

## Opinion Paper

Burak Arslan\*, Henrik Zetterberg and Nicholas J. Ashton

# Blood-based biomarkers in Alzheimer's disease – moving towards a new era of diagnostics

<https://doi.org/10.1515/cclm-2023-1434>

Received December 13, 2023; accepted January 5, 2024;

published online January 23, 2024

**Abstract:** Alzheimer's disease (AD), a primary cause of dementia globally, is traditionally diagnosed via cerebrospinal fluid (CSF) measures and positron emission tomography (PET). The invasiveness, cost, and limited accessibility of these methods have led to exploring blood-based biomarkers as a promising alternative for AD diagnosis and monitoring. Recent advancements in sensitive immunoassays have identified potential blood-based biomarkers, such as A $\beta$ 42/A $\beta$ 40 ratios and phosphorylated tau (p-tau) species. This paper briefly evaluates the clinical utility and reliability of these biomarkers across various AD stages, highlighting challenges like refining plasma A $\beta$ 42/A $\beta$ 40 assays and enhancing the precision of p-tau, particularly p-tau181, p-tau217, and p-tau231. The discussion also covers other plasma biomarkers like neurofilament light (NfL), glial

fibrillary acidic protein (GFAP), and synaptic biomarkers, assessing their significance in AD diagnostics. The need for ongoing research and development of robust assays to match the performance of CSF and PET biomarkers is underscored. In summary, blood-based biomarkers are increasingly crucial in AD diagnosis, follow-up, prognostication, treatment response evaluation, and population screening, particularly in primary care settings. These developments are set to revolutionize AD diagnostics, offering earlier and more accessible detection and management options.

**Keywords:** Alzheimer's disease; amyloid; blood; biomarker; CSF; tau

## Introduction

Alzheimer's disease (AD) is a chronic, neurodegenerative disease and the leading cause of dementia that affects more than 50 million people worldwide [1]. This number is projected to be greater than 150 million by 2050 [1]. The pathological hallmarks of AD are extracellular amyloid- $\beta$  (A $\beta$ ) plaques, intracellular tau tangles, and neurodegeneration, which are needed to make a definitive diagnosis *post-mortem*. However, considerable progress has been made to detect and quantify these pathologies *in vivo* using biomarkers. Cerebrospinal (CSF) fluid measures of A $\beta$ 42, A $\beta$ 42/A $\beta$ 40, phosphorylated tau (p-tau), total tau, and neurofilament light (NfL) chain are routinely examined as accessible measures of A $\beta$ , tau and neurodegeneration [2]. CSF A $\beta$ 42 has been validated analytically and clinically; assays to measure the marker are now also fully standardized through the development of certified reference methods and materials [3, 4]. This work is well underway for A $\beta$ 40 and has just started for p-tau and NfL. Positron emission tomography (PET) allows for regional visualization of A $\beta$  and tau pathologies, which have a high concordance with CSF measures. Glucose metabolism (fluorodeoxyglucose [FDG]-PET) and temporal atrophy by magnetic resonance imaging (MRI) provide imaging measures of neurodegeneration [5]. These biomarkers are now defined in the context of AD research frame ATN (amyloid/tau/neurodegeneration)

**\*Corresponding author: Burak Arslan**, MD, Department of Psychiatry and Neurochemistry, Institute of Neuroscience and Physiology, Sahlgrenska Academy at Gothenburg University, Mölndal Hospital, Hus V3, 43180 Mölndal, Sweden; and Clinical Neurochemistry Laboratory, Sahlgrenska University Hospital, Mölndal, Sweden, E-mail: burak.arslan@gu.se

**Henrik Zetterberg**, Department of Psychiatry and Neurochemistry, Institute of Neuroscience and Physiology, The Sahlgrenska Academy at the University of Gothenburg, Mölndal, Sweden; Clinical Neurochemistry Laboratory, Sahlgrenska University Hospital, Mölndal, Sweden; Wisconsin Alzheimer's Disease Research Center, School of Medicine and Public Health, University of Wisconsin-Madison, Madison, WI, USA; Department of Neurodegenerative Disease, UCL Institute of Neurology, London, UK; UK Dementia Research Institute at UCL, London, UK; and Hong Kong Center for Neurodegenerative Diseases, Clear Water Bay, Hong Kong, P.R. China

