

Review

Ingrid Fricke-Galindo, Adrián Llerena and Marisol López-López*

An update on *HLA* alleles associated with adverse drug reactions

DOI 10.1515/dmpt-2016-0025

Received July 27, 2016; accepted February 7, 2017; previously published online March 18, 2017

Abstract: Adverse drug reactions (ADRs) are considered as an important cause of morbidity and mortality. The hypersensitivity reactions are immune-mediated ADRs, which are dose-independent, unpredictable and have been associated with several *HLA* alleles. The present review aimed to describe *HLA* alleles that have been associated with different ADRs in populations worldwide, the recommendations of regulatory agencies and pharmacoeconomic information and databases for the study of *HLA* alleles in pharmacogenetics. A systematic search was performed in June 2016 of articles relevant to this issue in indexed journals and in scientific databases (PubMed and PharmGKB). The information of 95 association studies found was summarized. Several *HLA* alleles and haplotypes have been associated with ADRs induced mainly by carbamazepine, allopurinol, abacavir and nevirapine, among other drugs. Years with the highest numbers of publications were 2013 and 2014. The majority of the reports have been performed on Asians and Caucasians, and carbamazepine was the most studied ADR drug inducer. Two *HLA* alleles' databases are described, as well as the recommendations of the U.S. Food and Drug Administration, the European Medicine Agency and the Clinical Pharmacogenetics Implementation Consortium. Pharmacoeconomic studies on this issue are also mentioned. The strongest associations remain for *HLA-B*58:01*, *HLA-B*57:01*, *HLA-B*15:02* and *HLA-A*31:01* but only in certain populations; therefore, studies on different ethnic groups would be useful.

***Corresponding author: Marisol López-López**, PhD, Biological Systems Department, Universidad Autónoma Metropolitana, Campus Xochimilco, Calzada del Hueso 1100, Col. Villa Quietud, Coyoacán, 04960 Ciudad de México (CDMX), Mexico, Phone: (+52) (55) 5483 7250, Fax: (+52) (55) 5483 7237, E-mail: mlopez@correo.xoc.uam.mx

Ingrid Fricke-Galindo: Doctorate in Biological and Health Sciences, Metropolitan Autonomous University, Campus Xochimilco, Mexico City, Mexico

Adrián Llerena: CICAB Clinical Research Centre, Extremadura University Hospital and Medical School, Badajoz, Spain

Due to the improvement of drug therapy and the economic benefit that *HLA* screening represents, investigations on *HLA* alleles associated with ADR should continue.

Keywords: abacavir; allopurinol; adverse drug reactions; carbamazepine; human leukocyte antigen (HLA); hypersensitivity syndrome; pharmacogenetics; Stevens-Johnson syndrome.

Introduction

Immunologic-mediated adverse drug reactions (ADRs) are included in type B ADRs which are not related to the dose and that are uncommon and unpredictable in that they are not related with the pharmacodynamics of the drug and present high mortality [1]. There is evidence that susceptibility to at least some Type B ADR is found under strong genetic influence [2], and identifying genetic risk factors for this type of ADRs could significantly decrease health-care costs and improve the process of drug development [3].

The drug-activated immune response is known to be mediated by T cells when the drug molecule binds to T-cell receptors (TCR). The majority of existing drug molecules have a comparable size of one to three amino acids, which is much smaller than the peptide ligands of human leukocyte antigen (HLA) class I (8–12 mers) and class II (9–25 mers) molecules [4]. Accordingly, the following five hypotheses have been proposed in order to explain T-cell recognition of a drug antigen presented by a HLA molecule: (i) the hapten/pro-hapten theory, (ii) the p.i. concept, (iii) the “altered repertoire” model, (iv) the “altered T-cell receptor repertoire” model and (v) the danger hypothesis [4–6] (Figure 1).

Briefly, the first hypothesis postulates that the drug or its immunogenic metabolite binds by covalent bonds to an endogenous peptide and to the HLA molecule, which particularly explains the hypersensitivity to beta-lactam antibiotics. The p.i. (“pharmacological interaction with the immune receptor”) concept considers a non-covalent and reversible binding of the molecule directly to the HLA

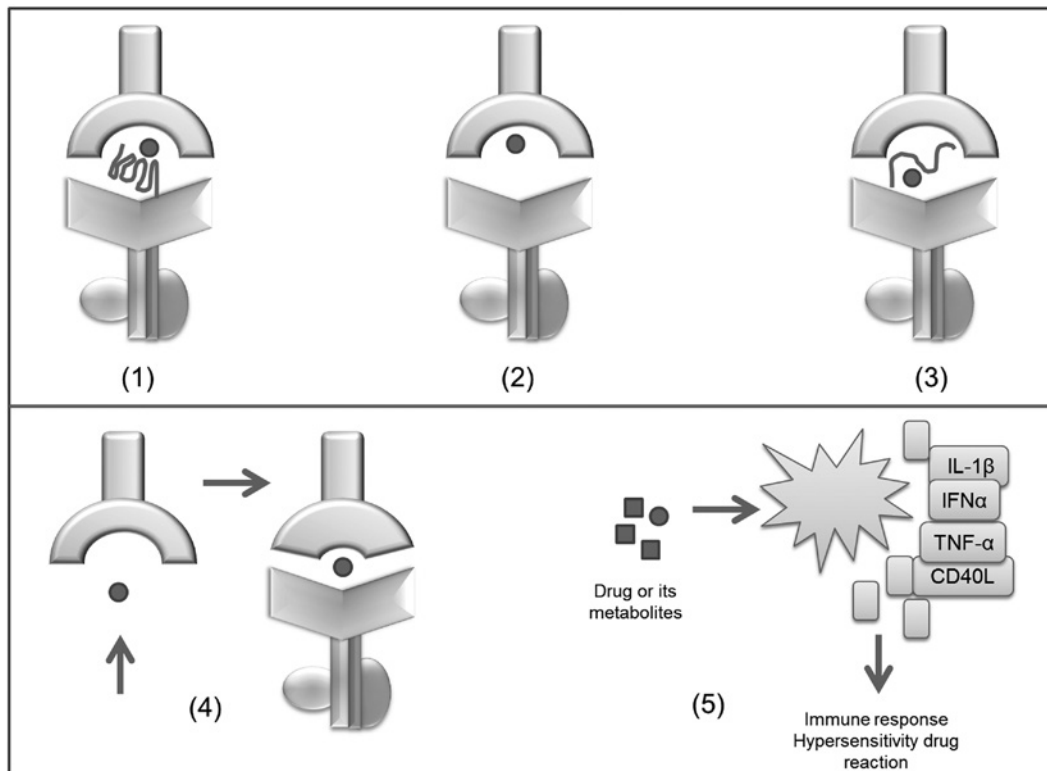


Figure 1: Models of drug interactions with immunology molecules.

(1) Hapten/pro-hapten theory, (2) p.i. concept, (3) “altered repertoire” model, (4) “altered T-cell receptor repertoire” model and (5) danger hypothesis.

molecule and/or the TCR, which could be the case for carbamazepine (CBZ)-induced Stevens-Johnson syndrome (SJS) and the *HLA-B*15:02* allele. The “altered repertoire” model proposes that a drug can non-covalently bind to the self-peptide repertoire presented to HLA and TCR protein, inducing cutaneous adverse drug reactions (cADRs); in this case, the drug may not directly bind to HLA, and this hypothesis elucidates in part the association of *HLA-B*57:01* and abacavir-induced cADRs. The “altered TCR repertoire” model suggests that the drug interacts first with the TCR, which undergoes a conformational change that, in turn, allows binding with an HLA molecule to induce the immune response. Finally, the danger hypothesis explains that immune responses are caused by endogenous cellular alarm signals (i.e. CD40L, TNF- α , IL-1 β and IFN- α) from distressed or injured cells, which could occur after the processing of the drug as antigen [4–6].

As mentioned previously, several *HLA* alleles have been associated with different induced ADRs. The majority of these are related with cADRs, which comprise mild ailments such as maculopapular exanthema (MPE)

and severe syndromes such as the SJS, toxic epidermal necrolysis (TEN) and the drug-induced hypersensitivity syndrome (DIHS)/drug reaction with eosinophilia and systemic symptoms (DRESS) [6, 7]. Furthermore, there are reports that also support the implication of variations in immune genes with drug-induced liver injury (DILI) [8–12], drug-induced proteinuria [13] and agranulocytosis [14].

The most common descriptions of associations of *HLA* alleles with ADRs were reported for *HLA-B*15:02* and CBZ-induced SJS [15], *HLA-B*58:01* and allopurinol-induced SJS or DIHS [16] and *HLA-B*57:01* with a hypersensitivity reaction to abacavir [17]. Over time, several association studies have been performed in different populations, and some of these have described that the associated *HLA* allele is drug-specific and that ethnicity matters [18]. Moreover, guideline recommendations by regulatory agencies have been published in order to prevent different cADRs by means of *HLA* genotyping prior to drug prescription, and databases are now available to supply information on *HLA* alleles associated with ADR throughout the world. Therefore, in the light of these advances in *HLA* alleles in ADRs the present review

aimed to describe *HLA* alleles that have been associated with several ADRs in worldwide populations, the recommendations of regulatory agencies, pharmacoeconomic information and databases for the study on *HLA* alleles in pharmacogenetics.

Methods

We performed a systematic search in June 2016 of relevant articles for this issue in indexed journals and scientific databases (PubMed and PharmGKB). Search words included “HLA and adverse drug reactions”. Association studies in which at least one *HLA* allele was related with the risk of ADR occurrences were included in the corresponding section. Articles were selected according to their relevance on the subject. The whole body of information was analyzed and summarized.

HLA alleles associated with adverse drug reactions

Ninety-five articles reporting that *HLA* alleles have been associated with ADRs were selected, and the main information of the association study was summarized in Table 1 for CBZ-induced cADRs, Table 2 for *HLA* alleles related to other antiepileptic drugs-induced cADRs, Table 3 for allopurinol, abacavir and nevirapine studies with *HLA* and ADRs and Table 4 for reports of *HLA* alleles in ADRs produced by different drugs.

The association of *HLA* alleles with CBZ-induced cADRs has been widely reported for several populations, including Asians (Han Chinese, Japanese, Korean, Thai, Malaysian, Indian, Vietnamese and Singaporean), European, North American and Mexican mestizos. The most significant associations have been described for *HLA-B*15:02* allele in Asian populations; however, important associations have been found for the *HLA-A*31:01* allele in European, North American, Mexican mestizos and Japanese populations. Other alleles associated with CBZ-induced cADRs are *HLA-A*01:01:01*, *-A*02:01*, *-DRB1*14:05*, *-B*51:01*, *-B*15:11*, *-A*02:06*, *-A*24:02*, *-C*08:01* and *-DRB1*12:02* (Table 1). The *HLA-B*15:02* allele also has been associated with other antiepileptic drugs-induced cADRs, such as phenytoin and oxcarbazepine. In addition, other *HLA* alleles have exhibited an association with lamotrigine-, phenobarbital-, phenytoin- and zonisamide-induced cADRs (Table 2).

