Impacts of cold exposure on energy metabolism

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Abstract

Cold stimulation has been shown to regulate glucose, lipid, and amino acid metabolism, while also increasing heat production and energy expenditure in the body. Disordered energy metabolism is a key factor in the onset and progression of chronic metabolic conditiones such as diabetes, obesity, and cardiovascular disease. Recent research has unveiled the myriad pathways through which cold stimulation affects human energy metabolism. This article provides an overview of how cold stimulation affects energy metabolism across the three major metabolic pathways. Furthermore, it explores the implications and potential therapeutic applications of cold stimulation in the prevention and treatment of various metabolic diseases.

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Keywords

cold stimulation; energy metabolism; glucose metabolism; lipid metabolism; amino acid metabolism; chronic metabolic diseases

Received 22 December 2023, accepted 31 January 2024

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1 Introduction

Climate change has led to extreme environmental temperatures, which are strongly associated with increased mortality rates. The Global Burden of Disease Study recently identified non-optimal temperatures as one of the leading risk factors for global mortality^[1]. Interestingly, low temperatures have been associated with a more pronounced increase in mortality risk compared to high temperatures^[2]. Approximately 30 countries worldwide are situated in cold northern regions, cand over half of China's territory qualifies as cold cities, defined by an average temperature below 0°C. Heilongjiang Province stands as a representative region of cold areas in China, where the prevalence of chronic diseases characterized by metabolic disorders is notably high. Addressing the disease burden associated with cold stimulation-related chronic conditions is a critical challenge in current clinical research.

Environmental conditions can significantly impact energy metabolism^[3]. Studies indicate that cold stimulation exerts a notable influence on human energy metabolism. The exploration of cold stimulation's effects on energy metabolism aims to uncover the metabolic regulatory mechanisms of the human body in the cold environment. This endeavor seeks novel approaches for preventing or treating metabolic diseases and provides fresh perspectives and directions for clinical application

in relevant fields. Thus, this paper provides a comprehensive overview of energy metabolism under cold stimulation, focusing on glucose metabolism, lipid metabolism, amino acid metabolism and metabolic diseases, this paper summarizes the characteristics of energy metabolism, and their implications for metabolic diseases based on recent research findings (Fig.1).

2 Definition of cold stimuli

The definition of cold stimuli varies depending on geographical location. Cold stimulation encompasses any form of stimulation that induces cold sensations in human skin and deep tissues. This includes activities such as cold-water bathing, ice application, cold exposure, and other modalities^[4-5]. In animal studies, appropriate temperatures typically range between 22°C and 26°C, with low temperature exposure defined as 4°C^[6-8]. For human studies, the optimal temperature is set between 21°C and 24°C, with cold exposure defined as temperatures ranging from 14°C to 19°C or individualized cooling regimens^[9-12].

3 Cold stimulation and regulation of glucose metabolism

Under normal conditions, approximately 70% of the energy supply required by the human body is derived from glucose metabolism^[13]. Cold stimulation activates thermogenic

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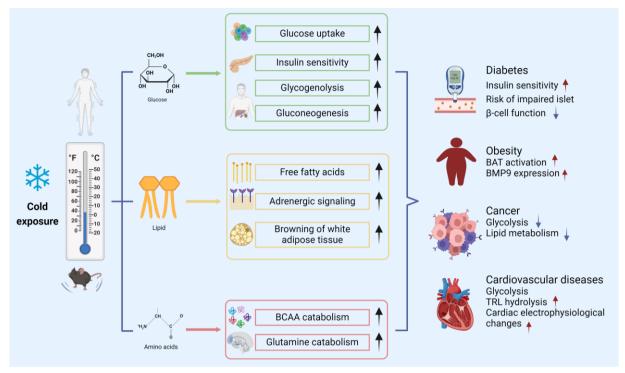


Fig. 1 Cold exposure affects energy metabolism and participates in the occurrence and development of metabolic diseases BCAA, branched chain amino acids; TRL, triglyceride-rich lipoproteins.

metabolism and promotes glucose uptake by cells. Recent studies employing deuterium metabolic imaging (DMI) have shown a higher labeled glucose signal in rats exposed to cold stimulation (9°C for 1 week), indicating increased sugar uptake [14]. The mammalian target of rapamycin (mTOR), an atypical serine/threonine protein kinase, integrates various extracellular signals such as nutrition, energy, and growth factors [15-16]. Loss of rapamycin complex 2 (mTORC2) impairs glucose uptake in brown adipose tissue (BAT) during cold exposure. However, research by Castro $et\ al.$ [17] suggests that rapamycin complex 1 (mTORC1) activity and BAT glucose uptake were upregulated upon cold stimulation (10 \pm 1°C), independent of mTORC2, indicating that mTORC2 deficiency in adipocytes is not necessary for glucose uptake.

