

Research Article

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Fractionating microplastics by density gradient centrifugation: a novel approach using LuerLock syringes in a low-cost density gradient maker

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Abstract: Microplastics are now ubiquitous in the environment and are even considered “technofossils” of the Anthropocene. Given their omnipresence and potential impact, identifying and analyzing these particles becomes increasingly crucial. Novel approaches suggest density gradient centrifugation for simultaneous extraction and fractionation of microplastic particles based on their plastic-specific densities. In this article we describe a cheap and harmless experimental setting to fractionate microplastic particles by density gradient centrifugation. An innovative low-cost Do-It-Yourself (DIY) gradient maker using Luer-Lock syringes is presented. With this gradient maker it is possible to produce density gradients with water and sucrose solutions, covering a density range of 1.00–1.32 g/cm³, as well as with water and saturated potassium carbonate solutions, covering a density range of 1.06–1.53 g/cm³. The separation performance was tested with the most broadly used plastics polyamide, polyurethane, polycarbonate, polyethylene terephthalate and polyvinylchloride. Both density gradients show centrifugation stability and clear banding patterns after centrifugation. Due to its cheap and easy-to-build-easy-to-use nature, this experimental setting for microplastic fractionation by density gradient centrifugation offers an approach for schools not only to address the microplastic problems, but also to integrate new methods of microplastic analysis in upper secondary school laboratories.

Keywords: microplastics; density centrifugation; low-cost; density gradient maker; qualitative analysis

1 Introduction

Plastic products are ubiquitous in our everyday lives, and with their use it is no surprise that plastic production is still increasing (Plastics Europe, 2022). As a result of human activities, macroplastics and microplastics (plastic particles smaller than 5 mm), are lost to the environment at different points across the plastic life cycle. For example, in 2015 approximately 6.2 Mt of macroplastics and 3.0 Mt of microplastics were lost to the environment (Ryberg et al., 2019). Macroplastic products that have entered the environment are usually of no use to nature.

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Instead, they are broken down into microplastic particles by external environmental influences, such as photooxidation and mechanical degradation (Andrady, 2015). Sediments are thought to be a major sink for these microplastic particles. It is therefore not surprising that microplastics have been detected in sediments worldwide, even in the deep sea (Van Cauwenberghe et al., 2013, 2015). The particles were shown to be enriched and preserved there. First plastiglomerates, molten and hardened plastic fused with other sediment, shells or other inorganic compounds were already found (Corcoran et al., 2014). Thus, plastic particles of all forms are more and more often proposed as “technofossils” of the Anthropocene for determining the stratigraphic age of core layers (Chen et al., 2022; Zalasiewicz et al., 2016). They have the potential being the “golden spike” besides radionuclides that emerged in the atmosphere since 1945 as a result of nuclear bomb tests. This makes the analysis of microplastic particles in sediments even more important for the future.

Nowadays density-based extractions with saturated salt solutions are often used to separate microplastic particles from sediments. They are based on the difference in density between plastics (0.8 g/cm^3 – 1.6 g/cm^3) and sediments ($\sim 2.7\text{ g/cm}^3$). For density separation saturated high-density salt solutions, such as sodium chloride (NaCl), sodium bromide (NaBr), sodium iodide (NaI) and zinc bromide (ZnBr_2) are often used (Prata et al., 2019; Quinn et al., 2017). A new density medium that is also suited for a school context is a saturated potassium carbonate solution (Gohla et al., 2021). For most protocols, the sediment sample is transferred to a flotation medium and shaken or stirred. After a while, the higher-density sediment grains sink to the bottom, while the microplastic particles float at the surface of the solution, from where they are removed and filtered. One limitation of this method is that it does not allow the identification of the plastic type(s) floating atop the medium. Subsequent identification is usually done only visually or by IR, Raman or GC-MS (Prata et al., 2019).

New approaches in research use density gradient centrifugation to identify different polymer types according to their specific density, but they also use highly hazardous density media such as caesium chloride and the methods usually require expensive materials (Jakobs et al., 2023; Jing et al., 2022).