Several investigations have found a strong association of *HLA-B*58:01* with cADRs induced by allopurinol, mainly in patients with Asian ancestry; however, there are also reports that include European and Portuguese populations in whom this allele has also been associated. Other alleles that have been related with allopurinol-induced cADRs include *HLA-C*03:02* and *-A*33:03* in Koreans and *HLA-DRB1*15:02* and *-DRB1*13:02* in Caucasians. Studies of abacavir hypersensitivity have been mostly performed in Caucasians from the U.S. and Australia. The main associated *HLA* allele is *HLA-B*57:01*, and only one study (to our knowledge) has reported the association of two different alleles from *HLA* class II (*HLA-DR7* and *-DQ3*) with abacavir-induced hypersensitivity. For nevirapine, four different *HLA* alleles in four studies have been found associated with ADRs induced by this drug; two of these reports identified an association between the *HLA-C*04* allele and hypersensitivity in Han Chinese and SJS/NET in Malawian patients. There are also reports that have investigated the relation of *HLA* alleles with hepatotoxicity induced by this drug (Table 3).

There are several reports of other *HLA* alleles associated with other different drugs-induced ADRs, for instance, amoxicillin-clavulanate, antituberculosis drugs, asparaginase, aspirin, clozapine, cold medications, co-trimazole, dapsone and penicillins, among others. The type of ADRs included in these studies comprises hypersensitivity, cADR, drug-induced liver injury, proteinuria, agranulocytosis and other specific reactions, such as aspirin-exacerbated respiratory disease and cold medicine-related SJS/TEN with severe ocular-surface complications. In some cases, the studies have only been performed in one population, and the information remains not sufficiently strong to be considered in clinical recommendations (Table 4).

The studies included comprise years of publication from 1993 to 2016 with at least one study reporting an association between *HLA* alleles and ADRs published per year. The highest number of reports was published in 2013 and 2014 (13 per year), which is 6 years after the U.S. Food and Drug Administration (FDA) recommendation on genetic screening for patients of Asian ancestry prior to the initiation of CBZ therapy [112] and when the number of studies in this subject increased. The majority of the investigations have studied CBZ-induced cADRs (29%), followed by allopurinol- and lamotrigine-induced cADRs (13% and 7%, respectively). Few studies have been performed for abacavir- and phenytoin-induced hypersensitivity (6% for each) and for ADRs induced by amoxicillin-clavulanate, nevirapine and oxcarbazepine (4% for each). Fifty-five percent of the studies have included

Table 1: HLA alleles associated with carbamazepine-induced cutaneous adverse drug reactions.

ADR type	Population	HLA allele	n +/tot	OR	95% CI	p-Value	References
cADR	Japanese	-A*31	10/15	11.20	2.668–47.105	0.001	[19]
cADR	Japanese	-A*31:01	37/61	10.80	5.9–19.6	3.64×10^{-15}	[20]
DRESS	European	-A*31:01	7/10	57.60	11.0–341.0	<0.001	[21]
DRESS	Chinese	-A*31:01	5/10	23.00	4.2–125	<0.001	[21]
HSS	North American	-A*31:01	3/6	26.36	2.53–307.89	0.002	[22]
HSS	European	-A*31:01	10/27	12.41	1.27–121.03	3.5×10^{-8}	[23]
MPE	Mexican mestizo	-A*01:01:01	3/5	7.29	1.02–52.01	0.028	[24]
MPE	Mexican mestizo	-A*31:01:02	2/5	ND	ND	0.006	[24]
MPE	Han Chinese	-A*02:01	11/80 (2n)	ND	ND	0.033	[25]
MPE	Han Chinese	-DRB1*14:05	7/80 (2n)	ND	ND	0.003	[25]
MPE	North America	-A*31:01	6/26	8.57	1.67–57.50	0.004	[22]
MPE	European	-A*31:01	23/106	8.33	3.59–19.36	8.0×10^{-7}	[23]
MPE/DRESS	Han Chinese	-A*31:01	14/74	6.86	2.4–19.9	2.7×10^{-3}	[26]
MPE/DRESS	Han Chinese	-B*51:01	18/74	4.56	2.0–10.5	0.01	[26]
MPE/HSS	Chinese	-A*31:01	8/31	12.17	3.6–41.2	0.0021	[27]
SCAR	Japanese	-A*31:01	11/44 (2n)	4.33	2.07–9.06	0.0004	[28]
SJS	Han Chinese	-B*15:02	44/44	2504	126–49,522	3.1×10^{-27}	[15]
SJS	North American	-B*15:02	3/9	38.65	2.68–2239.5	0.002	[22]
SJS	Chinese and Malaysian	-B*15:02	6/6	ND	ND	0.0345	[29]
SJS	Korean	-B*15:11	3/7	18.00	2.3–141.2	0.011	[30]
SJS	Indian	-B*15:02	6/8	71.40	3.0–1698	0.0014	[31]
SJS	Thai	-B*15:02	6/6	25.50	2.68–242.61	0.0005	[32]
SJS/TEN	Vietnamese	-B*15:02	32/38	33.78	7.55–151.03	<0.001	[33]
SJS/TEN	Han Chinese	-B*15:02	24/26	89.25	19.25–413.83	3.51×10^{-18}	[34]
SJS/TEN	North Indian	-B*15:02	2/9	ND	ND	0.035	[35]
SJS/TEN	Han Chinese	-B*15:02	8/35	18.22	3.66–90.66	0.000	[36]
SJS/TEN	Singaporean	-B*15:02	13/13	181.00	8.7–3785	6.9×10^{-8}	[37]
SJS/TEN	Chinese	-B*15:02	41/53	58.10	17.6–192	<0.001	[21]
SJS/TEN	Singaporean	-B*15:02	5/5	27.20	2.67 to ∞	<0.05	[38]
SJS/TEN	Han Chinese	-B*15:02	24/26	89.25	19.25–413.83	3.51×10^{-18}	[39]
SJS/TEN	Japanese	-A*02:06	51/110	5.07	ND	6.9×10^{-10}	[40]
SJS/TEN	Han Chinese	-A*24:02	9/16	3.18	1.11–9.11	0.03	[41]
SJS/TEN	Han Chinese	-B*15:02	13/18	17.55	5.31–58.06	<0.001	[41]
SJS/TEN	Thai	-B*15:02	32/34	75.40	13.0–718.9	<0.001	[42]
SJS/TEN	European	-A*31:01	5/12	25.93	4.93–116.18	8.0×10^{-5}	[23]
SJS/TEN	Han Chinese	-B*15:02	16/17	152.00	12–1835	<0.0001	[43]
SJS/TEN	Han Chinese	-B*15:02	9/9	114.83	6.25–2111.03	<0.001	[44]
SJS/TEN	Malaysian	-B*15:02	12/17	16.15	4.57–62.4	7.87×10^{-6}	[45]
SJS/TEN	Japanese	-B*15:11	4/28	16.30	4.76–55.6	0.0004	[46]
SJS/TEN	Chinese	-B*15:02	8/8	184.00	33.2–1021.0	<0.05	[47]
SJS/TEN	Thai	-B*15:02	37/42	54.76	14.62–205.13	2.89×10^{-12}	[48]
SJS/TEN	Chinese	-B*15:02	59/60	1357.00	193.4–8838.3	1.6×10^{-41}	[27]
SJS/TEN	Chinese	-C*08:01	56/60	86.80	29.3–254.6	7.8×10^{-27}	[27]
SJS/TEN	Chinese	-DRB1*12:02	41/60	11.40	5.6–22.9	2.3×10^{-11}	[27]
SJS/TEN > 5% skin detachment	Han Chinese	-B*15:02	25/25	97.60	42.0–226.8	5.8×10^{-43}	[26]

ADR, adverse drug reaction; cADR, cutaneous adverse drug reaction; CI, confidence interval; DRESS, drug reactions with eosinophilia and systemic symptoms; HLA, human leukocyte antigen; HSS, hypersensitivity syndrome; MPE, maculopapular exanthema; ND, not determined; n +/tot, number of subjects positive for the HLA allele associated/total of subjects included in the association study; OR, odds ratio; SCAR, severe cutaneous adverse drug reactions; SJS, Stevens-Johnson syndrome; TEN, toxic epidermal necrolysis.

Asian patients in whom there is a notably higher incidence of cADRs than in Caucasians [113]; however, an important percentage of these reports was also performed

in individuals of Caucasian ancestry (42%); Africans have only been included in one study and individuals from America in three.

Table 2: *HLA* alleles associated with antiepileptic drugs-induced cutaneous adverse drug reactions.

Drug	ADR	Population	<i>HLA</i> allele	n + /tot	OR	95% CI	p-Value	References
Antiepileptic drugs	SJS/TEN	Han Chinese	-B*15:02	6/6	71.9	3.7–1,415.8	1.48×10^{-4}	[49]
	SJS/TEN	Han Chinese	-B*15:02	15/27	18.75	5.29–66.44	0.000	[50]
Lamotrigine	cADR	Norwegian	-A*24:02	10/28	ND	ND	0.027	[51]
	cADR	Japanese	-DRB1*04:05	6/16	ND	ND	0.01	[52]
	cADR	Japanese	-DQB1*04:01	6/16	ND	ND	0.01	[52]
	MPE	Korean	-A*24:02	15/21	4.09	1.22–13.69	0.025	[53]
	MPE	Korean	-A*24:02/ -C*01:02	10/21	7.88	1.81–34.28	0.007	[53]
	MPE	Mexican mestizo	-A*02:01:01/ -B*35:01:01/ -C*04:01:01	4/10	18.33	1.99–169.08	0.0048	[24]
	MPE	Han Chinese	-A*30:01	6/86 (2n)	ND	ND	0.013	[25]
	MPE	Han Chinese	-B*13:02	6/86 (2n)	ND	ND	0.013	[25]
	SCAR	European	-A*68:01	4/44 (2n)	19.22	1.01–365	0.012	[54]
	SJS/TEN	Korean	-B*44:03	3/5	ND	ND	0.099	[55]
Oxcarbazepine	MPE	Han Chinese	-B*38:02	3/28 (2n)	3.33	1.78–22.46	0.018	[56]
	MPE	Han Chinese	-B*13:02	4/28 (2n)	7.83	2.32–26.41	0.001	[57]
	MPE	Han Chinese	-B*15:02	4/9	8.8	1.85–41.79	0.011	[58]
	SJS	Han Chinese	-B*15:02	3/3	80.7	3.8–1714.4	8.4×10^{-4}	[59]
Phenobarbital	SJS/TEN	Japanese	-B*51:01	6/8	16.71	3.66–83.06	0.004	[60]
Phenytoin	DRESS	Malay	-B*15:13	3/3	59	2.49–1395.74	0.003	[61]
	MPE	Mexican mestizos	-C*08:01	2/2	ND	ND	0.002	[24]
	SJS	Thai	-B*15:02	4/4	18.5	1.82–188.40	0.005	[32]
	SJS/TEN	Malay	-B*15:13	7/13	11.28	2.25–59.60	0.003	[61]
	SJS/TEN	Malay	-B*15:02	8/13	5.71	1.41–23.10	0.016	[61]
	SJS/TEN	Han Chinese	-B*15:02	7/15	3.44	1.08–11.00	0.047	[34]
	SJS/TEN	Han Chinese	-B*15:02	7/15	3.5	1.10–11.18	0.045	[39]
	SJS/TEN	Han Chinese	-B*15:02	8/26	5.1	1.8–15.1	0.0041	[59]
	SJS/TEN	Han Chinese	-B*13:01	9/26	3.7	1.4–10.0	0.0154	[59]
	SJS/TEN	Han Chinese	-C*08:01	9/26	3	1.1–7.8	0.0281	[59]
Zonisamide	SJS/TEN	Han Chinese	-DRB1*16:02	7/26	4.3	1.4–12.8	0.0128	[59]
	SJS/TEN	Japanese	-A*02:07	5/12	9.77	3.07–31.1	0.018	[60]