Insulin, an anabolic hormone secreted by beta cells, plays a crucial role in carbohydrate metabolism^[18-19]. Scheel *et al.*^[20] proposed that non-shivering thermogenesis (NST) induced by cold stimulation increases energy expenditure and promotes negative energy balance, potentially improving insulin resistance (IR) in skeletal muscle and adipose tissue. T-box1 (TBX1), a transcription factor involved in mesoderm development, has been found to maintain glucose homeostasis in cold conditions (4°C), and its expression is crucial for proper insulin signaling^[21]. These findings suggest the involvement of

insulin signaling in glucose metabolism under cold conditions, although the precise mechanism remain to be fully elucidated. Recently evidence suggests that hypothermia exposure enhances whole-body insulin sensitivity[22], potentially mediated by various proteins, adipokines, and mRNAs related to glucose metabolism. Nascimento et al. [23] demonstrated that coldinduced increases in peripheral insulin sensitivity involve translocation of glucose transporter 4 (GLUT4). Another study found that cold exposure increased insulin sensitivity through AMP-activated protein kinase (AMPK) or peroxisome proliferator-activated receptor delta (PPAR-delta)-dependent signaling^[24]. In addition, cold stimulation (4°C) induces microRNA-485 (miR-485) expression, which promotes the proliferation of BAT progenitor cells, increases BAT mass, enhances NST, and significantly improves insulin sensitivity^[25]. Interestingly, Sellers et al. [26] confirmed the existence of IR after cold exposure-induced shivering in experiments involving healthy human, providing evidence for the impact of glucose metabolism after cold exposure. These findings suggest that shivering and NST may have distinct effects on insulin signaling in the body under cold stimulation.

Regarding the effects of cold stimulation on glycolysis and gluconeogenesis, it was previously believed that cold stimulation increased the secretion of thyroid hormones, thereby accelerating energy metabolism, glycogenolysis and gluconeogenesis^[27]. However, recent cross-sectional studies by Merchan-Ramirez *et al.*^[28] in young healthy adults demonstrated that cold exposure affects thyroid hormone levels independent of glucose metabolism assessed 2 hours after cold exposure. Further investigations into the effects of altered thyroid hormone levels on glucose metabolism during prolonged cold exposure are warranted. Another study showed that cold stimulation (4°C) increased intramuscular conversion of lactate to glycogen (gluconeogenesis) by promoting myofructose-1, 6-bisphosphatase 2 (Fbp2)^[29]. The mechanisms underlying these processes remain unclear, and further studies are needed to elucidate the effects of cold stimulation on the different stages of the glucose metabolic pathway.

4 Cold stimulation and regulation of fat metabolism

The human body primarily obtains and absorbs fatty acids through the ingestion and degradation of triglycerides. These fatty acids are then either directly catabolized to provide energy or re-synthesize into triglycerides for transportation to tissues for storage^[30-31]. A randomized crossover study has shown that cold exposure (9°C) induces an increase in circulating free fatty acid (FFA) levels in humans, suggesting that cold exposure mobilizes lipids to provide nutrients for thermogenesis in organs^[32]. Under periods of energy expenditure, such as cold exposure, triglycerides are released as FFA to support a shift towards catabolism^[33]. Pernes et al.^[34] further observed that triglycerides were the primary lipids affected by cold stimulation, with significant reductions observed at temperatures of 22°C and 5°C. However, studies analyzing serum metabolites in humans have shown a tendency for triglyceride levels to increase during cold exposure[35]. Additionally, Straa et al.[4] confirmed through serum lipid profiling that cold exposure gradually increased circulating FFA, peaking at 60 minutes, while total triacylglycerols (TAG) only transiently decreased at 30 minutes. These findings suggest that the effect of cold stimulation on triglyceride is time-dependent and that cold stimulation affects fat catabolism through other pathways.