In this publication we want to present an affordable and less harmful method for fractionating microplastic particles with density gradient centrifugation, suitable for educational settings. Furthermore, by replacing problematic substances, this method might serve as an example of green chemistry and can even be used by students. First, we introduce the low-cost density gradient maker. Second, cheap and harmless density media that are suitable for school use are discussed. Third, the separation performance of the gradient for selected microplastic particles is evaluated.

As this project is an example of a successful collaborative research and development approach between students, teachers and researchers, the following chapter will briefly describe the collaboration.

2 From school to university and vice versa

Originating from the course “marine sciences-oceanography” in the upper secondary level at Lise-Meitner Gymnasium in Willich-Anrath (Germany), two students took on the challenging task of microplastic analysis. Intrigued by the topic, the two students together with their teacher continued the project in a voluntary special course. Based on the principle of density separation, they optimized the method of density centrifugation for school use to identify different polymer types. The aim was to fractionate microplastic particles according to their specific density. Using only cheap materials and harmless chemicals, they wanted to develop a method that can easily be used by other students at the upper secondary or freshman level or researchers in citizen science projects. With this aim in mind, a literature review revealed that gradient makers are either heavily expensive when purchased, or there are self-built devices that cannot be easily made anymore. This is because materials are no longer manufactured or the device would need up-to-date modifications (Flurkey, 2000; Marks, 1998; Salo & Kouns, 1965).

First experiments were conducted with plastic tubes glued to a perspex disc and connected to each other. This horizontal setup was ruled out due to reoccurring mechanical issues like leakage. Afterwards, a more robust and easy-to-use vertical design was built using Luer-Lock components with minor modifications.

Subsequently first tests of density gradients using a 66 % sucrose solution were done by the two students. Microplastic particles were added for some experiments. Their promising initial results marked the start of the cooperation with the University of Graz and further development steps were discussed between students and researchers. The researchers tested the reproducibility of the gradient maker, as well as further cheap and harmless density media that are suitable for school use. Coming from school with a newly developed device, the university validated and enhanced the method with equipment that is not available to schools, and then brought the device back to school in a broadly usable way. Furthermore, the method has already been used and disseminated in teacher workshops to once again bridge the gap between research and school.

3 Materials and methods

3.1 Luer-Lock low-cost density gradient maker

The setup of the low-cost density gradient maker is simple and based on the Luer-Lock-system. This ISO-standardized medical system offers a simple option for leak-free connections through their special and accurate fittings. Normally used for medical equipment, it is increasingly used in laboratories due to its user-friendly design and compatibility with various products such as syringes, needles and tubing. To reduce costs, it can be helpful to ask hospitals or doctors for expired Luer-Lock items, which are often provided for free.

The only modification needed is piercing a stopper with a needle. All other parts used in the gradient maker are standard issue lab materials (see Figure 1).

The Luer-Lock low-cost density gradient maker is assembled vertically as shown in Figure 1. The lower syringe acts as a mixing chamber and is filled with a high-density medium. The upper syringe, filled with a low-density medium, serves as the reservoir. Due to the airtight system, the maker can produce the gradients based on the principle of pressure equalization.

For every drop leaving the lower syringe through the tube, a drop from the upper syringe enters the lower syringe via the needle, diluting the solution. Thus, the gradient maker can ensure a continuous dilution of the lower solution that leads to a continuous density distribution.

To make a gradient, the apparatus has to be prepared with all three-way-stopcocks closed. The lower syringe is filled with 5 mL of the higher density medium (e.g. a saturated potassium carbonate or sucrose solution in water). Next, the upper (bigger) syringe reservoir is filled with the lower density medium, e.g. water. A fitting stirring bar or a neodymium magnet is placed in the lower syringe, so that the liquid in the lower syringe is in motion during use and can ensure permanent mixing and consequentially