ADR, adverse drug reaction; cADR, cutaneous adverse drug reaction; CI, confidence interval; DRESS, drug reactions with eosinophilia and systemic symptoms; HLA, human leukocyte antigen; MPE, maculopapular exanthema; ND, not determined; n + /tot, number of subjects positive for the *HLA* allele associated/total of subjects included in the association study; OR, odds ratio; SCAR, severe cutaneous adverse drug reactions; SJS, Stevens-Johnson syndrome; TEN, toxic epidermal necrolysis.

Databases with *HLA* alleles and adverse drug reactions

As can be observed in Tables 1–4, several class I and II *HLA* alleles have been associated with ADRs in different populations. The *HLA* allele associated in a certain ethnic group is largely influenced by the frequency of the specific allele among the studied population [18, 114]. In order to facilitate the investigation and associations of *HLA* alleles and ADRs, two databases have been designed [115, 116]. Both of these permit easy and rapid access to relevant information about a specific drug, ADR or *HLA* allele related with ADR pharmacogenomics.

One of these belongs to the Allele Frequency Net Database (<http://www.allelefrequencies.net>), a free, centralized resource that contains information on the allele, the haplotype and the genotype frequencies of several immune genes, including *HLA* alleles, in different populations [117]. This website was released in 2003 [118], and since then, the database has grown significantly in terms of the number of populations covered and the number of users and citations [115]. Therefore, to assist *HLA* and pharmacogenetic studies, this database contains collated data sets from the literature, with information that not only facilitates meta-analyses but that also enables users to examine the quality of published studies by comparing the frequencies of *HLA* alleles reported in healthy volunteers' cohorts with those

Table 3: HLA alleles associated with adverse reactions induced by allopurinol, abacavir and nevirapine.

Drug	ADR	Population	HLA allele	n +/tot	OR	95% CI	p-Value	References
Allopurinol	cADR	Japanese	-B*58:01	4/7	65.6	2.9–1497.0	9.73×10^{-4}	[62]
	cADR	Han Chinese	-B*58:01	38/38	580.07	32.18–10456.80	7.01×10^{-18}	[63]
	MPE	Han Chinese	-B*58:01	26/40	8.5	4.2–17.5	2.3×10^{-9}	[64]
	SCAR	Han Chinese	-B*58:01	96/106	44	21.5–90.3	2.6×10^{-41}	[64]
	SCAR	Han Chinese	-B*58:01	87/92	127.6	40.8–398.6	<0.001	[65]
	SCAR	Han Chinese	-B*58:01	45/48	108.5	33.7–346.3	<0.0001	[66]
	SCAR	Portuguese	-B*58:01	16/25	39.11	4.49–340.51	5.9×10^{-4}	[67]
	SCAR	Han Chinese	-B*58:01	19/19	229.7	11.7–4520.4	<0.0001	[68]
	SCAR	Korean	-B*58:01	23/25	97.8	18.3–521.5	2.45×10^{-11}	[69]
	SCAR	Korean	-C*03:02	23/25	82.1	15.8–426.5	9.39×10^{-11}	[69]
	SCAR	Korean	-A*33:03	22/25	20.5	5.4–78.6	3.31×10^{-6}	[69]
	SCAR	Han Chinese	-B*58:01	51/51	580.3	34.4–9780.9	4.7×10^{-24}	[16]
	SCAR	Caucasian	-B*58:01	3/7	13.6	2.77–69.45	0.003	[70]
	SCAR	Caucasian	-DRB1*15:02	2/7	22.6	3.28–160.72	0.005	[70]
	SCAR	Caucasian	-DRB1*13:02	2/7	11.1	1.94–7.64	0.037	[70]
	SJS/TEN	Korean	-B*58:01	5/7	ND	ND	0.013	[71]
	SJS/TEN	Thai	-B*58:01	27/27	348.3	19.2–6336.9	<0.001	[72]
	SJS/TEN	Japanese	-B*58:01	4/20 (2n)	40.83	10.5–158.9	<0.0001	[73]
	SJS/TEN	European	-B*58:01	15/27	80	34–187	< 10^{-6}	[74]
Abacavir	AHS	Caucasian	-B*57:01	18/20	6934	321–149,035	<0.0001	[75]
	AHS	White from U.S.	-B*57:01	42/42	1945	110–34,352	<0.0001	[17]
	AHS	Black from U.S.	-B*57:01	5/5	900	38–21,045	<0.0001	[17]
	AHS	Caucasian	-B*57:01	7/7	ND	ND	<0.001	[76]
	AHS	Western Australian	-B*57:01	17/18	960	NS	<0.00001	[77]
	AHS	North American	-B*57	39/84	23.6	8.0–70.0	<0.01	[78]
	AHS	Western Australian	-B*57:01	14/18	117	29–481	<0.0001	[79]
	AHS	Western Australian	-B*5701, -DR7, -DQ3	13/18	822	43–15,675	<0.0001	[79]
Nevirapine	cADR	Thai	-B*35:05	25/243	18.96	4.87–73.44	4.6×10^{-6}	[80]
	Hepatotoxicity	South African	-B*58:01	12/51	ND	ND	0.033	[81]
	Hepatotoxicity	South African	-DRB1*01:02	8/51	ND	ND	0.044	[81]
	Hypersensitivity	Han Chinese	-C*04	8/64	3.61	1.13–11.49	0.03	[82]
	SJS/TEN	Malawian	-C*04:01	23/36	17.52	3.31–92.8	NS	[83]
Nevirapine and Efavirenz	Hypersensitivity	French	-DRB1*01	5/6	ND	ND	0.04	[84]

ADR, adverse drug reaction; AHS, abacavir hypersensitivity; cADR, cutaneous adverse drug reaction; CI, confidence interval; HLA, human leukocyte antigen; MPE, maculopapular exanthema; ND, not determined; NS, not specified; n +/tot, number of subjects positive for the HLA allele associated/total of subjects included in the association study; OR, odds ratio; SCAR, severe cutaneous adverse drug reactions; SJS, Stevens-Johnson syndrome; TEN, toxic epidermal necrolysis.

of worldwide populations. This resource only includes case-control studies with statistical evidence provided for the association and in which high-resolution HLA genotyping was performed. From each report, information about ethnicity, the drug of interest and the proportion of cases and controls carrying the HLA allele implicated in ADRs is extracted and summarized in the database [115].

The second of these is denominated the HLADR database (<http://pgx.fudan.edu.cn/hladr/>). This is an in-house database that manually collected the information of 1786 records after an extensive search and review of the literature and curation of reports of HLA alleles associated with ADRs. The information compiled from each report contains the following: drugs, ADRs, HLA alleles, the ethnic and

geographic origins of the study subjects, a 2×2 contingency table reflecting the association strengths (p-value, odds ratio [OR], sensitivity, specificity, positive-predictive values and negative-predictive values) and FDA drug-labeling changes resulting from the drug-HLA associations. The association strengths across different studies were incomparable because different statistical methods were applied in the original reports; consequently, the designers of this database performed the Fisher's exact test and the Haldane's modification to recalculate the p-value and the OR for each association study, respectively. In addition, this resource standardized the names of ADRs and HLA alleles based on reference databases such as PharmGKB and international ImmunoGeneTics (IMGT) project/HLA [116].

Table 4: HLA alleles associated with different adverse reactions induced by several drugs.

Drug	ADR	Population	HLA allele	n + /tot	OR	95% CI	p-Value	References
Amoxicillin-clavulanate	DILI	Spaniard	-A*30:02	11/75	6.7	2.8–15.9	5×10^{-5}	[85]
	DILI	European	-DQB1*06:02	NS	4.2	2.7–6.6	4.6×10^{-10}	[10]
	DILI	European	-DRB1*15	32/61	2.59	1.44–4.68	0.002	[86]
	DILI	Belgian	-DRB1*15:01/ -DRB5*01:01/-DQB1*06:02 -DQB1*05/*05	20/35	ND	ND	<0.0002	[87]
Antituberculosis treatment	ATLI	Chinese	-DQB1*05/*05	10/88	4.56	0.98–21.22	0.053	[88]
Asparaginase Aspirin	DIH	Indian	-DQB1*02:01	25/47	2.2	1.19–4.15	<0.01	[89]
	HSS	Korean	-C*04:01	7/60	6.9	2.20–21.66	0.0204	[90]
	Allergy	European descent	-DRB1*07:01	66/143	1.92	5.129–2.87	0.023	[91]
	Aspirin desensitization	Iranian	-DQB1*03:02	7/14	0.12	0.02–0.76	0.022	[92]
	related to AERD							
Benznidazole Bucillamine Clozapine	AERD	Iranian	-DQB1*03:02	16/66 (2n)	5.49	2.40–12.59	<0.01	[93]
	AERD	Iranian	-DQA1*03:01	20/66 (2n)	2.9	1.49–5.67	<0.01	[93]
	AERD	Iranian	-DRB4	28/66 (2n)	2.94	1.61–5.36	<0.01	[93]
	Moderate and severe cADRs	Spaniard	-B*3505	5/11	NS	NS	0.033	[94]
	Proteinuria	Japanese	-DRB1*08:02	7/25	25.17	7.98–79.38	1.96×10^{-5}	[95]
	Proteinuria	Japanese	-DQB1*04:02	8/25	10.35	3.99–26.83	2.69×10^{-4}	[95]
	Agranulocytosis	Non-Jewish Caucasian	-DRB5*0201	5/84 (2n)	22.15	1.74–∞	0.005	[96]
Cold medications	Agranulocytosis	Jewish Caucasian	-DRB1*04:02, -DRB4*01:01, DQB1*03:02, DQA1*03:01, DPB1*04:01	11/24 (2n)	6.8	ND	0.02	[97]
	CM-SJS/TEN with SOC	Indian	-B*44:03	12/20	12.25	3.57–42.01	2.14.E–05	[98]
	CM-SJS/TEN with SOC	Brazilian	-B*44:03	10/39	2.74	1.12–6.71	0.0478	[98]
Co-trimazole	CM-SJS/TEN with SOC	Korean	-A*02:06	11/31	3	1.18–7.57	0.0362	[98]
	SJS/TEN	Thai	-B*15:02	14/43	3.91	1.42–10.92	0.02	[99]
	SJS/TEN	Thai	-C*06:02	5/43	11.84	1.24–566.04	0.013	[99]
	SJS/TEN	Thai	-C*08:01	12/43	3.53	1.21–10.40	0.011	[99]
	SJS/TEN	Thai	-B*15:02/-C*08:01	11/43	4.87	1.48–17.22	0.004	[99]
Dapsone	HSS	Chinese descent	-B*13:01	65/76	20.53	11.55–36.48	6.84×10^{-25}	[100]
	DIHR	Southern Chinese	-B*13:01	18/20	122.1	23.5–636.2	6.038×10^{-12}	[101]
Flucloxacillin	DILI	European	-B*57:01	43/51	80.6	22.8–284.9	8.97×10^{-19}	[8]
	Liver injury	Majority White, not Hispanic/Latino	-DRB1*07:01	29/37	14.12	6.36–31.32	2.4×10^{-13}	[102]
Methazolamide	SJS/TEN	Han Chinese	-B*59:01	7/8	305	11.3–8259.9	6.3×10^{-7}	[103]
	SJS/TEN	Korean	-B*59:01	5/5	249.8	13.4–4813.5	<0.001	[104]
Multiple drugs	SJS/TEN with ocular complications	Japanese	-A*02:06	19/40	5.1	NS	<0.0005	[105]
	Anaphylactoid reaction	Spaniard	-DR11	25/42 (2n)	7.3	2.8–19.0	0.02	[106]