The mechanisms triggering changes in lipid metabolism induced by cold stimulation are still poorly understood. $\beta3$ -adrenergic receptors ($\beta3$ -AR), primarily distributed in adipose tissue, play a crucial role in fat metabolism^[36-37]. Zinc-alpha2-glycoprotein (ZAG) is a lipid mobilization factor discovered in recent years that promotes lipolysis^[38-39]. Fan *et al.*^[40] found that cold stress (6°C) promotes lipid metabolism by increasing $\beta3$ -AR expression in the ZAG signaling pathway. Moreover, the cold environment can promote triglyceride degradation by stimulating the production of thyroxine (T4), which binds to $\beta3$ -AR^[41]. Hong

et al. [42] further demonstrated that b-cell translocation gene 2 (BTG2)-deficient mice exhibited reduced fat degradation and lipase expression in response to adrenergic signaling under cold exposure, confirming the essential role of BTG2 in the correct response to β -adrenergic signaling under cold stimulation. These mechanisms are proposed as links between cold stimulation and fat metabolism or as factors that may trigger enhanced fat catabolism.

Cold stimulation promotes lipolysis and metabolism by inducing the browning of white adipose tissue. In mice, this browning process is associated with increased expression of the key thermogenic protein uncoupling protein 1 (UCP1)[43]. The adipokine asprosin reduced the expression of the browning marker UCP1 and other browning-related genes through the transcription factor nuclear factor-E2-related factor 2 (Nrf2), thereby negatively regulating browning and enhancing lipid deposition in adipose tissue^[44-45]. Miao et al. [44] demonstrated that cold stimulation (4°C) significantly downregulated asprosin expression, thereby promoting the browning of white adipose tissue. In addition, fibroblast growth factor 6/9 (FGF6/9) is a potent inducer of UCP1 expression^[46]. A recent study found that FGF6/9 expression is upregulated in response to cold stimulation (5°C), further inducing UCP1 expression [47]. Although potential mechanisms by which cold stimulation affects lipid metabolism through various pathways have been identified, the time dependence of these effects and the specific roles of each mechanism remain to be fully elucidated.

5 Cold stimulation and regulation of amino acid metabolism

Amino acid metabolism plays an important role in maintaining the metabolic balance of the body, involving the conversion of amino acids into sugars, lipids, or the resynthesis of non-essential amino acids. Dysregulation of amino acid metabolism can lead to various related diseases^[48-49]. When amino acid metabolism is dysregulated, it will lead to the occurrence of a variety of related diseases^[50].

Limited research has been conducted on the effects of cold stimulation on amino acid metabolism. However, recent studies focusing on the metabolism of branched-chain amino acids (BCAAs) and glutamine have garnered attention. BCAAs, including valine, leucine, and isoleucine, serve as signaling molecules regulating various signaling pathways *in vivo*^[51]. The dysregulation of BCAA metabolism is implicated in metabolic diseases such as obesity and tumors^[52]. Li *et al.*^[53] showed through KEGG analysis that cold stimulation (4°C) promotes amino acid catabolism, especially BCAA. Teng *et al.*^[54] further found that prolonged cold exposure (7 ± 3°C) promotes BCAA

degradation and inhibits amino acid absorption by inhibiting the expression of amino acid transporters. Similarly, recent findings indicate that cold stimulation mediates BCAA transport by inducing SLC25A44 expression, thereby enhancing BCAA catabolism *in vivo*^[55]. These observations suggest that cold stimulation may regulate amino acid metabolism by affecting amino acid transport. In addition, it is found that the knockout of FAM195A, a gene involved in BCAA metabolism, downregulates the expression of multiple enzymes associated with BCAA metabolism, significantly impairing mice tolerance to cold stimulation. This suggests that FAM195A may not only regulate BCAA metabolism but also serve as a potential target for cold stimulation-induced BCAA metabolic disorders^[56]. Nonetheless, further studies are needed to identify other factors through which cold stimulation affects BCAA metabolism.

Glutamine (Gln) is an abundant free amino acid in human tissues both intracellularly and extracellularly, with physiological functions including improving immune function, antioxidation, and gastrointestinal nutrition^[57-58]. A recently microarray analysis unveiled that cold exposure (10°C) significantly increases glutamine content and the expression of genes related to glutamine metabolism in mice^[59]. However, there is still some debates on this issue. Lian *et al.*^[60] found, through serum metabolite analysis, that glutamine was downregulated in the 3-day cold stress group (4°C), but no significant change was observed in the 7-day cold stress group, indicating a potential adaptation to cold stress. Further studies are needed to elucidate the specific changes in glutamine metabolism and the underlying mechanisms in response to cold exposure.