**Nicholas J. Ashton**, Department of Psychiatry and Neurochemistry, Institute of Neuroscience and Physiology, The Sahlgrenska Academy at the University of Gothenburg, Mölndal, Sweden; Department of Old Age Psychiatry, Psychology & Neuroscience, King's College London, Institute of Psychiatry, London, UK; King's College London, Institute of Psychiatry, Psychology and Neuroscience, Maurice Wohl Institute Clinical Neuroscience Institute, London, UK; NIHR Biomedical Research Centre for Mental Health & Biomedical Research Unit for Dementia at South London & Maudsley, NHS Foundation, London, UK; and Centre for Age-Related Medicine, Stavanger University Hospital, Stavanger, Norway

which proposes a shift in the definition of the disease from a clinical syndrome to a biological construct [3]. This definition is of immediate clinical importance given that disease-modifying therapies that effectively remove A $\beta$  pathology from the brain have received regulatory approval in the US, and will likely become widely available globally in the coming years. The ATN criteria are currently undergoing revision to describe the progression of biomarker changes over time and to encourage their clinical use for diagnosis and staging of the disease [6]. The perceived invasiveness or reluctance of a lumbar puncture and the relatively high cost of PET examination restrict their use for large-scale ATN classification of patients—especially in primary care settings or population screening. Therefore, current efforts have been made to develop blood-based biomarkers to replace or complement the currently available CSF and PET biomarkers. With the advent of highly sensitive immunoassays, the development of blood-based biomarkers has accelerated; mounting evidence now support their clinical usefulness and reliability in both prodromal and dementia stages of the disease.

Blood-based biomarkers in AD

In the context of AD, the most used blood-based biomarkers are A $\beta$ 42/A $\beta$ 40 and phosphorylated tau (p-tau) species (e.g., p-tau181, p-tau217, p-tau231). A detailed summary of key biomarkers relevant to Alzheimer's disease is presented in Table 1.

Plasma amyloid beta biomarkers

Initially, plasma A $\beta$ 42 concentration was measured using enzyme-linked immunosorbent assays (ELISA); there was, however, not much hope for its usefulness due to the low performance of the assay and inconsistent findings in different cohorts [18]. This changed with the advent of Single molecule array (Simoa) technology that demonstrated significant reduction of A $\beta$ 42/A $\beta$ 40 in the AD *continuum* despite weak correlations with CSF and PET [19, 20]. Other assays (e.g., Elecsys using electrochemiluminescence [ECL] [21],

Table 1: Comprehensive overview of the blood-based biomarkers in Alzheimer's disease: methods, correlations, and implications.

Biomarker	Methods and process	Remarks
Beta amyloid 1–42/beta amyloid 1–40 (A $\beta$ 42/A $\beta$ 40) ratio	Mass spectrometry assays, immunoassays; Cerebral A $\beta$ pathology	A decline in A $\beta$ 42 levels is indicative of amyloid accumulation in the brain [2, 7, 8] A lower A $\beta$ 42/A $\beta$ 40 ratio aligns closely with amyloid deposition as identified by amyloid PET imaging [9, 10] There is a high correlation between cerebrospinal fluid (CSF) and blood A $\beta$ 42/A $\beta$ 40 levels [4, 9]
Phosphorylated tau protein (P-tau)	Mass spectrometry assays, immunoassays; Neuronal tau phosphorylation and secretion	P-tau levels are increased in both manifest and asymptomatic stages of Alzheimer's disease [9] There is a significant correlation between CSF and blood levels of P-tau [4, 9]
Subtypes of P-tau (P-tau 181, P-tau-217, P-tau-205, P-tau231)	Mass spectrometry assays, immunoassays; neuronal tau phosphorylation and secretion	Variants of phosphorylated tau protein, including P-tau 181, P-tau 217, P-tau 205, and P-tau 231, show increased levels in both symptomatic and asymptomatic [11–14] Alzheimer's disease. These subtypes of P-tau are highly sensitive and specific markers for Alzheimer's disease [7, 9]
Neurofilament light (NFL) protein	Immunoassays, neurodegeneration	NfL mirrors the pathology of neurofibrillary tangles and the overall severity of Alzheimer's disease [15, 16] There is a close correlation between CSF concentrations of NFL and its levels in Alzheimer's disease [16]
Glial fibrillary acidic protein (GFAP)	Immunoassays, astrocyte activation	GFAP indicates the presence of neuroinflammation and neurodegeneration [17] The levels of GFAP in the plasma are indicative of the extent of astrogliosis [9, 17]

This Table provides a detailed summary of key biomarkers relevant to Alzheimer's disease, including A $\beta$ 42/A $\beta$ 40 ratio, phosphorylated tau protein variants, neurofilament light protein, and glial fibrillary acidic protein. Each biomarker is accompanied by its detection methods, primarily mass spectrometry and immunoassays, and remarks on its significance in Alzheimer's pathology.