Table 4 (continued)

Drug	ADR	Population	<i>HLA</i> allele	n + /tot	OR	95% CI	p-Value	References
Penicillins	Immediate hypersensitive reaction	Han Chinese	- <i>DRB9</i>	9/37	ND	ND	0.019	[107]
Salazosulfapyridine	Urticaria		- <i>DRB9</i>	10/37	ND	ND	0.005	[107]
Sodium aurothiomalate	DRESS	Han Chinese	- <i>B*13:01</i>	4/6	13	ND	0.004	[108]
Sulphasalazine	Mucocutaneous side effects	NS	- <i>DR1</i>	11/16	ND	ND	0.004	[109]
Ticlopidine	SLE-like symptoms	Caucasian	- <i>DRB1*03:01</i>	3/4	ND	ND	0.012	[110]
	Hepatotoxicity	Japanese	- <i>A*33:03</i>	15/22	13.04	4.40–38.59	1.24×10^{-5}	[111]
	Cholestatic hepatotoxicity	Japanese	- <i>A*33:03</i>	12/14	36.5	7.25–183.82	7.32×10^{-7}	[111]

ADR, adverse drug reaction; AERD, aspirin-exacerbated respiratory disease; ATLL, antitubercular drug-induced liver injuries; cADR, cutaneous adverse drug reaction; CI, confidence interval; CM-SJS/TEN with SOC, cold medicine-related SJS/TEN with severe ocular-surface complications; DIH, drug-induced hepatotoxicity; DIHR, dapsone-induced hypersensitivity reactions; *HLA*, human leukocyte antigen; DILI, drug-induced liver injury; DRESS, drug reactions with eosinophilia and systemic symptoms; HSS, hypersensitivity syndrome; MPE, maculopapular exanthema; ND, not determined; NS, not specified; n + /tot, number of subjects positive for the *HLA* allele associated/total of subjects included in the association study; OR, odds ratio; SJS, Stevens-Johnson syndrome; SLE, systemic lupus erythematosus; TEN, toxic epidermal necrolysis.

International medicine agencies recommendations on *HLA* alleles and drug prescriptions

International medicine agencies as the FDA in the U.S. and the European medicine agency (EMA) have made recommendations based on evidence published about *HLA* alleles associated with ADRs in drugs such as CBZ, allopurinol, abacavir and phenytoin, among others. These data have been added to the specific drug-label information in order to improve the safety of the drug and to inform physicians about the usefulness of performing *HLA* genotyping prior to prescribing the drug [112, 119]. In addition, the clinical pharmacogenetics implementation consortium (CPIC) has published guidelines for the dosing of abacavir, allopurinol, CBZ and phenytoin considering the patient's *HLA* genotype [120–127]. Recommendations from the three previously mentioned international medicine agencies are summarized in Table 5. For abacavir, the international medicine agencies and the CPIC highly recommend *HLA* genotyping in all patients with HIV of any ethnicity prior the prescription of this drug and avoiding the administration of abacavir in patients who are carriers of the *HLA-B*57:01* allele. In contrast, for allopurinol there is no well-established recommendation for routine *HLA* testing, although EMA and CPIC consider that the prescription of allopurinol should be evaluated carefully if it is known that the patient is an *HLA-B*58:01* carrier. In the case of CBZ, genetic testing for *HLA* is recommended in patients with Asian ancestry or, specifically, for Han Chinese and Thai population, but the use of this antiepileptic is contraindicated in any patient positive for the *HLA-B*15:02* allele regardless of the patient's ancestry. In addition, the agencies recommend that the use of CBZ in patients carrying *HLA-A*31:01*, especially of Caucasian or Japanese origin, should be considered if the benefits are thought to exceed the risks. Additionally, the recommendation for antiepileptic drug prescription is to avoid phenytoin and, in some cases, oxcarbazepine, eslicarbazepine acetate and lamotrigine in patients positive for *HLA-B*15:02*.

Pharmacoeconomic perspective of *HLA* genotyping prior to drug prescription

Among the benefits of the use of pharmacogenomics in clinical practice are those of guiding the initial drug

Table 5: International medicine agencies' recommendations on HLA genotyping and drug hypersensitivity.

Drug	HLA allele	Publisher	Recommendation ^a	References
Abacavir	-B*57:01	FDA	All patients should be screened for the <i>HLA-B*57:01</i> allele prior to initiating therapy with abacavir or reinitiation of therapy with abacavir, unless patients have a previously documented <i>HLA-B*57:01</i> allele assessment. Abacavir is contraindicated in patients with a prior hypersensitivity reaction to abacavir and in <i>HLA-B*57:01</i> -positive patients	[128]
	-B*57:01	EMA	Before initiating treatment with abacavir, screening for carriage of the <i>HLA-B*57:01</i> allele should be performed in any HIV-infected patient, irrespective of racial origin. Abacavir should not be used in patients known to carry the <i>HLA-B*57:01</i> allele	[129]
	-B*57:01	CPIC	<i>HLA-B*57:01</i> screening should be performed in all abacavir-naïve individuals before initiation of abacavir-containing therapy. In abacavir-naïve individuals who are <i>HLA-B*57:01</i> -positive, abacavir is not recommended and should be considered only under exceptional circumstances when the potential benefit, based on resistance patterns and treatment history, outweighs the risk	[122, 123]
Allopurinol	-B*58:01	EMA	The use of genotyping as a screening tool to make decisions about treatment with allopurinol has not been established. Routine testing for <i>HLA-B*58:01</i> is not recommended in any patient. If the patient is a known carrier of <i>HLA-B*58:01</i> , the use of allopurinol may be considered if the benefits are thought to exceed risks. Extra vigilance for signs of hypersensitivity syndrome or SJS/TEN is required, and the patient should be informed of the need to stop treatment immediately at the first appearance of symptoms	[130]
	-B*58:01	CPIC	Allopurinol is contraindicated in patients who are carriers of <i>HLA-B*58:01</i> /*X, <i>HLA-B*58:01</i> / <i>HLA-B*58:01</i>	[124, 125]
Carbamazepine	-B*15:02	FDA	Prior to initiating carbamazepine therapy, testing for <i>HLA-B*15:02</i> should be performed in patients with ancestry in populations in which <i>HLA-B*15:02</i> may be present. Carbamazepine should not be used in patients positive for <i>HLA-B*15:02</i> unless the benefits clearly outweigh the risks	[128]
	-A*31:01	FDA	The risks and benefits of Tegretol therapy should be weighed before considering administering Tegretol in patients known to be positive for <i>HLA-A*31:01</i>	[128]
	-B*15:02	EMA	Individuals of Han Chinese and Thai origin should, whenever possible, be tested for the <i>HLA-B*15:02</i> allele prior to treatment with carbamazepine. Testing for the <i>HLA-B*15:02</i> allele in other Asian populations at genetic risk may be considered	[130]
Phenytoin	-A*31:01	EMA	Routine testing for the <i>HLA-A*31:01</i> allele is not recommended. If European Caucasians or patients of Japanese descent are known to be positive for the <i>HLA-A*31:01</i> allele, the use of carbamazepine may be considered if the benefits are thought to exceed the risks	[130]
	-B*15:02	CPIC	Regardless of the individual's ancestry or age, if the genetic testing results are "positive" for the presence of at least one copy of the <i>HLA-B*15:02</i> allele, it is recommended that a different agent be employed depending on the underlying disease, unless the benefits clearly outweigh the risk	[127]
	-B*15:02	FDA	Consideration should be given to avoid phenytoin as an alternative for carbamazepine in patients positive for <i>HLA-B*15:02</i>	[128]
	-B*15:02	CPIC	Regardless of the <i>CYP2C9</i> genotype and individual's ancestry or age, if the <i>HLA-B*15:02</i> test result is "positive", the recommendation is to consider administering an anticonvulsant other than carbamazepine and phenytoin unless the benefits of treating the underlying disease clearly outweigh the risks. Alternative medications such as oxcarbazepine, eslicarbazepine acetate and lamotrigine have some evidence linking SJS/TEN with the <i>HLA-B*15:02</i> allele; thus, caution should be exercised in choosing alternatives to phenytoin	[126]

CPIC, clinical pharmacogenetics implementation consortium; FDA, food and drug administration; EMA, European medicine agency; HLA, human leukocyte antigen. ^aThis information is part of the text included as a recommendation in published articles and labeling information of each drug

regimen, individualizing the regimen, increasing efficacy and avoiding ADRs [131]; thus, the implementation of pharmacogenomics in routine practice can improve different drug treatments [132].