6 Cold stimulation and metabolic diseases

In recent years, the prevalence of various metabolic diseases, including diabetes, obesity, and cardiovascular disease, has been steadily increasing, largely due to disorders in energy metabolism $^{[61-63]}$. Mild cold acclimation has been shown to significantly improve insulin sensitivity in patients with type 2 diabetes. Notably, Remie et al. $^{[64]}$ conducted a 10-day mild cold adaptation study (16-17°C) and found that observable overt shivering was necessary to produce the beneficial effects of mild cold adaptation on insulin sensitivity. An epidemiologic study has revealed a direct correlation between elevated environmental temperatures and the risk of gestational diabetes mellitus (GDM) and impaired β -cell function, with a higher prevalence of GDM observed during warmer seasons. Cold-induced BAT thermogenesis may explain this seasonal difference in insulin sensitivity $^{[65]}$.

Obesity is a global epidemic, and cold stimulation is being explored as a potential adjunctive therapeutic strategy to alleviate obesity and associated metabolic disorders by triggering coldinduced thermogenesis. While reduced BAT activation is observed in obese individuals, cold exposure activates BAT in humans, promoting weight loss by modulating metabolic levels^[66]. Um *et al.*^[67] discovered that cold exposure (4°C) enhances fat metabolism by upregulating the expression of bone morphogenetic protein 9 (BMP9) and activating the thermogenic gene program in adipocytes, suggesting BMP9 as a potential pharmacological intervention for obesity prevention. Interestingly, a study demonstrated that reducing the ambient temperature from 28°C to 20°C resulted in a 125% increase in body weight and subcutaneous fat mass in obese rats, highlighting the significant impact of temperature reduction on promoting weight gain^[68]. These findings underscore the need for further investigation into the effects of cold stimulation on obesity, considering specific temperatures and duration of cold exposure.

Most cancers rely on glycolysis for energy generation to fuel their uncontrolled growth, invasion, and metastasis. Cold exposure decreases glucose catabolism systemically and glucose utilization in tumors to inhibit tumor progression[69]. Similarly, Tseng et al. [70] found that cold exposure (4°C) decreases circulating glucose levels within tumors, attenuating glycolysis and lipid metabolism, thus impeding tumor growth in mice bearing various solid tumors. . Moreover, Seki et al.[71] demonstrated that cold exposure (4°C or 5°C) significantly inhibits the growth of diverse solid tumors, including clinically challenging cancers like pancreatic cancer, suggesting cold exposure as a potential universal approach for cancer treatment. However, some studies have reported increased tumor growth rates in mice implanted with 4T1 triple-negative breast cancer (TNBC) cells following mild cold stress, indicating diverse effects of cold stimulation on different cancer types^[72].

Cold stimulation can indeed have both beneficial and detrimental effects on lipid metabolism and cardiovascular health. On one hand, it can ameliorate hyperlipidemia and prevent atherosclerosis by enhancing the hydrolysis of triglyceriderich lipoproteins (TRL) and accelerating the production of highdensity lipoprotein (HDL)[73]. However, there are complexities to consider. For example, exposure of apolipoprotein E (ApoE) and low-density lipoprotein receptor (LDLR) deficient mice to cold exacerbates atherosclerosis, indicating that the effects of cold exposure on lipid metabolism and cardiovascular health are multifaceted^[74]. Furthermore, while cold exposure may promote lipoprotein clearance and improve hyperlipidemia, it can also lead to various cardiovascular damages, including alterations in cardiac electrophysiological properties and regulation of ionic currents by AMPK^[75-77]. As such, further research is needed to thoroughly understand the mechanisms underlying cold stress-related cardiovascular diseases and to identify potential therapeutic strategies.

7 Summary

Research on the impacts of cold stimulation on energy metabolism has thus far been rather limited and controversial in certain specific areas. These issues may be attributed at least partially to the present lack of established definition and standardized methodologies relevant to the research field. For example, the impacts of cold exposure on human metabolism are likely dependent on the exposure duration, gender differences, and individual variations, among others. However, these issues remain yet to be elucidated. Despite that many studies have generated evidence in support of cold stimulation as a potential approach for the prevention and treatment of metabolic diseases, large clinical trials, particularly those of randomized studies, are still lacking, leaving the findings from animal model studies skeptical in an effort to extrapolate to humans and whether prolonged cold stimulation exerts negative impacts on human metabolism unresolved. These limitations merit more comprehensive addresses through future studies not only with further in-depth mechanistic explorations but also with more clinically orientated analyses.