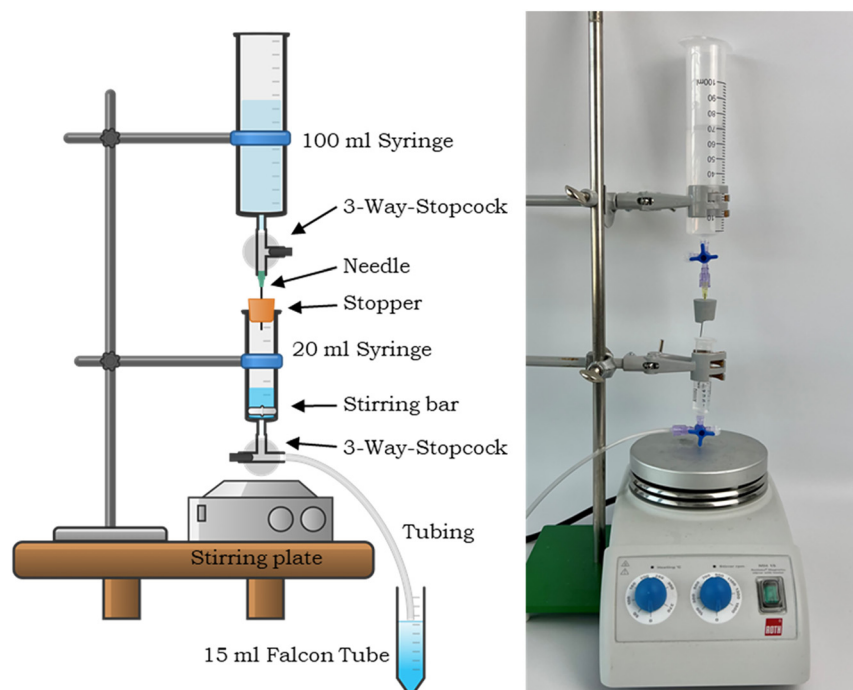


Figure 1: Setup of the Luer-Lock low-cost density gradient maker.

continuous dilution. Then it is crucial to connect the upper and lower syringe with the pierced stopper and the three-way-stopcock so that the connection is airtight.

To make a density gradient both three-way stopcocks need to be opened, allowing the media to flow through the apparatus. It is essential that this flow begins only after both three-way stopcocks have been opened. If there is any flow before this step, it indicates a lack of airtightness, making the apparatus unsuitable for use. To prevent the mixing of the gradient in the Falcon tube, it is important to raise the end of the tubing along the level of the liquid. We recommend filling the Falcon Tube with 12–13 mL of density medium so that there is still room in the tube for the sample to be analyzed.

3.2 Testing sucrose and potassium-carbonate solutions as harmless and functional density media

After it was shown that there was no leakage, the stability and reproducibility of the gradients were tested with different density media. At first a solution of 66 % sucrose in water as dense medium and water as the dilutant was tested. Thereafter, the density of each milliliter of the water-sugar gradient was determined using refractive indices and UV–VIS spectroscopy. This used methylene blue added to the sucrose solution, which was then used indirectly to determine the sucrose concentration (see Figure 3).

For further experiments, sucrose was ruled out due to insufficient maximum density (1.32 g/cm³) for separation of high-density plastics like PET and PVC. Hence a saturated potassium carbonate solution was used as a novel high-density medium (1.53 g/cm³) as suggested recently (Gohla et al., 2021). A big advantage of this setup is that the potassium carbonate solution can act as both the extraction medium for sample preparation as well as the separation medium in gradient centrifugation.

Continuity and reproducibility of the potassium-carbonate-and-water-gradient was measured 6-fold using a “DMA46” flexural vibrating meter from Anton Paar K.G. (0–3 g/cm³, accuracy $\pm 1 \times 10^{-4}$ g/cm³). The gradients were collected in a 15 mL Falcon tube and centrifuged in a SBS-LZ-400B benchtop centrifuge from Steinberg Systems, with an angular rotor at 5000 rpm for 30 min. For validation the gradients were fractionated before and after centrifugation using a pipette to take 1 mL fractions from the top of the gradient to the bottom. Then, the density of each fraction was determined.

3.3 Fractionation of microplastic particles by density gradient centrifugation

Finally, we tested the separation efficiency of gradient centrifugation to fractionate microplastic particles. We only tested plastic types with a density higher than seawater (>1.02 g/cm³), since they will sink and therefore accumulate in sediments (see Table 1) (Van Cauwenberghe et al., 2015). Plastic types with a lower density, such as low-density polyethylen (LD-PE), polypropylen (PP), high-density polyethylen (HD-PE) and expanded polystyrol (EPS) would float on seawater, hence they also float atop of the gradient and fractionation is therefore not possible with the density media used here.