Pharmacogenetic testing is also of relevance for the economic resources of the patient or the public health system. Several studies have demonstrated that *HLA* testing prior to various drug prescriptions is cost-effective but that this also depends on the population to which it will be applied. For instance, *HLA-B*15:02* genotyping prior to CBZ prescription is cost-effective for Singaporean Chinese, Malays and Thai patients [133, 134] but not for Singaporean Indians [133]. For Europeans, a study reports that testing for *HLA-A*31:01* represents a cost-effective use prior to initiation of CBZ therapy [135]. Contrariwise, testing for *HLA-B*58:01* did not represent a benefit in allopurinol therapy in Singaporean patients, in contrast with the profit observed in Thai and Korean populations [71, 136]. *HLA-B*57:01* screening prior to abacavir was found to be cost-effective for patients from Spain and Germany [137, 138]. However, a systematic review found that there is cost-effectiveness in the use of pharmacogenetic biomarkers to avoid ADRs in common drug therapies [139].

On the other hand, several investigations have been performed in order to develop assays that reduce the cost of *HLA* screening, i.e. nested allele-specific PCR and PCR-RFLP for *HLA-A*31:01* detection [140, 141], flow cytometry pre-screening [142], real-time PCR [143] for *HLA-B*57:01* and the flow cytometry test for *HLA-B*58:01* [144]. Furthermore, the search for single-nucleotide polymorphisms in linkage disequilibrium with specific *HLA* alleles could diminish the cost of *HLA* genotyping and make it more available for different populations [145, 146].

However, there are some limitations for *HLA* genotyping in cases of rare ADRs that should be taken into account. In general, the incidence of ADRs associated with *HLA* alleles is low; for instance, allopurinol hypersensitivity has been reported in 0.1% [147], and it represents a problem in association studies. This explains the small sample size observed in the majority of studies, which limits the study power for detecting significant associations [148]; however, meta-analyses can help solve this problem, especially in populations that are widely studied. In addition, investigations employing a scarce number of patients could lead to many associated *HLA* alleles, in which further studies are required in order to confirm their use as pharmacogenetic biomarkers. This situation could impact the cost-effectiveness of *HLA* typing prior to drug prescription in uncommon ADRs [149].

In sum, several *HLA* alleles have been associated with ADRs to common drugs; however, the strongest associations remain for *HLA-B*58:01*, *HLA-B*57:01*, *HLA-B*15:02* and *HLA-A*31:01*, but only in certain populations. In addition, the majority of the studies have been performed in Asians and Caucasians, and there is a lack of reports on other populations. This situation renders it difficult for international medicine agencies to offer global recommendations on *HLA* genotyping prior to drug prescription or for their including more populations in their warnings. Moreover, it might be possible that more *HLA* alleles are associated with ADRs in other, not yet studied ethnic groups; therefore, pharmacogenomic investigations on this issue should not be pushed aside. The existence of *HLA* allele databases that facilitate these studies and the reports that assure the cost-effectiveness of *HLA* screening could ease pharmacogenomic research of ADRs focused on *HLA* alleles.

Author contributions: All the authors have accepted responsibility for the entire content of this submitted manuscript and approved submission.

Research funding: This research was supported by a grant from Consejo Nacional de Ciencia y Tecnología de México (CONACyT) (#167261). IFG was supported by a grant (doctoral degree) from CONACyT (#369708).

Employment or leadership: None declared.

Honorarium: None declared.

Competing interests: The funding organization(s) played no role in the study design; in the collection, analysis and interpretation of data; in the writing of the report; or in the decision to submit the report for publication.

References

1. Edwards IR, Aronson JK. Adverse drug reactions: definitions, diagnosis, and management. *Lancet* (London, England) 2000;356:1255–9.
2. Molokhia M, McKeigue P. EUDRAGENE: European collaboration to establish a case-control DNA collection for studying the genetic basis of adverse drug reactions. *Pharmacogenomics* 2006;7:633–8.
3. Giacomini KM, Krauss RM, Roden DM, Eichelbaum M, Hayden MR, Nakamura Y. When good drugs go bad. *Nature* 2007;446:975–7.
4. Illing PT, Mifsud NA, Purcell AW. Allotype specific interactions of drugs and HLA molecules in hypersensitivity reactions. *Curr Opin Immunol* 2016;42:31–40.
5. Yun J, Cai F, Lee FJ, Pichler WJ. T-cell-mediated drug hypersensitivity: immune mechanisms and their clinical relevance. *Asia Pac Allergy* 2016;6:77–89.
6. Chung WH, Wang CW, Dao RL. Severe cutaneous adverse drug reactions. *J Dermatol* 2016;43:758–66.

7. Schöpf E, Stühmer A, Rzany B, Victor N, Zentgraf R, Kapp JF. Toxic epidermal necrolysis and Stevens-Johnson syndrome. An epidemiologic study from West Germany. *Arch Dermatol* 1991;127:839–42.
8. Daly AK, Donaldson PT, Bhatnagar P, Shen Y, Pe'er I, Floratos A, et al. HLA-B*5701 genotype is a major determinant of drug-induced liver injury due to flucloxacillin. *Nat Genet* 2009;41:816–9.
9. Kindmark A, Jawaid A, Harbron CG, Barratt BJ, Bengtsson OF, Andersson TB, et al. Genome-wide pharmacogenetic investigation of a hepatic adverse event without clinical signs of immunopathology suggests an underlying immune pathogenesis. *Pharmacogenomics J* 2008;8:186–95.
10. Lucena MI, Molokhia M, Shen Y, Urban TJ, Aithal GP, Andrade RJ, et al. Susceptibility to amoxicillin-clavulanate-induced liver injury is influenced by multiple HLA class I and II alleles. *Gastroenterology* 2011;141:338–47.
11. Parham LR, Briley LP, Li L, Shen J, Newcombe PJ, King KS, et al. Comprehensive genome-wide evaluation of lapatinib-induced liver injury yields a single genetic signal centered on known risk allele HLA-DRB1*07:01. *Pharmacogenomics J* 2016;16:180–5.
12. Singer JB, Lewitzky S, Leroy E, Yang F, Zhao X, Klickstein L, et al. A genome-wide study identifies HLA alleles associated with lumiracoxib-related liver injury. *Nat Genet* 2010;42:711–4.
13. Ueda S, Wakashin M, Wakashin Y, Yoshida H, Iesato K, Mori T, et al. Experimental gold nephropathy in guinea pigs: detection of autoantibodies to renal tubular antigens. *Kidney Int* 1986;29:539–48.
14. Hodinka L, Géher P, Merétey K, Gyódi EK, Petrányi GG, Bozsóky S. Levamisole-induced neutropenia and agranulocytosis: association with HLA B27 leukocyte agglutinating and lymphocytotoxic antibodies. *Int Arch Allergy Appl Immunol* 1981;65:460–4.
15. Chung WH, Hung SI, Hong HS, Hsieh MS, Yang LC, Ho HC, et al. Medical genetics: a marker for Stevens-Johnson syndrome. *Nature* 2004;428:486.
16. Hung SI, Chung WH, Liou LB, Chu CC, Lin M, Huang HP, et al. HLA-B*5801 allele as a genetic marker for severe cutaneous adverse reactions caused by allopurinol. *Proc Natl Acad Sci USA* 2005;102:4134–9.
17. Saag M, Balu R, Phillips E, Brachman P, Martorell C, Burman W, et al. High sensitivity of human leukocyte antigen-b*5701 as a marker for immunologically confirmed abacavir hypersensitivity in white and black patients. *Clin Infect Dis* 2008;46:1111–8.
18. Aihara M. Pharmacogenetics of cutaneous adverse drug reactions. *J Dermatol* 2011;38:246–54.
19. Niihara H, Kakamu T, Fujita Y, Kaneko S, Morita E. HLA-A31 strongly associates with carbamazepine-induced adverse drug reactions but not with carbamazepine-induced lymphocyte proliferation in a Japanese population. *J Dermatol* 2012;39:594–601.
20. Ozeki T, Mushiroda T, Yowang A, Takahashi A, Kubo M, Shirakata Y, et al. Genome-wide association study identifies HLA-A*3101 allele as a genetic risk factor for carbamazepine-induced cutaneous adverse drug reactions in Japanese population. *Hum Mol Genet* 2011;20:1034–41.
21. Genin E, Chen DP, Hung SI, Sekula P, Schumacher M, Chang PY, et al. HLA-A*31:01 and different types of carbamazepine-induced severe cutaneous adverse reactions: an international study and meta-analysis. *Pharmacogenomics J* 2014;14:281–8.
22. Amstutz U, Ross CJ, Castro-Pastrana LI, Rieder MJ, Shear NH, Hayden MR, et al. HLA-A 31:01 and HLA-B 15:02 as genetic markers for carbamazepine hypersensitivity in children. *Clin Pharmacol Ther* 2013;94:142–9.
23. McCormack M, Alfirevic A, Bourgeois S, Farrell JJ, Kasperavičiūtė D, Carrington M, et al. HLA-A*3101 and carbamazepine-induced hypersensitivity reactions in Europeans. *N Engl J Med* 2011;364:1134–43.
24. Fricke-Galindo I, Martínez-Juárez IE, Monroy-Jaramillo N, Jung-Cook H, Falfán-Valencia R, Ortega-Vázquez A, et al. HLA-A*02:01:01/-B*35:01:01/-C*04:01:01 haplotype associated with lamotrigine-induced maculopapular exanthema in Mexican mestizo patients. *Pharmacogenomics* 2014;15:1881–91.
25. Li LJ, Hu FY, Wu XT, An DM, Yan B, Zhou D. Predictive markers for carbamazepine and lamotrigine-induced maculopapular exanthema in Han Chinese. *Epilepsy Res* 2013;106:296–300.
26. Hsiao Y, Hui R, Wu T, Chang W, Hsieh M, Yang C, et al. Genotype-phenotype association between HLA and carbamazepine-induced hypersensitivity reactions: strength and clinical correlations. *J Dermatol Sci* 2014;73:101–9.
27. Hung SI, Chung WH, Jee SH, Chen WC, Chang YT, Lee WR, et al. Genetic susceptibility to carbamazepine-induced cutaneous adverse drug reactions. *Pharmacogenet Genomics* 2006;16:297–306.
28. Kashiwagi M, Aihara M, Takahashi Y, Yamazaki E, Yamane Y, Song Y, et al. Human leukocyte antigen genotypes in carbamazepine-induced severe cutaneous adverse drug response in Japanese patients. *J Dermatol* 2008;35:683–5.
29. Then SM, Rani ZZ, Raymond AA, Ratnaningrum S, Jamal R. Frequency of the HLA-B*1502 allele contributing to carbamazepine-induced hypersensitivity reactions in a cohort of Malaysian epilepsy patients. *Asian Pac J Allergy Immunol* 2011;29:290–3.
30. Kim SH, Lee KW, Song WJ, Kim SH, Jee YK, Lee SM, et al. Carbamazepine-induced severe cutaneous adverse reactions and HLA genotypes in Koreans. *Epilepsy Res* 2011;97:190–7.
31. Mehta TY, Prajapati LM, Mittal B, Joshi CG, Sheth JJ, Patel DB, et al. Association of HLA-B*1502 allele and carbamazepine-induced Stevens-Johnson syndrome among Indians. *Indian J Dermatol Venereol Leprol* 2009;75:579–82.
32. Lochareonkul C, Loplumert J, Limotai C, Korkij W, Desudchit T, Tongkobpetch S, et al. Carbamazepine and phenytoin induced Stevens-Johnson syndrome is associated with HLA-B*1502 allele in Thai population. *Epilepsia* 2008;49:2087–91.
33. Nguyen DV, Chu HC, Nguyen DV, Phan MH, Craig T, Baumgart K, et al. HLA-B*1502 and carbamazepine-induced severe cutaneous adverse drug reactions in Vietnamese. *Asia Pac Allergy* 2015;5:68–77.
34. Kwan PK, Ng MH, Lo SV. Association between HLA-B*15:02 allele and antiepileptic drug-induced severe cutaneous reactions in Hong Kong Chinese: a population-based study. *Hongkong Med J* 2014;20:16–8.
35. Aggarwal R, Sharma M, Modi M, Garg VK, Salaria M. HLA-B*1502 is associated with carbamazepine induced Stevens-Johnson syndrome in North Indian population. *Hum Immunol* 2014;75:1120–2.
36. He XJ, Jian LY, He XL, Wu Y, Xu YY, Sun XJ, et al. Association between the HLA-B 15:02 allele and carbamazepine-induced Stevens-Johnson syndrome/toxic epidermal necrolysis in Han individuals of northeastern China. *Pharmacol Rep* 2013;65:1256–62.
37. Toh DS, Tan LL, Aw DC, Pang SM, Lim SH, Thirumoorthy T, et al. Building pharmacogenetics into a pharmacovigilance program