Author contributions

Yan M and Wang S J wrote the manuscript. Obtained funding:

Fang S H, E M Y and Yu B. Administrative, technical, or material support: Fang S H. Supervision: Yu B. Yan M and Wang S J contributed equally to this work.

Source of funding

The research was supported by the National Natural Science Foundation of China (No. 82170262, to Fang S H). Heilongjiang Province Applied Technology Research and Development Plan (GA20C009, to E M Y). The Natural Science Foundation of Heilongjiang Province (TD2020H001, to Yu B).

Conflict of interest

Yu B is an Editorial Board Member of Frigid Zone Medicine. The article was subject to the journal's standard procedures, with peer review handled independently of this Member and his research groups.

Data availability statement

Data used to support the findings of this study are available from the corresponding author upon request.

References

- [1] Alahmad B, Khraishah H, Roye D, *et al.* Associations between extreme temperatures and cardiovascular cause-specific mortality: results from 27 countries. Circulation, 2023; 147(1): 35-46.
- [2] Bosy-Westphal A, Hagele F A, Muller M J. What is the impact of energy expenditure on energy intake? Nutrients, 2021; 13(10): 3508
- [3] Liu F, Xiao Y, Ji X L, *et al.* The cAMP-PKA pathway-mediated fat mobilization is required for cold tolerance in C. elegans. Sci Rep, 2017; 7(1): 638.
- [4] Straat M E, Jurado-Fasoli L, Ying Z, et al. Cold exposure induces dynamic changes in circulating triacylglycerol species, which is dependent on intracellular lipolysis: a randomized cross-over trial. EBioMedicine, 2022; 86: 104349.
- [5] Kovanicova Z, Kurdiova T, Balaz M, et al. Cold exposure distinctively modulates parathyroid and thyroid hormones in cold-acclimatized and non-acclimatized humans. Endocrinology, 2020; 161(7): bqaa051
- [6] Chen R, Yin P, Wang L, *et al.* Association between ambient temperature and mortality risk and burden: time series study in 272 main Chinese cities. BMJ, 2018; 363: k4306.
- [7] Chen Y, Ji H, Guo J, *et al.* Non-targeted metabolomics analysis based on LC-MS to assess the effects of different cold exposure times on piglets. Front Physiol, 2022; 13: 853995.
- [8] Ruperez C, Blasco-Roset A, Kular D, et al. Autophagy is Involved in

- cardiac remodeling in response to environmental temperature change. Front Physiol, 2022; 13: 864427.
- [9] Hanssen M J, Hoeks J, Brans B, *et al.* Short-term cold acclimation improves insulin sensitivity in patients with type 2 diabetes mellitus. Nat Med, 2015; 21(8): 863-865.
- [10] Hanssen M J, Wierts R, Hoeks J, *et al.* Glucose uptake in human brown adipose tissue is impaired upon fasting-induced insulin resistance. Diabetologia, 2015; 58(3): 586-595.
- [11] Lahesmaa M, Oikonen V, Helin S, et al. Regulation of human brown adipose tissue by adenosineand A2A receptors studies with [150]H2O and [11C]TMSX PET/CT. Eur J Nucl Med Mol Imaging, 2019; 46(3): 743-750
- [12] Matsushita M, Nirengi S, Hibi M, *et al.* Diurnal variations of brown fat thermogenesis and fat oxidation in humans. Int J Obes (Lond), 2021; 45(11): 2499-2505.
- [13] Lee W C, Guntur A R, Long F. Energy metabolism of the osteoblast: implications for osteoporosis. Endocr Rev, 2017; 38(3): 255-266.
- [14] Riis-Vestergaard M J, Laustsen C, Mariager C O, *et al.* Glucose metabolism in brown adipose tissue determined by deuterium metabolic imaging in rats. Int J Obes (Lond), 2020; 44(6): 1417-1427.
- [15] Kim Y C, Guan K L. mTOR: a pharmacologic target for autophagy regulation. J Clin Invest. 2015; 125(1): 25-32.

