile and green approach to prepare fluorescent carbon dots: emergent nanomaterial for cell imaging and detection of vitamin B₂. J Colloid Interface Sci. 2016;468:276-83. doi: 10.1016/ j.jcis.2016.01.070.
- [144] Barati A, Shamsipur M, Abdollahi H. Hemoglobin detection using carbon dots as a fluorescence probe. Biosens. Bioelectron. 2015;71:470-5. PMID: 25988918. doi: 10.1016/ j.bios.2015.04.073.
- [145] Lim SY, Shen W, Gao Z. Carbon quantum dots and their applications. Chem Soc Rev. 2015;44(1):362-81. doi: 10.1039/C4CS00269E.
- [146] Jiang K, Sun S, Zhang L, Lu Y, Wu A, Cai C, et al. Red, green, and blue luminescence by carbon dots: full-color emission tuning and multicolor cellular imaging. Angew Chem Int Ed. 2015;54(18):5360-3. doi: 10.1002/anie.201501193.
- [147] Ge J, Lan M, Zhou B, Liu W, Guo L, Wang H, et al. A graphene quantum dot photodynamic therapy agent with high singlet oxygen generation. Nat Commun. 2014;5:4596. doi: 10.1038/ ncomms5596.
- [148] Bourlinos AB, Bakandritsos A, Kouloumpis A, Gournis D, Krysmann M, Giannelis EP, et al. Gd(iii)-doped carbon dots as a dual fluorescent-MRI probe. J Mater Chem. 2012;22(44):23327–30. doi: 10.1039/C2JM35592B.
- [149] Xu Y, Jia XH, Yin XB, He XW, Zhang YK. Carbon quantum dot stabilized gadolinium nanoprobe prepared via a one-Pot hydrothermal approach for magnetic resonance and fluorescence dual-modality bioimaging. Anal Chem. 2014;86(24):12122-9. doi: 10.1021/ac503002c.
- [150] Feng J, Wang WJ, Hai X, Yu YL, Wang JH. Green preparation of nitrogen-doped carbon dots derived from silkworm chrysalis for cell imaging. J Mater Chem B. 2016;4(3):387–93. doi: 10.1039/C5TB01999K.
- [151] Ding H, Wei JS, Zhong N, Gao QY, Xiong HM. Highly efficient red-emitting carbon dots with gram-scale yield for bioimaging. Langmuir. 2017;33(44):12635-42. doi: 10.1021/ acs.langmuir.7b02385.

- [152] Yan F, Jiang Y, Sun X, Bai Z, Zhang Y, Zhou X. Surface modification and chemical functionalization of carbon dots: a review. Microchim Acta. 2018;185:424. doi: 10.1007/s00604-018-2953-9.
- [153] Zhou L, Lin Y, Huang Z, Ren J, Qu X. Carbon nanodots as fluorescence probes for rapid, sensitive, and label-free detection of Hg²⁺ and biothiols in complex matrices. Chem Commun. 2012;48:1147–9. doi: 10.1039/C2CC16791C.
- [154] Wen X, Shi L, Wen G, Li Y, Dong C, Yang J, et al. Green synthesis of carbon nanodots from cotton for multicolor imaging, patterning, and sensing. Sens Actuators B Chem. 2015;221:769–76. doi: 10.1016/j.snb.2015.07.019.
- [155] Das P, Ganguly S, Bose M, Mondal S, Das AK, Banerjee S, et al. A simplistic approach to green future with eco-friendly luminescent carbon dots and their application to fluorescent nano-sensor 'turn-off' probe for selective sensing of copper ions. Mater Sci Eng C. 2017;75(1):1456–64. doi: 10.1016/j.msec.2017.03.045.
- [156] Fan H, Zhang M, Bhandari B, Yang CH. Food waste as a carbon source in carbon quantum dots technology and their applications in food safety detection. Trends Food Sci Tech. 2020;95:86–96. doi: 10.1016/j.tifs.2019.11.008.