- in Singapore: using serious skin rash as a pilot study. *Pharmacogenomics J* 2014;14:316–21.
38. Chong KW, Chan DW, Cheung YB, Ching LK, Hie SL, Thomas T, et al. Association of carbamazepine-induced severe cutaneous drug reactions and HLA-B*1502 allele status, and dose and treatment duration in paediatric neurology patients in Singapore. *Arch Dis Child* 2014;99:581–4.
 39. Cheung YK, Cheng SH, Chan EJ, Lo SV, Ng MH, Kwan P. HLA-B alleles associated with severe cutaneous reactions to antiepileptic drugs in Han Chinese. *Epilepsia* 2013;54:1307–14.
 40. Ueta M, Tokunaga K, Sotozono C, Sawai H, Tamiya G, Inatomi T, et al. HLA-A*0206 with TLR3 polymorphisms exerts more than additive effects in Stevens-Johnson syndrome with severe ocular surface complications. *PLoS One* 2012;7:1–7.
 41. Shi YW, Min FL, Qin B, Zou X, Liu XR, Gao MM, et al. Association between HLA and Stevens-Johnson syndrome induced by carbamazepine in southern Han Chinese: genetic markers besides B*1502? *Basic Clin Pharmacol Toxicol* 2012;111:58–64.
 42. Kulkantrakorn K, Tassaneeyakul W, Tiamkao S, Jantararoungtong T, Prabmechai N, Vannaprasaht S, et al. HLA-B*1502 strongly predicts carbamazepine-induced Stevens-Johnson syndrome and toxic epidermal necrolysis in Thai patients with neuropathic pain. *Pain Pract* 2012;12:202–8.
 43. Zhang Y, Wang J, Zhao LM, Peng W, Shen GQ, Xue L, et al. Strong association between HLA-B*1502 and carbamazepine-induced Stevens-Johnson syndrome and toxic epidermal necrolysis in mainland Han Chinese patients. *Eur J Clin Pharmacol* 2011;67:885–7.
 44. Wang Q, Zhou JQ, Zhou LM, Chen ZY, Fang ZY, Chen SD, et al. Association between HLA-B*1502 allele and carbamazepine-induced severe cutaneous adverse reactions in Han people of southern China mainland. *Seizure* 2011;20:446–8.
 45. Chang CC, Too CL, Murad S, Hussein SH. Association of HLA-B1502 allele with carbamazepine-induced toxic epidermal necrolysis and Stevens-Johnson syndrome in the multi-ethnic Malaysian population. *Int J Dermatol* 2011;50:221–4.
 46. Kaniwa N, Saito Y, Aihara M, Matsunaga K, Tohkin M, Kurose K, et al. HLA-B*1511 is a risk factor for carbamazepine-induced Stevens-Johnson syndrome and toxic epidermal necrolysis in Japanese patients. *Epilepsia* 2010;51:2461–5.
 47. Wu XT, Hu FY, An DM, Yan B, Jiang X, Kwan P, et al. Association between carbamazepine-induced cutaneous adverse drug reactions and the HLA-B*1502 allele among patients in central China. *Epilepsy Behav* 2010;19:405–8.
 48. Tassaneeyakul W, Tiamkao S, Jantararoungtong T, Chen P, Lin SY, Chen WH, et al. Association between HLA-B*1502 and carbamazepine-induced severe cutaneous adverse drug reactions in a Thai population. *Epilepsia* 2010;51:926–30.
 49. Man CB, Kwan P, Baum L, Yu E, Lau KM, Cheng AS, et al. Association between HLA-B*1502 allele and antiepileptic drug-induced cutaneous reactions in Han Chinese. *Epilepsia* 2007;48:1015–8.
 50. Wang W, Hu FY, Wu XT, An DM, Yan B, Zhou D. Genetic predictors of Stevens-Johnson syndrome and toxic epidermal necrolysis induced by aromatic antiepileptic drugs among the Chinese Han population. *Epilepsy Behav* 2014;37:16–9.
 51. Shirzadi M, Thorstensen K, Helde G, Moen T, Brodtkorb E. Do HLA-A markers predict skin-reactions from aromatic antiepileptic drugs in a Norwegian population? A case control study. *Epilepsy Res* 2015;118:5–9.
 52. Ito A, Shimada H, Ishikawa K, Takeo N, Hatano Y, Katagiri K, et al. Association between HLA-DRB1*0405, -DQB1*0401 and -DQA1*0303 alleles and lamotrigine-induced cutaneous adverse drug reactions. A pilot case-control study from Japan. *J Affect Disord* 2015;179:47–50.
 53. Moon J, Park HK, Chu K, Sunwoo JS, Byun JI, Lim JA, et al. The HLA-A*2402/Cw*0102 haplotype is associated with lamotrigine-induced maculopapular eruption in the Korean population. *Epilepsia* 2015;56:e161–7.
 54. Kazeem GR, Cox C, Aponte J, Messenheimer J, Brazell C, Nelsen AC, et al. High-resolution HLA genotyping and severe cutaneous adverse reactions in lamotrigine-treated patients. *Pharmacogenet Genomics* 2009;19:661–5.
 55. Park HJ, Kim SR, Leem DW, Moon IJ, Koh BS, Park KH, et al. Clinical features of and genetic predisposition to drug-induced Stevens-Johnson syndrome and toxic epidermal necrolysis in a single Korean tertiary institution patients – investigating the relation between the HLA-B*4403 allele and lamotrigine. *Eur J Clin Pharmacol* 2015;71:35–41.
 56. Lv YD, Min FL, Liao WP, He N, Zeng T, Ma DH, et al. The association between oxcarbazepine-induced maculopapular eruption and HLA-B alleles in a northern Han Chinese population. *BMC Neurol* 2013;13:75.
 57. He N, Min FL, Shi YW, Guo J, Liu XR, Li BM, et al. Cutaneous reactions induced by oxcarbazepine in southern Han Chinese: incidence, features, risk factors and relation to HLA-B alleles. *Seizure* 2012;21:614–8.
 58. Hu FY, Wu XT, An DM, Yan B, Stefan H, Zhou D. Pilot association study of oxcarbazepine-induced mild cutaneous adverse reactions with HLA-B*1502 allele in Chinese Han population. *Seizure* 2011;20:160–2.
 59. Hung S, Chung W, Liu Z, Chen C, Hsieh M, Hui RC, et al. Common risk allele in aromatic antiepileptic-drug induced Stevens-Johnson syndrome and toxic epidermal necrolysis in Han Chinese. *Pharmacogenomics* 2010;11:349–56.
 60. Kaniwa N, Sugiyama E, Saito Y, Kurose K, Maekawa K, Hasegawa R, et al. Specific HLA types are associated with antiepileptic drug-induced Stevens-Johnson syndrome and toxic epidermal necrolysis in Japanese subjects. *Pharmacogenomics* 2013;14:1821–31.
 61. Chang CC, Ng CC, Too CL, Choon SE, Lee CK, Chung WH, et al. Association of HLA-B*15:13 and HLA-B*15:02 with phenytoin-induced severe cutaneous adverse reactions in a Malay population. *Pharmacogenomics J* 2016;1–4. [Epub ahead of print].
 62. Niihara H, Kaneko S, Ito T, Sugamori T, Takahashi N, Kohno K, et al. HLA-B 58:01 strongly associates with allopurinol-induced adverse drug reactions in a Japanese sample population. *J Dermatol Sci* 2013;71:150–2.
 63. Cao Z, Wei Z, Zhu Q, Zhang J, Yang L, Qin S, et al. HLA-B*58:01 allele is associated with augmented risk for both mild and severe cutaneous adverse reactions induced by allopurinol in Han Chinese. *Pharmacogenomics* 2012;13:1193–201.
 64. Ng CY, Yeh YT, Wang CW, Hung SI, Yang CH, Chang YC, et al. Impact of the HLA-B(*)58:01 allele and renal impairment on allopurinol-induced cutaneous adverse reactions. *J Invest Dermatol* 2016;136:1373–81.
 65. Cheng L, Xiong Y, Qin CZ, Zhang W, Chen XP, Li J, et al. HLA-B*58:01 is strongly associated with allopurinol-induced severe cutaneous adverse reactions in Han Chinese patients: a multicentre retrospective case-control clinical study. *Br J Dermatol* 2015;173:555–8.