The tested microplastic particles were produced by cutting, chopping or grating of everyday items.

The density gradients were then prepared as described previously. The fractionation was repeated 10-fold where each time 3–5 particles of each plastic type were placed on the density gradient. The density gradients were then centrifuged at 5000 rpm for 30 min. Afterwards, the microplastic particles were identified visually based on their shape or color.

In Figure 2, a summary of the process is shown that can also be used in schools. First, the gradient maker is used to produce a centrifugation-stable gradient. Setting up the gradient maker requires 15 min, making the gradient takes around 10 min. Second, microplastic particles are placed in the Falcon tube. After 30 min of centrifugation, the microplastic particles are fractionated and can assigned to a density range.

Table 1: Tested common plastic types and their specific density (Pacher, 2021; Quinn et al., 2017) and examples for their source.

Plastic-Type	Density [g/cm ³]	Source
Polyamide (PA)	1.13–1.15	Tights, dowels, cable ties
Polyurethane (PU)	1.05–1.28	Rolls, mattresses
Polycarbonate (PC)	1.2	CDs, glasses of safety goggles
Polyethylene terephthalate (PET)	1.38	Bottles
Plasticised polyvinyl chloride (PVC)	1.10–1.35	Cables, tubes
Unplasticised polyvinyl chloride (PVC)	1.35–1.45	Pipes, flooring

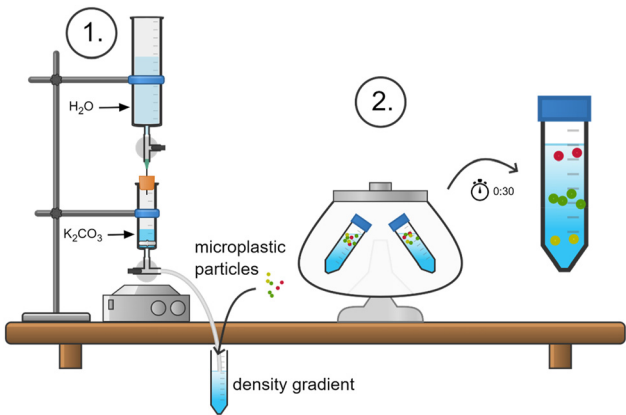


Figure 2: Overview of the two-step fractionation process of microplastic particles.

4 Results and discussion

4.1 Luer-Lock low-cost density gradient maker is suitable to produce continuous density gradients

With the Luer-Lock low-cost density gradient maker, continuous and reproducible density gradients can be produced both with a sucrose solution and water (see Figure 3) and with a saturated potassium carbonate solution and water (see Figure 4). A characteristic of a continuous gradient is its exponential decay, as can be seen in Figure 3.