- [157] Huang CC, Hung YS, Weng YM, Chen W, Lai YS. Review: sustainable development of carbon nanodots technology: natural products as a carbon source and applications to food safety. Trends Food Sci Technol. 2019;86:144–52. doi: 10.1016/j.tifs.2019.02.016.
- [158] Himaja AL, Karthik PS, Singh SP. Carbon dots: the newest member of the carbon nanomaterials family. Chem Rec. 2015;15(3):595-615. doi: 10.1002/tcr.201402090.
- [159] Tuerhong M, Xu Y, Yin XB. Review on carbon dots and their applications. Chin J Anal Chem. 2007;45(1):139-50. doi: 10.1016/S1872-2040(16)60990-8.
- [160] Varma RS. Journey on greener pathways: from the use of alternate energy inputs and benign reaction media to sustainable applications of nano-catalysts in synthesis and environmental remediation. Green Chem. 2014;16:2027–41. doi: 10.1039/C3GC42640H.
- [161] Tyagi A. From lemon peel waste: applications in sensing and photocatalysis. RSC Adv. 2016;6:72423-32. doi: 10.1039/ C6RA10488F.
- [162] Bano D, Kumar V, Singha VK, Hasan SH. Green synthesis of fluorescent carbon quantum dots for the detection of mercury (II) and glutathione. New J Chem. 2018;42:5814–21. doi: 10.1039/C8NJ00432C.
- [163] de Yro PAN, Quaichon GMO, Cruz RAT, Emolaga CS, Que MCO, Magdaluyo Jr ER, et al. Hydrothermal synthesis of carbon quantum dots from biowaste for bio-imaging. AIP Conf Proc. 2019;2083(1):1–6. doi: 10.1063/1.5094310.
- [164] Liu X, Hao J, Liu J, Tao H. Green synthesis of carbon quantum dots from lignite coal and the application in Fe³⁺ detection. IOP Conf Series Earth Environ Sci. 2018;113:012063. doi: 10.1088/1755-1315/113/1/012063.
- [165] Ramezani Z, Qorbanpour M, Rahbar N. Green synthesis of carbon quantum dots using quince fruit (Cydonia oblonga) powder as carbon precursor: application in cell imaging and As³⁺ determination. Colloids Surf A. 2018;549:58–66. doi: 10.1016/j.colsu rfa.2018.04.006.
- [166] Tadessea A, Devi DR, Hagosa M, Battud G, Basavaiah K. Facile green synthesis of fluorescent carbon quantum dots

- from citrus lemon juice for live cell imaging. Asian J Nanosci Mater. 2018;1:36–46. doi: 10.26655/ajnan omat.2018.1.5.
- [167] Li H, Shao F-Q, Huang H, Feng J-J, Wang A-J. Eco-friendly and rapid microwave synthesis of green fluorescent graphitic carbon nitride quantum dots for vitro bioimaging. Sens Actuators B Chem. 2016;226:506–11. doi: 1016/ i.snb.2015.12.018.
- [168] Edison TN, Atchudan R, Shim JJ, Kalimuthu S, Ahn BC, Lee YR. Turn-off fluorescence sensor for the detection of ferric ion in water using green synthesized N-doped carbon dots and its bio-imaging. J Photochem Photobiol B. 2016;158:235–42. doi: 10.1016/j.jphot obiol.2016.03.010.
- [169] Edison TN, Atchudan R, Sethuraman MG, Shim JJ, Lee YR. Microwave assisted green synthesis of fluorescent N-doped carbon dots: cytotoxicity and bio-imaging applications. J Photochem Photobiol B. 2016;161:154–61. doi: 10.1016/ j.jphot obiol.2016.05.017.
- [170] Peng J, Gao W, Gupta BK, Liu Z, Aburto RR, Ge L, et al. Graphene quantum dots derived from carbon fibers. Nano Lett. 2012;12:844–9. doi: 10.1021/nl2038979.
- [171] Hola K, Zhang Y, Wang Y, Gianneils EP, Zboril R, Rogach AL. Carbon dots Emerging light emitters for bioimaging, cancer therapy and optoelectronics. Nano Today. 2014;9:590–603. doi: 10.1016/j.nantod.2014.09.004.