66. Zhang X, Ma H, Hu C, Yu B, Ma W, Wu Z, et al. Detection of HLA-B*58:01 with TaqMan assay and its association with allopurinol-induced sCADR. *Clin Chem Lab Med* 2015;53:383–90.
67. Gonçalves M, Coutinho I, Teixeira V, Gameiro AR, Brites MM, Nunes R, et al. HLA-B*58:01 is a risk factor for allopurinol-induced DRESS and Stevens-Johnson syndrome/toxic epidermal necrolysis in a Portuguese population. *Br J Dermatol* 2013;169:660–5.
68. Chiu ML, Hu M, Ng MH, Yeung CK, Chan JC, Chang MM, et al. Association between HLA-B*58:01 allele and severe cutaneous adverse reactions with allopurinol in Han Chinese in Hong Kong. *Br J Dermatol* 2012;167:44–9.
69. Kang HR, Jee YK, Kim YS, Lee CH, Jung JW, Kim SH, et al. Positive and negative associations of HLA class I alleles with allopurinol-induced SCARs in Koreans. *Pharmacogenet Genomics* 2011;21:303–7.
70. Cristallo AF, Schroeder J, Citterio A, Santori G, Ferrioli GM, Rossi U, et al. A study of HLA class I and class II 4-digit allele level in Stevens-Johnson syndrome and toxic epidermal necrolysis. *Int J Immunogenet* 2011;38:303–9.
71. Park DJ, Kang JH, Lee JW, Lee KE, Wen L, Kim TJ, et al. Cost-effectiveness analysis of HLA-B5801 genotyping in the treatment of gout patients with chronic renal insufficiency in Korea. *Arthritis Care Res* 2015;67:280–7.
72. Tassaneeyakul W, Jantararoungtong T, Chen P, Lin PY, Tiamkao S, Khunarkornsiri U, et al. Strong association between HLA-B*5801 and allopurinol-induced Stevens-Johnson syndrome and toxic epidermal necrolysis in a Thai population. *Pharmacogenet Genomics* 2009;19:704–9.
73. Kaniwa N, Saito Y, Aihara M, Matsunaga K, Tohkin M, Kurose K, et al. HLA-B locus in Japanese patients with anti-epileptics and allopurinol-related Stevens-Johnson syndrome and toxic epidermal necrolysis. *Pharmacogenomics* 2008;9:1617–22.
74. Lonjou C, Borot N, Sekula P, Ledger N, Thomas L, Halevy S, et al. A European study of HLA-B in Stevens-Johnson syndrome and toxic epidermal necrolysis related to five high-risk drugs. *Pharmacogenet Genomics* 2008;18:99–107.
75. Berka N, Gill JM, Liacini A, O'Bryan T, Khan FM. Human leukocyte antigen (HLA) and pharmacogenetics: screening for HLA-B*57:01 among human immunodeficiency virus-positive patients from southern Alberta. *Hum Immunol* 2012;73:164–7.
76. Phillips EJ, Wong GA, Kaul R, Shahabi K, Nolan DA, Knowles SR, et al. Clinical and immunogenetic correlates of abacavir hypersensitivity. *AIDS* 2005;19:979–81.
77. Martin AM, Nolan D, Gaudieri S, Almeida CA, Nolan R, James I, et al. Predisposition to abacavir hypersensitivity conferred by HLA-B*5701 and a haplotypic Hsp70-Hom variant. *Proc Natl Acad Sci U S A* 2004;101:4180–5.
78. Hetherington S, Hughes AR, Mosteller M, Shortino D, Baker KL, Spreen W, et al. Genetic variations in HLA-B region and hypersensitivity reactions to abacavir. *Lancet* 2002;359:1121–2.
79. Mallal S, Nolan D, Witt C, Masel G, Martin AM, Moore C, et al. Association between presence of HLA-B*5701, HLA-DR7, and HLA-DQ3 and hypersensitivity to HIV-1 reverse-transcriptase inhibitor abacavir. *Lancet* 2002;359:727–32.
80. Chantarangsou S, Mushiroad T, Mahasirimongkol S, Kiertiburanakul S, Sungkanuparph S, Manosuthi W, et al. HLA-B*3505 allele is a strong predictor for nevirapine-induced skin adverse drug reactions in HIV-infected Thai patients. *Pharmacogenet Genomics* 2009;19:139–46.
81. Phillips E, Bartlett JA, Sanne I, Lederman MM, Hinkle J, Rousseau F, et al. Associations between HLA-DRB1*0102, HLA-B*5801, and hepatotoxicity during initiation of nevirapine-containing regimens in South Africa. *J Acquir Immune Defic Syndr* 2013;62:55–7.
82. Gao S, Gui XE, Liang K, Liu Z, Hu J, Dong B. HLA-dependent hypersensitivity reaction to nevirapine in Chinese Han HIV-infected patients. *AIDS Res Hum Retroviruses* 2012;28:540–3.
83. Carr DF, Chaponda M, Jorgensen AL, Castro EC, Van Oosterhout JJ, Khoo SH, et al. Association of human leukocyte antigen alleles and nevirapine hypersensitivity in a Malawian HIV-infected population. *Clin Infect Dis* 2013;56:1330–9.
84. Vitezica ZG, Milpied B, Lonjou C, Borot N, Ledger TN, Lefebvre A, et al. HLA-DRB1*01 associated with cutaneous hypersensitivity induced by nevirapine and efavirenz. *AIDS* 2008;22:540–1.
85. Stephens C, López-Nevot MÁ, Ruiz-Cabello F, Ulzurrun E, Soriano G, Romero-Gómez M, et al. HLA alleles influence the clinical signature of amoxicillin-clavulanate hepatotoxicity. *PLoS One* 2013;8:e68111.
86. Donaldson PT, Daly AK, Henderson J, Graham J, Pirmohamed M, Bernal W, et al. Human leucocyte antigen class II genotype in susceptibility and resistance to co-amoxiclav-induced liver injury. *J Hepatol* 2010;53:1049–53.
87. Hautekeete ML, Horsmans Y, Van Waeyenberge C, Demanet C, Henrion J, Verbist L, et al. HLA association of amoxicillin-clavulanate-induced hepatitis. *Gastroenterology* 1999;117:1181–6.
88. Chen R, Zhang Y, Tang S, Lv X, Wu S, Sun F, et al. The association between HLA-DQB1 polymorphism and antituberculosis drug-induced liver injury: a case-control study. *J Clin Pharm Ther* 2015;40:110–5.
89. Sharma SK, Balamurugan A, Saha PK, Pandey RM, Mehra NK. Evaluation of clinical and immunogenetic risk factors for the development of hepatotoxicity during antituberculosis treatment. *Am J Respir Crit Care Med* 2002;166:916–9.
90. Kim SH, Lee SK, Kim SH, Park HW, Chang YS, Lee KW, et al. Antituberculosis drug-induced hypersensitivity syndrome and its association with human leukocyte antigen. *Tuberculosis (Edinb)* 2013;93:270–4.
91. Fernandez CA, Smith C, Yang W, Dat M, Bashford D, Larsen E, et al. HLA-DRB1*07:01 is associated with a higher risk of asparaginase allergies. *Blood* 2014;124:1266–76.
92. Esmailzadeh H, Nabavi M, Aryan Z, Akbar A. Pharmacogenetic tests to predict the efficacy of aspirin desensitization in patients with aspirin-exacerbated respiratory diseases; HLA-DQB302. *Expert Rev Respir Med* 2015;9:511–8.
93. Esmailzadeh H, Nabavi M, Amirzargar AA, Aryan Z, Arshi S, Bermanian MH, et al. HLA-DRB and HLA-DQ genetic variability in patients with aspirin-exacerbated respiratory disease. *Am J Rhinol Allergy* 2015;29:63–9.
94. Salvador F, Sánchez-Montalvá A, Martínez-Gallo M, Sala-Cunill A, Viñas L, García-Prat M, et al. Evaluation of cytokine profile and HLA association in benzimidazole related cutaneous reactions in patients with chagas disease. *Clin Infect Dis* 2015;61:1688–94.
95. Furukawa H, Oka S, Shimada K, Sugii S, Hashimoto A, Komiya A, et al. HLA-DRB1*08:02 is associated with bucillamine-induced proteinuria in Japanese rheumatoid arthritis patients. *Biomark Insights* 2014;9:23–8.
96. Dettling M, Cascorbi I, Opgen-Rhein C, Schaub R. Clozapine-induced agranulocytosis in schizophrenic Caucasians: confirming clues for associations with human leukocyte class I and II antigens. *Pharmacogenomics J* 2007;7:325–32.