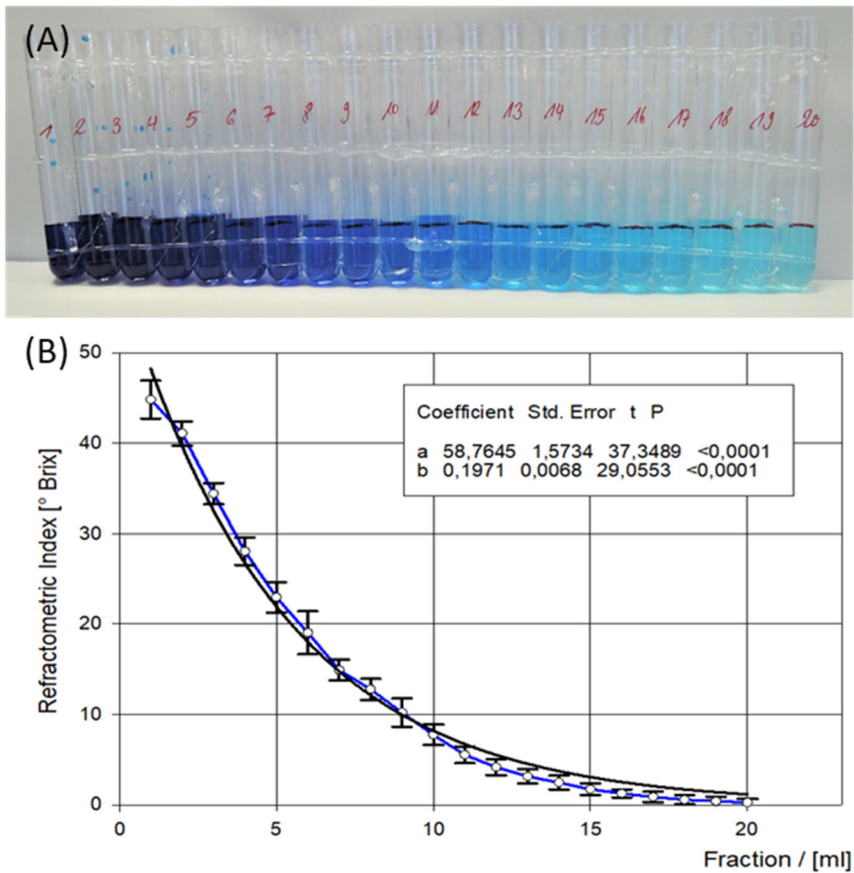


Figure 3: The sucrose – water gradient achieved by the low-cost gradient maker, separated milliliter by milliliter. (A) Methylene blue was added to the sucrose solution for visualisation (B) the refractive index for each fraction is shown as a mean of 10 measurements. The black line shows a regression for exponential decay [$f = a \cdot \exp(-b \cdot x)$].

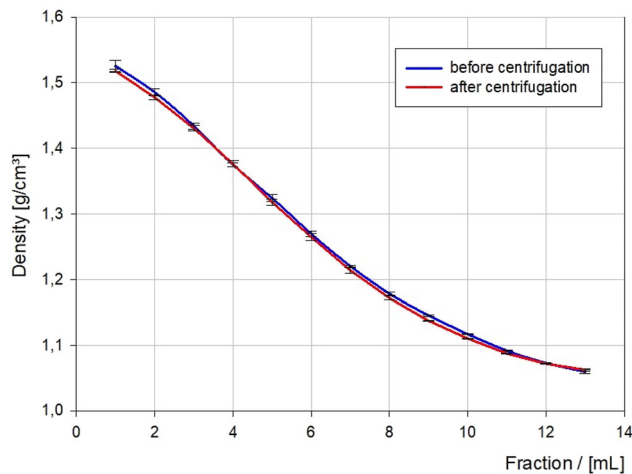


Figure 4: Densities of each fraction of the potassium-carbonate-and-water gradient before (blue) and after (red) centrifugation.

As shown in Figure 4 there are no significant changes in the gradient before and after centrifugation. Generally, the gradients are characterized by their high stability. Therefore, the handling of the gradients is also suitable for students handling the gradients with little caution.

The initial phase of exponential density decay can be approximated by a linear decay, especially in fractions 2–9, allowing for reliable separation of microplastic particles in this density range.

4.2 Fractionation of microplastic particles by density gradient centrifugation

Preliminary results with the sucrose gradient were promising separating plastic types polypropylene (PP), polystyrol (PS), polyamide (PA), polycarbonate (PC) and polyethylene terephthalate (PET) (see Figure 5) as confirmed by ATR-IR spectroscopy (see Supplementary Information).

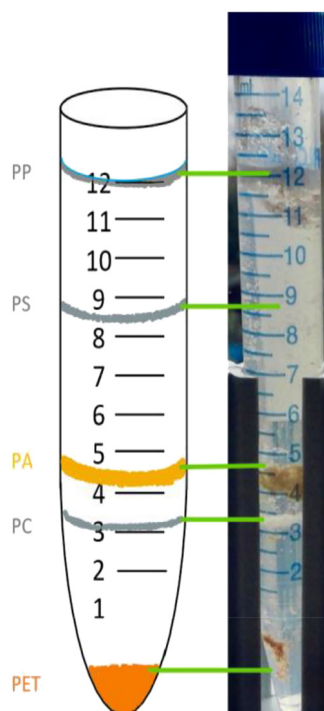


Figure 5: A picture of the sucrose gradient with microplastic particles of the plastic types PP, PS, PA, PC and PET after centrifugation showing clear bands for each plastic type as confirmed by ATR-IR spectroscopy.