Lumipulse using chemiluminescent enzyme immunoassay [CLEIA] [22], and immunoprecipitation mass-spectrometry [IP-MS] [23, 24]) independently confirmed the decrease in plasma A $\beta$ 42 and A $\beta$ 42/A $\beta$ 40 ratio in those with confirmed cerebral amyloid pathology. In a head-to-head study to detect brain A $\beta$  pathology via blood-based measurements, mass spectrometry-based methods (IP-MS) have been shown to perform slightly better than immunoassays [25], but the latter are catching up [26]. Previous research has indicated that reductions in both CSF and plasma A $\beta$ 42/A $\beta$ 40 ratios occur early in the Alzheimer's *continuum* and predict the transition from amyloid-negative to amyloid-positive PET status [24, 27]. Nevertheless, the main confounding factor with plasma A $\beta$  measurements that must be overcome is the small-fold change between A $\beta$ -positive and -negative individuals (only 8–15 % in plasma compared with 40–60 % in CSF [9]). In addition, multiple pharmacodynamic effects on plasma A $\beta$  by common drugs (e.g., neprilysin inhibitors) may further restrict the use of A $\beta$  peptides in older adults (Brum et al., [28]). Overall, the low robustness of plasma A $\beta$ 42/A $\beta$ 40 is a significant challenge to making use of the biomarker in clinical settings [10] – small drifts in assay performance or variation in pre-analytical factors could cause individuals to be misclassified. Plasma A $\beta$  could be highly important for establishing novel disease-modifying drugs. Gamma-secretase modulators (GSMs), shifting A $\beta$  production from the amyloidogenic A $\beta$ 42 to the less pathogenic A $\beta$ 38 and maintaining overall A $\beta$  levels, hold promise for primary prevention of AD pathology, making A $\beta$ 38 an important biomarker in blood [29].

## Plasma tau biomarkers

So far, several plasma p-tau assays have been developed and used to show AD-related changes. Immunoassay and mass spectrometric measures of p-tau181, p-tau217 and p-tau231 discriminate A $\beta$ -positive from -negative individuals with high performance in many studies [11, 30, 31]. High-performing p-tau blood tests exhibit a substantial increase in AD patients with increases occurring concurrently with extracellular A $\beta$  plaque deposition. This relationship is observed across the AD *continuum*, including the presymptomatic phase in sporadic and familial AD [7]. This includes individuals with Down syndrome who have a genetically determined form of AD [32]. P-tau species (especially p-tau217 and p-tau231) start to increase just after the drop in CSF A $\beta$ 42/A $\beta$ 40 ratio and prior to the cut-point for A $\beta$ -PET positivity has been reached. This is also a promising finding for including people in the pre-dementia stage in clinical trials, which is of great importance for disease-modifying treatments. Certain p-tau

species [12], however, are associated with neurofibrillary tangle pathology [33, 34], as indexed by tau-PET imaging and neuropathological examination. Unlike plasma A $\beta$ 42/40, plasma p-tau is reliably measured using immunoassays with large fold-changes, which is an encouraging finding to implement this biomarker in routine clinical practice widely. There is much debate as to which p-tau biomarker has the most potential as a clinical biomarker. While all p-tau tests have high discriminative accuracy, p-tau217 has the largest fold-change between AD and non-AD disorders [8] and is more related to AD progression [7]. All in all, plasma p-tau is a promising biomarker to show AD-type brain pathologies, detect individuals with pre-clinical AD, stage them as they progress, and, potentially, track treatment efficacy [35].

Building on the research of p-tau, the role of the microtubule-binding region (MTBR) of tau in AD becomes particularly pertinent [36, 37]. This region of tau contains the building blocks of neurofibrillary tangles and could thus be a very different biomarker compared with the N-terminal p-tau fragments discussed above, which mostly reflect A $\beta$  pathology.