97. Corzo D, Yunis JJ, Salazar M, Lieberman JA, Howard A, Awdeh Z, et al. The major histocompatibility complex region marked by HSP70-1 and HSP70-2 variants is associated with clozapine-induced agranulocytosis in two different ethnic groups. *Blood* 1995;86:3835–40.
98. Ueta M, Kannabiran C, Wakamatsu TH, Kim MK, Yoon KC, Seo KY, et al. Trans-ethnic study confirmed independent associations of HLA-A*02:06 and HLA-B*44:03 with cold medicine-related Stevens-Johnson syndrome with severe ocular surface complications. *Sci Rep* 2014;4:5981.
99. Kongpan T, Mahasirimongkol S, Konyoung P, Kanjanawart S, Chumworathayi P, Wichukchinda N, et al. Candidate HLA genes for prediction of co-trimoxazole-induced severe cutaneous reactions. *Pharmacogenet Genomics* 2015;25:402–11.
100. Zhang FR, Liu H, Irwanto A, Fu XA, Li Y, Yu GQ, et al. HLA-B*13:01 and the dapsone hypersensitivity syndrome. *N Engl J Med* 2013;369:1620–8.
101. Wang H, Yan L, Zhang G, Chen X, Yang J, Li M, et al. Association between HLA-B*1301 and dapsone-induced hypersensitivity reactions among leprosy patients in China. *J Invest Dermatol* 2013;133:2642–4.
102. Schaid DJ, Spraggs CF, McDonnell SK, Parham LR, Cox CJ, Ejlersen B, et al. Prospective validation of HLA-DRB1*07:01 allele carriage as a predictive risk factor for lapatinib-induced liver injury. *J Clin Oncol* 2014;32:2296–303.
103. Yang F, Xuan J, Chen J, Zhong H, Luo H, Zhou P, et al. HLA-B*59:01: a marker for Stevens-Johnson syndrome/toxic epidermal necrolysis caused by methazolamide in Han Chinese. *Pharmacogenomics J* 2016;16:83–7.
104. Kim SH, Kim M, Lee KW, Kim SH, Kang HR, Park HW, et al. HLA-B*5901 is strongly associated with methazolamide-induced Stevens-Johnson syndrome/toxic epidermal necrolysis. *Pharmacogenomics* 2010;11:879–84.
105. Ueta M, Sotozono C, Tokunaga K, Yabe T, Kinoshita S. Strong association between HLA-A*0206 and Stevens-Johnson syndrome in the Japanese. *Am J Ophthalmol* 2007;143:367–8.
106. Quirarte J, Sánchez-García F, Torres MJ, Blanco C, Castillo R, Ortega N, et al. Association of HLA-DR11 with the anaphylactoid reaction caused by nonsteroidal anti-inflammatory drugs. *J Allergy Clin Immunol* 1999;103:685–9.
107. Yang J, Qiao H, Zhang Y, Jia L, Tian X, Gao N. HLA-DRB genotype and specific IgE responses in patients with allergies to penicillins. *Chin Med J (Engl)* 2006;119:458–66.
108. Yang F, Gu B, Zhang L, Xuan J, Luo H, Zhou P, et al. HLA-B*13:01 is associated with salazosulfapyridine-induced drug rash with eosinophilia and systemic symptoms in Chinese Han population. *Pharmacogenomics* 2014;15:1461–9.
109. Pickl WF, Fischer GF, Fad I, Kolarz G, Scherak O. HLA-DR1-positive patients suffering from rheumatoid arthritis are at high risk for developing mucocutaneous side effects upon gold therapy. *Hum Immunol* 1993;38:127–31.
110. Gunnarsson I, Nordmark B, Hassan Bakri A, Gröndal G, Larsson P, Forslid J, et al. Development of lupus-related side-effects in patients with early RA during sulphasalazine treatment – the role of IL-10 and HLA. *Rheumatology (Oxford)* 2000;39:886–93.
111. Hirata K, Takagi H, Yamamoto M, Matsumoto T, Nishiya T, Mori K, et al. Ticlopidine-induced hepatotoxicity is associated with specific human leukocyte antigen genomic subtypes in Japanese patients: a preliminary case-control study. *Pharmacogenomics J* 2008;8:29–33.
112. Ferrell PB, McLeod HL. Carbamazepine, HLA-B*1502 and risk of Stevens-Johnson syndrome and toxic epidermal necrolysis: US FDA recommendations. *Pharmacogenomics* 2008;9:1543–6.
113. Lim KS, Kwan P, Tan CT. Association of HLA-B * 1502 allele and carbamazepine-induced severe adverse cutaneous drug reaction among Asians, a review. *Neurol Asia* 2008;13:15–21.
114. Lonjou C, Thomas L, Borot N, Ledger N, de Toma C, LeLouet H, et al. A marker for Stevens-Johnson syndrome ethnicity matters. *Pharmacogenomics J* 2006;6:265–8.
115. González-Galarza FF, Takeshita LY, Santos EJ, Kempson F, Maia MH, Da Silva AL, et al. Allele frequency net 2015 update: new features for HLA epitopes, KIR and disease and HLA adverse drug reaction associations. *Nucleic Acids Res* 2015;43:D784–8.
116. Du T, Yang L, Luo H, Xuan J, Xing Q, Ning B, et al. HLADR: a database system for enhancing the discovery of biomarkers for predicting human leukocyte antigen-mediated idiosyncratic adverse drug reactions. *Biomark Med* 2015;9:1079–93.
117. Gonzalez-Galarza FF, Christmas S, Middleton D, Jones AR. Allele frequency net: a database and online repository for immune gene frequencies in worldwide populations. *Nucleic Acids Res* 2011;39:913–9.
118. Middleton D, Menchaca L, Rood H, Komerofsky R. New allele frequency database: <http://www.allelefrequencies.net>. *Tissue Antigens* 2003;61:403–7.
119. Ehmann F, Caneva L, Prasad K, Paulmichl M, Maliepaard M, Llerena A, et al. Pharmacogenomic information in drug labels: European Medicines Agency perspective. *Pharmacogenomics J* 2015;15:201–10.
120. Caudle KE, Klein TE, Hoffman JM, Muller DJ, Whirl-Carrillo M, Gong L, et al. Incorporation of pharmacogenomics into routine clinical practice: the Clinical Pharmacogenetics Implementation Consortium (CPIC) guideline development process. *Curr Drug Metab* 2014;15:209–17.
121. Relling MV, Klein TE. CPIC: clinical pharmacogenetics implementation consortium of the pharmacogenomics research network. *Clin Pharmacol Ther* 2011;89:464–7.
122. Martin MA, Hoffman JM, Freimuth RR, Klein TE, Dong BJ, Pirmohamed M, et al. Clinical pharmacogenetics implementation consortium guidelines for HLA-B genotype and Abacavir dosing: 2014 update. *Clin Pharmacol Ther* 2014;95:499–500.
123. Martin MA, Klein TE, Dong BJ, Pirmohamed M, Haas DW, Kroetz DL, et al. Clinical pharmacogenetics implementation consortium guidelines for HLA-B genotype and abacavir dosing. *Clin Pharmacol Ther* 2012;91:734–8.
124. Hershfield MS, Callaghan JT, Tassaneeyakul W, Mushiroda T, Thorn CF, Klein TE, et al. Clinical pharmacogenetics implementation consortium guidelines for human leukocyte antigen-B genotype and allopurinol dosing. *Clin Pharmacol Ther* 2013;93:153–8.
125. Saito Y, Stamp LK, Caudle KE, Hershfield MS, McDonagh EM, Callaghan JT, et al. Clinical pharmacogenetics implementation consortium (CPIC) guidelines for human leukocyte antigen B (HLA-B) genotype and allopurinol dosing: 2015 update. *Clin Pharmacol Ther* 2016;99:36–7.
126. Caudle KE, Rettie AE, Whirl-Carrillo M, Smith LH, Mintzer S, Lee MT, et al. Clinical pharmacogenetics implementation consortium guidelines for CYP2C9 and HLA-B genotypes and phenytoin dosing. *Clin Pharmacol Ther* 2014;96:542–8.

127. Leckband SG, Kelsoe JR, Dunnenberger HM, George AL, Tran E, Berger R, et al. Clinical pharmacogenetics implementation consortium guidelines for HLA-B genotype and carbamazepine dosing. *Clin Pharmacol Ther* 2013;94:324–8.
128. FDA. Genomics – table of pharmacogenomic biomarkers in drug labeling [Internet]. Center for Drug Evaluation and Research. Food and Drug Administration. Available at: <http://www.fda.gov/drugs/scienceresearch/researchareas/pharmacogenetics/ucm083378.htm>. Accessed: 16 June 2015.
129. European Medicines Agency. Ziagen (abacavir) [Internet]. Available at: http://www.ema.europa.eu/ema/index.jsp?curl=pages/medicines/human/medicines/000252/human_med_001179.jsp&mid=WC0b01ac058001d124. Accessed: 21 July 2016.
130. European Medicines Agency. PhVWP monthly report on safety concerns, guidelines and general matters. Available at: http://www.ema.europa.eu/docs/en_GB/document_library/Report/2012/04/WC500124972.pdf. Accessed on 1 March, 2017.
131. Bushyakanist A, Puangpetch A, Sukasem C, Kiertiburanakul S. The use of pharmacogenetics in clinical practice for the treatment of individuals with HIV infection in Thailand. *Pharmacogenomics Pers Med* 2015;8:163–70.
132. Powell G, Holmes EA, Plumpton CO, Ring A, Baker GA, Jacoby A, et al. Pharmacogenetic testing prior to carbamazepine treatment of epilepsy: patients' and physicians' preferences for testing and service delivery. *Br J Clin Pharmacol* 2015;80:1149–59.
133. Dong D, Sung C, Finkelstein EA. Cost-effectiveness of HLA-B*1502 genotyping in adult patients with newly diagnosed epilepsy in Singapore. *Neurology* 2012;79:1259–67.
134. Tiamkao S, Jitpimolmard J, Sawanyawisuth K, Jitpimolmard S. Cost minimization of HLA-B*1502 screening before prescribing carbamazepine in Thailand. *Int J Clin Pharm* 2013;35:608–12.
135. Plumpton CO, Yip VL, Alfirevic A, Marson AG, Pirmohamed M, Hughes DA. Cost-effectiveness of screening for HLA-A*31:01 prior to initiation of carbamazepine in epilepsy. *Epilepsia* 2015;56:556–63.
136. Saokaew S, Tassaneeyakul W, Maenthaisong R, Chaiyakunapruk N. Cost-effectiveness analysis of HLA-B*5801 testing in preventing allopurinol-induced SJS/TEN in Thai population. *PLoS One* 2014;9:1–9.
137. Nieves Calatrava D, Calle-Martín Ode L, Iribarren-Loyarte J, Rivero-Román A, García-Bujalance L, Pérez-Escolano I, et al. Cost-effectiveness analysis of HLA-B*5701 typing in the prevention of hypersensitivity to abacavir in HIV+ patients in Spain. *Enferm Infecc Microbiol Clin* 2010;28:590–5.
138. Wolf E, Blankenburg M, Bogner JR, Becker W, Gorriahn D, Mueller MC, et al. Cost impact of prospective HLA-B*5701-screening prior to abacavir/lamivudine fixed dose combination use in Germany. *Eur J Med Res* 2010;15:145–51.
139. Plumpton CO, Roberts D, Pirmohamed M, Hughes DA. A systematic review of economic evaluations of pharmacogenetic testing for prevention of adverse drug reactions. *Pharmacoeconomics* 2016;34:771–93.
140. Thorstensen K, Kvitland M, Shirzadi M, Helde G, Moen T, Brodtkorb E. Carbamazepine-induced cutaneous reactions: a simple assay to identify patients carrying the HLA-A*31:01 allele. *Scand J Clin Lab Invest* 2014;74:644–7.
141. Uchiyama K, Kubota F, Ariyoshi N, Matsumoto J, Ishii I, Kitada M. Development of a simple method for detection of the HLA-A*31:01 allele. *Drug Metab Pharmacokinet* 2013;28:435–8.
142. De Spiegelaere W, Philippé J, Vervisch K, Verhofstede C, Malatinkova E, Kiselina M, et al. Comparison of methods for in-house screening of HLA-B*57:01 to prevent abacavir hypersensitivity in HIV-1 care. *PLoS One* 2015;10:1–10.
143. Dello Russo C, Lisi L, Fabbiani M, Gagliardi D, Fanti I, Di Giambenedetto S, et al. Detection of HLA-B*57:01 by real-time PCR: implementation into routine clinical practice and additional validation data. *Pharmacogenomics* 2014;15:319–27.
144. Scarsi M, Bosio C, Coccoli S, Barucco A, Tavelli G, Airò P. Flow cytometry test to screen for HLA-B*58:01-associated allopurinol hypersensitivity. *Clin Rheumatol* 2014;33:873–5.
145. Borgiani P, Di Fusco D, Erba F, Marazzi MC, Mancinelli S, Novelli G, et al. HCP5 genetic variant (RS3099844) contributes to nevirapine-induced Stevens Johnsons syndrome/toxic epidermal necrolysis susceptibility in a population from Mozambique. *Eur J Clin Pharmacol* 2014;70:275–8.
146. Vidal C, Li J, Fulton R, Fernando SL. A polymorphism within the psoriasis susceptibility 1 candidate 1 (PSORS1C1) gene is not linked to HLA-B*58:01 in an Australian cohort. *Drug Metab Pharmacokinet* 2016;31:252–5.
147. Dalbeth N, Stamp L. Allopurinol dosing in renal impairment: walking the tightrope between adequate urate lowering and adverse events. *Semin Dial* 2007;20:391–5.
148. Daly AK. Pharmacogenomics of adverse drug reactions. *Genome Med* 2013;5:5.
149. Karlin E, Phillips E. Genotyping for severe drug hypersensitivity. *Curr Allergy Asthma Rep* 2014;14:418.