- [16] Hua H, Kong Q, Zhang H, et al. Targeting mTOR for cancer therapy. J Hematol Oncol, 2019; 12(1): 71.
- [17] Castro E, Vieira T S, Oliveira T E, et al. Adipocyte-specific mTORC2 deficiency impairs BAT and iWAT thermogenic capacity without affecting glucose uptake and energy expenditure in cold-acclimated mice. Am J Physiol Endocrinol Metab, 2021; 321(5): E592-E605.
- [18] Lewis G F, Brubaker P L. The discovery of insulin revisited: lessons for the modern era. J Clin Invest, 2021; 131(1): e142239.
- [19] Harrison V S, Khan M H, Chamberlain C E, *et al.* The noble and often nobel role played by insulin-focused research in modern medicine. Diabetes Care, 2022; 45(1): 23-27.
- [20] Scheel A K, Espelage L, Chadt A. Many ways to rome: exercise, cold exposure and diet-do they all affect BAT activation and WAT browning in the same manner? Int J Mol Sci, 2022; 23(9): 4759.
- [21] Markan K R, Boland L K, King-McAlpin A Q, *et al.* Adipose TBX1 regulates beta-adrenergic sensitivity in subcutaneous adipose tissue and thermogenic capacity *in vivo*. Mol Metab, 2020; 36: 100965.
- [22] Monfort-Pires M, U-Din M, Nogueira G A, *et al.* Short dietary intervention with olive oil increases brown adipose tissue activity in lean but not overweight subjects. J Clin Endocrinol Metab, 2021; 106(2): 472-484
- [23] Nascimento E B M, Hangelbroek R W J, Hooiveld G, *et al.* Comparative transcriptome analysis of human skeletal muscle in response to cold acclimation and exercise training in human volunteers. BMC Med Genomics, 2020; 13(1): 124.
- [24] Huang S, Cao L, Cheng H, *et al.* The blooming intersection of subfatin and metabolic syndrome. Rev Cardiovasc Med, 2021; 22(3): 799-805.
- [25] Du Q, Wu J, Fischer C, et al. Generation of mega brown adipose tissue in adults by controlling brown adipocyte differentiation in vivo. Proc Natl Acad Sci USA, 2022; 119(40): e2203307119.
- [26] Sellers A J, Pallubinsky H, Rense P, *et al.* The effect of cold exposure with shivering on glucose tolerance in healthy men. J Appl Physiol, 2021; 130(1): 193-205.
- [27] Bahler L, Verberne H J, Admiraal W M, et al. Differences in sympathetic nervous stimulation of brown adipose tissue between the young and old, and the lean and obese. J Nucl Med, 2016; 57(3): 372-377. [28] Merchan-Ramirez E, Sanchez-Delgado G, Arrizabalaga-Arriazu C, et al. Thyroid function is not associated with brown adipose tissue volume and 18F-fluorodeoxyglucose uptake in young euthyroid adults. Eur J Endocrino, 2021; 185(2): 209-218.
- [29] Park H J, Jang H R, Park S Y, *et al.* The essential role of fructose-1,6-bisphosphatase 2 enzyme in thermal homeostasis upon cold stress. Exp Mol Med, 2020; 52(3): 485-496.
- [30] Remmerie A, Scott CL. Macrophages and lipid metabolism. Cell Immunol, 2018; 330: 27-42.
- [31] Li Z, Zhang H. Reprogramming of glucose, fatty acid and amino acid metabolism for cancer progression. Cell Mol Life Sci, 2016; 73(2): 377-392
- [32] Straat M E, Martinez-Tellez B, Sardjoe Mishre A, *et al.* Cold-induced thermogenesis shows a diurnal variation that unfolds differently in males and females. J Clin Endocrinol Metab, 2022; 107(6): 1626-1635.
- [33] Von Bank H, Kirsh C, Simcox J. Aging adipose: depot location dictates age-associated expansion and dysfunction. Ageing Res Rev, 2021; 67: 101259.
- [34] Pernes G, Morgan P K, Huynh K, *et al.* Characterization of the circulating and tissue-specific alterations to the lipidome in response to moderate and major cold stress in mice. Am J Physiol Regul Integr Comp