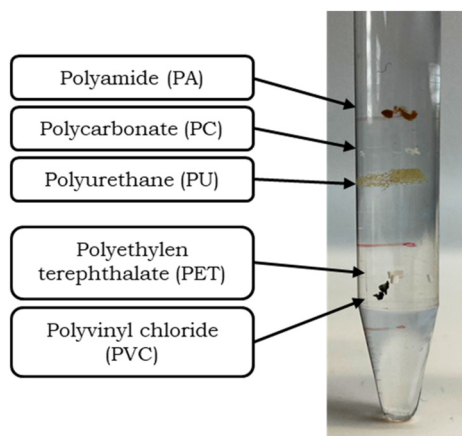


Figure 6: Picture of fractioned microplastic particles after centrifugation in a saturated-potassium-carbonate-and-water density gradient showing clear bands for each plastic type, attributed to the density of each fraction.

Furthermore, using the potassium carbonate gradient for fractionation makes the separation of PA, PC, PU, PET and PVC easily accessible. After centrifugation, the tested microplastic particles were separated into five well-defined bands (see Figure 6). The bands were in the same place in all 10 samples tested.

Particles were then identified visually, and the density of the fraction was compared to the specific density of the plastic types. All five polymers formed a band in their specific density range, showing this method is viable for separation of multiple microplastic particles by plastic type. Separation efficiency is similar to other protocols using caesium chloride solution (Jakobs et al., 2023; Jing et al., 2022), making a saturated potassium carbonate solution suitable and favourable as a non-hazardous and cost-effective alternative for school context. It should be noted that the separation does not allow for a clear identification due to the partially overlapping density ranges of some plastic types. Density can also vary due to the use of additives (such as plasticizers, flame retardants, slip additives and thermostabilizers) or non-polymeric fillers (Andrady, 2015; Hale et al., 2020). Nevertheless, density separation by gradient centrifugation allows an approximate estimation of the type of the microplastic particles.

4.3 Workshop for teachers

In order to transfer the knowledge created by the students from school and extended by further development at university back to school, a hands-on experience working with this device has already been integrated in workshops for teachers. In addition to theoretical information on the topic of microplastics, the teachers also received an introduction to the analysis of microplastics in the workshop. Afterwards, the teachers had enough time to practice the methods actively and independently with the help of instruction sheets that were developed to be used with students (see Supplementary Information). A subsequent plenum discussion showed potentials for integration of the device into upper secondary chemistry classes. The workshop received a lot of positive feedback. Following the workshops, teachers reported that they had already successfully used the gradient maker and method in their laboratory lessons with minor, class-specific modifications and plan to implement it on a more regular basis.

5 Conclusion and outlook

Using the novel Luer-Lock low-cost density gradient maker it is possible to easily produce stable and reproducible density gradients in a school and university context. Both a sucrose solution and a saturated potassium carbonate solution were shown to be suitable, low-cost, easy-to-handle and non-hazardous alternatives to a caesium chloride solution, which is state of the art in current research. We have also shown that with our approach density gradient centrifugation can be used to fractionate microplastic particles of the types PA, PUR, PC, PET and PVC

based on their specific particle densities. Lower density polymers such as PE, PP and EPS might be fractionated in an ethanol-water gradient. First tests have shown promising results.

This simple yet powerful method can be used not only to address the environmental microplastic problem in classes, but also to show some limitations as well as challenges of microplastic analysis. In addition, it might contribute to career guidance towards analytical chemistry. Furthermore, the experimental setting can be used to quickly visualize density using a specific example from a real-world-context. Another fruitful field of application may be Citizen Science projects for identification of microplastic particles in real-world samples. First tests at a marine biology station in Croatia are already showing promising results.

Supplementary information

ATR-IR-Spectra of microplastic particles in the sucrose gradient; further information and a picture of a non-functional prototype gradient maker; statements on hazards and precautionary measures.

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