- [172] Liu ML, Chen BB, Li CM, Huang CZ. Carbon dots: synthesis, formation mechanism, fluorescence origin and sensing applications. Green Chem. 2019;21:449-71. doi: 10.1039/ C8GC02736F.
- [173] Arts R, Aper SJA, Merkx M. Engineering BRET-sensor proteins. Methods Enzymol. 2017;589:87-114. doi: 10.1016/ bs.mie.2017.01.010.
- [174] Rizzo MA, Springer GH, Granada B, Piston DW. An improved cyan fluorescent protein variant useful for FRET. Nat Biotechnol. 2004;22:445–9. doi: 10.1038/nbt945.
- [175] Chen L, Tse WH, Chen Y, Mcdonald MW, Melling J, Zhang J. Nanostructured biosensor for detecting glucose in tear by applying fluorescence resonance energy transfer quenching mechanism. Biosens Bioelectron. 2017;91:393–9. doi: 10.1016/j.bios.2016.12.044.
- [176] De A, Loening AM, Gambhir SS. An improved bioluminescence resonance energy transfer strategy for imagingintra-

- cellular events in single cells and living subjects. Cancer Res. 2007;67:7175–83. doi: 10.1158/0008-5472.
- [177] Fereja TH, Hymete A, Gunasekaran T. A recent review on chemiluminescence reaction, principle and application on pharmaceutical analysis. Int Sch Res Not. 2013;2013:1–12. doi: 10.1155/2013/230858. Article ID 230858.
- [178] Baklaushev VP, Kilpeläinen A, Petkov S, Abakumov MA, Grinenko NF, Yusubalieva GM, et al. Luciferase expression allows bioluminescence imaging but imposes limitations on the orthotopic mouse (4T1) model of breast cancer. Sci Rep. 2017;7:7715. doi: 10.1038/s41598-017-07851-z.
- [179] Hwang E, Song J, Zhang J. Integration of nanomaterials and bioluminescence resonance energy transfer techniques for sensing biomolecules. Biosensors. 2019;9(1)42:1–16. doi: 10.3390/bios901004258.
- [180] Bhaumik S, Gambhir SS. Optical imaging of Renilla luciferase reporter gene expression in living mice. Proc Natl Acad Sci U S A. 2002;99(1):377–82. PMID: 11752410, PMCID: PMC117568. doi: 10.1073/pnas.012611099.
- [181] Chen L, Bao Y, Denstedt J, Zhang J. Nanostructured bioluminescent sensor for rapidly detecting thrombin. Biosens Bioelectron. 2016;77:83–9. doi: 10.1016/j.bios.2015.09.010.
- [182] Kamkaew A, Sun H, England CG, Cheng L, Liu Z, Cai W. Quantum dot-NanoLuc bioluminescence resonance energy transfer enables tumor imaging and lymph node mapping in vivo. Chem Commun Camb Engl. 2016;52:6997–7000. doi: 10.1039/C6CC02764D.
- [183] Hsu CY, Chen CW, Yu HP, Lin YF, Lai PS. Bioluminescence resonance energy transfer using luciferase-immobilized quantum dots for self-illuminated photodynamic therapy. Biomaterials. 2012;34(4):1204–12. doi: 10.1016/j.biomaterials.2012.08.044.
- [184] So M-K, Loening AM, Gambhir SS, Rao J. Creating self-illuminating quantum dot conjugates. Nat Protoc. 2006;1:1160-4. doi: 10.1038/nprot.2006.162.
- [185] Iravani S, Varma RS. Green synthesis, biomedical and biotechnological applications of carbon and graphene quantum dots. A review. Environ Chem Lett. 2020;18:703–27. doi: 10.1007/s10311-020-00984-0.
- [186] Song J, Zhang J. Self-illumination of carbon dots by bioluminescence resonance energy transfer. Sci Rep. 2019;9:13796. doi: 10.1038/s41598-019-50242-9.