- Physiol, 2021; 320(2): R95-104.
- [35] Crandall J P, Fraum T J, Wahl R L. Brown adipose tissue: a protective mechanism against "preprediabetes"? J Nucl Med, 2022; 63(9): 1433-1440.
- [36] Cero C, Lea H J, Zhu K Y, et al. beta3-Adrenergic receptors regulate human brown/beige adipocyte lipolysis and thermogenesis. JCI Insight, 2021; 6(11): e139160.
- [37] Chen CC, Kuo C H, Leu Y L, *et al.* Corylin reduces obesity and insulin resistance and promotes adipose tissue browning through SIRT-1 and beta3-AR activation. Pharmacol Res, 2021; 164: 105291.
- [38] Sorensen-Zender I, Bhayana S, Susnik N, et al. Zinc-alpha2-Glycoprotein exerts antifibrotic effects in kidney and heart. J Am Soc Nephrol, 2015; 26(11): 2659-2668.
- [39] Ryden M, Agustsson T, Andersson J, *et al.* Adipose zinc-alpha2-glycoprotein is a catabolic marker in cancer and noncancerous states. J Intern Med, 2012; 271(4): 414-420.
- [40] Fan G, Li Y, Ma F, *et al.* Zinc-alpha2-glycoprotein promotes skeletal muscle lipid metabolism in cold-stressed mice. Endocr J, 2021; 68(1): 53-62
- [41] Bjertnaes L J, Naesheim T O, Reierth E, *et al.* Physiological changes in subjects exposed to accidental hypothermia: an update. Front Med (Lausanne), 2022; 9: 824395.
- [42] Hong A E, Ryu M S, Lim I K. Proper regulation of beta-adrenergic signal requires Btg2 gene for lipolysis and thermogenesis in response to starvation or cold acclimation in female mice. J Nutr Biochem, 2023; 111: 109160.
- [43] Bleher M, Meshko B, Cacciapuoti I, et al. Egr1 loss-of-function promotes beige adipocyte differentiation and activation specifically in inguinal subcutaneous white adipose tissue. Sci Rep, 2020; 10(1): 15842.
- [44] Miao Y, Qin H, Zhong Y, *et al.* Novel adipokine asprosin modulates browning and adipogenesis in white adipose tissue. J Endocrinol, 2021; 249(2): 83-93.
- [45] Romere C, Duerrschmid C, Bournat J, et al. Asprosin, a fasting-induced glucogenic protein hormone. Cell, 2016; 165(3): 566-579.
- [46] Shamsi F, Wang C H, Tseng Y H. The evolving view of thermogenic adipocytes ontogeny, niche and function. Nat Rev Endocrinol, 2021; 17(12): 726-744.
- [47] Shamsi F, Xue R, Huang T L, et al. FGF6 and FGF9 regulate UCP1 expression independent of brown adipogenesis. Nat Commun, 2020; 11(1): 1421.
- [48] Tabe Y, Lorenzi P L, Konopleva M. Amino acid metabolism in hematologic malignancies and the era of targeted therapy. Blood, 2019; 134(13): 1014-1023.
- [49] Li C, Wu B, Li Y, et al. Amino acid catabolism regulates hematopoietic stem cell proteostasis via a GCN2-elF2alpha axis. Cell Stem Cell, 2022; 29(7): 1119-1134
- [50] Yang L, Chu Z, Liu M, *et al.* Amino acid metabolism in immune cells: essential regulators of the effector functions, and promising opportunities to enhance cancer immunotherapy. J Hematol Oncol, 2023; 16(1): 59.
- [51] McGarrah R W, White P J. Branched-chain amino acids in cardiovascular disease. Nat Rev Cardiol, 2023; 20(2): 77-89.
- [52] Huang Y, Zhou M, Sun H, et al. Branched-chain amino acid metabolism in heart disease: an epiphenomenon or a real culprit? Cardiovasc Res, 2011; 90(2): 220-223.
- [53] Li Y, Ping X, Zhang Y, et al. Comparative transcriptome profiling of cold exposure and beta3-AR agonist CL316,243-induced browning of white fat. Front Physiol, 2021; 12: 667698.

- [54] Teng T, Song X, Sun G, *et al.* Glucose supplementation improves intestinal amino acid transport and muscle amino acid pool in pigs during chronic cold exposure. Anim Nutr, 2023; 12: 360-374.
- [55] Yoneshiro T, Wang Q, Tajima K, *et al.* BCAA catabolism in brown fat controls energy homeostasis through SLC25A44. Nature, 2019; 572(7771): 614-619.
- [56] Cannavino J, Shao M, An YA, *et al.* Regulation of cold-induced thermogenesis by the RNA binding protein FAM195A. Proc Natl Acad Sci USA, 2021; 118(23): e2104650118.
- [57] Cruzat V, Macedo Rogero M, Noel Keane K, et al. Glutamine: metabolism and immune function, supplementation and clinical translation. Nutrients, 2018; 10(11): 1564
- [58] Smedberg M, Wernerman J. Is the glutamine story over? Crit Care, 2016; 20(1): 361.
- [59] Okamatsu-Ogura Y, Kuroda M, Tsutsumi R, et al. UCP1-dependent and UCP1-independent metabolic changes induced by acute cold exposure in brown adipose tissue of mice. Metabolism, 2020; 113: 154396.
- [60] Lian S, Li W, Wang D, *et al.* Effects of prenatal cold stress on maternal serum metabolomics in rats. Life Sci, 2020; 246: 117432.
- [61] Shahcheraghi S H, Aljabali A A A, Al Zoubi M S, et al. Overview of key molecular and pharmacological targets for diabetes and associated diseases. Life Sci, 2021; 278: 119632.
- [62] Aubert G, Vega R B, Kelly D P. Perturbations in the gene regulatory pathways controlling mitochondrial energy production in the failing heart. Biochim Biophys Acta, 2013; 1833(4): 840-847.
- [63] Samuel V T, Petersen K F, Shulman G I. Lipid-induced insulin resistance: unravelling the mechanism. Lancet, 2010; 375(9733): 2267-2277
- [64] Remie C M E, Moonen M P B, Roumans K H M, et al. Metabolic responses to mild cold acclimation in type 2 diabetes patients. Nat Commun, 2021; 12(1): 1516.
- [65] Pace N P, Vassallo J, Calleja-Agius J. Gestational diabetes, environmental temperature and climate factors from epidemiological evidence to physiological mechanisms. Early Hum Dev, 2021; 155:

- 105219
- [66] Perez L C, Perez L T, Nene Y, et al. Interventions associated with brown adipose tissue activation and the impact on energy expenditure and weight loss: a systematic review. Front Endocrinol (Lausanne), 2022; 13: 1037458.
- [67] Um J H, Park S Y, Hur J H, *et al.* Bone morphogenic protein 9 is a novel thermogenic hepatokine secreted in response to cold exposure. Metabolism, 2022; 129: 155139.
- [68] Aldiss P, Lewis J E, Lupini I, *et al.* Cold exposure drives weight gain and adiposity following chronic suppression of brown adipose tissue. Int J Mol Sci, 2022; 23(3): 1869
- [69] Yuneva M. Cold exposure as anti-cancer therapy. Cancer Cell, 2022; 40(10): 1092-1094.
- [70] Tseng Y H. Adipose tissue in communication: within and without. Nat Rev Endocrinol, 2023; 19(2): 70-71.
- [71] Seki T, Yang Y, Sun X, et al. Brown-fat-mediated tumour suppression by cold-altered global metabolism. Nature, 2022; 608(7922): 421-428.
- [72] Gandhi S, Oshi M, Murthy V, et al. Enhanced thermogenesis in triple-negative breast cancer is associated with pro-tumor immune microenvironment. Cancers (Basel), 2021; 13(11): 2559.
- [73] Schaltenberg N, John C, Heine M, et al. Endothelial lipase is involved in cold-induced high-density lipoprotein turnover and reverse cholesterol transport in Mice. Front Cardiovasc Med, 2021; 8: 628235.
- [74] Ying Z, van Eenige R, Beerepoot R, et al. Mirabegron-induced brown fat activation does not exacerbate atherosclerosis in mice with a functional hepatic ApoE-LDLR pathway. Pharmacol Res, 2023; 187: 106634
- [75] Kong X, Liu H, He X, et al. Unraveling the mystery of cold stress-induced myocardial injury. Front Physiol, 2020; 11: 580811.
- [76] Kashiwagi Y, Komukai K, Kimura H, *et al.* Therapeutic hypothermia after cardiac arrest increases the plasma level of B-type natriuretic peptide. Sci Rep, 2020; 10(1): 15545.
- [77] Wu C I, Lu Y Y, Chen Y C, et al. The AMP-activated protein kinase modulates hypothermia-induced J wave. Eur J Clin Invest, 2020; 50(6): e13247.