## Plasma neurodegeneration biomarkers

In the ATN criteria, total tau is classified as biomarker for neurodegeneration in CSF. However, this relationship has not translated well to blood [38]. Plasma total tau levels are not well correlated with its CSF levels, possibly because of peripheral production of total tau, and are not specific for AD [4]. Thus, NfL has been used as a neuroaxonal injury marker research settings and specialized clinical routine laboratories where NfL has already been implemented [15, 39]. As a general neurodegeneration marker, NfL plasma levels are also elevated in amyotrophic lateral sclerosis, frontotemporal dementia, traumatic brain injury, and peripheral neuropathy, which should be considered while assessing possible AD cases [16]. NfL is also an excellent prognostic marker for neurological outcome acutely after cardiac arrest [40]. In AD cases, plasma NfL levels are found to be increased in relation to A $\beta$  and tau positivity [41], but the lack of specificity to AD, minimal longitudinal change associated to atrophy [7], cognitive decline and amyloid clearance [42] may limit NfL in the context of AD. To accurately detect and track AD-specific brain-derived tau levels in the blood, a novel brain-derived tau assay has recently been developed, and it discriminated autopsy-confirmed AD and non-AD tauopathies, which was better than NfL performance [43]. To

overcome the lack of robustness of plasma total tau assays currently used, and distinguish AD-type neurodegeneration, brain-derived total tau has great promise for its future implementation in AD management [43]. In addition, recent studies highlight the significance of plasma N-terminal tau (NTA) in AD, demonstrating its ability to predict future cognitive decline and neurodegeneration even in pre-symptomatic stages of AD, with higher baseline levels of NTA strongly associated with the progression to mild cognitive impairment (MCI) to AD dementia, and effectively differentiating between normal, mildly-impaired, and AD dementia populations, thereby underlining the potential of NTA as a non-invasive, sensitive biomarker for early detection and monitoring of AD pathology [44, 45].

## Plasma astrocytic activation biomarker

Glial fibrillary acidic protein (GFAP) is one of the most studied glial marker, which is likely released as a quick response to A $\beta$  pathology in the context of AD. Interestingly, it has been shown that plasma GFAP levels are more related to brain A $\beta$  pathology than CSF levels of GFAP, which may be due to instability of GFAP in CSF [17, 46, 47]. Although GFAP is not an AD-specific marker, the magnitude of GFAP change is relatively large in AD compared with those of non-AD neurodegenerative diseases such as FTD [47]. Furthermore, along with the plasma markers of amyloid, tau, and neurodegeneration, plasma GFAP levels were also observed to decrease after amyloid-removal therapy (lecanemab) in a phase 3 clinical trial [35].

In addition to GFAP, the role of S100B in AD appears multifaceted and warrants careful interpretation. While various studies suggest its potential as a biomarker due to its correlation with brain atrophy [48] and inverse relationship with MMSE scores [49] in AD patients, the expression of S100B in other cell types beyond the brain necessitates caution in interpreting serum levels, as they might reflect peripheral changes rather than central nervous system pathology [50]. Moreover, the involvement of S100B in AD pathophysiology is further supported by its astrocytic over-expression driven by IL-1 from activated microglia, indicating its potential as a marker of underlying pathological processes in AD [51]. However, considering the complexity of its expression and the factors influencing its levels, the utility of S100B utility in AD diagnosis and monitoring is uncertain.

YKL-40, chitinase 3-L1, CHI3L1, shows promise as a biomarker for early detection and progression monitoring of

MCI-AD and dementia due to AD, with its levels correlating with disease severity [52, 53]. However, its effectiveness is enhanced when combined with other AD biomarkers, due to its non-specificity to AD dementia alone [54]. Given its lack of specificity to AD, the diagnostic potential of YKL-40 could be enhanced when used in conjunction with other biomarkers like A $\beta$ 42/40, total tau, and p-tau, offering a more comprehensive approach to AD diagnosis and monitoring.

## Synaptic biomarkers

The loss of synapses is generally considered the most accurate indicator of the deterioration in cognitive abilities associated with AD [55]. In relation to this, the dendritic protein neurogranin (Ng) has been the subject of thorough research. Studies have shown that the level of Ng in the CSF is elevated in individuals with AD. This elevation correlates with the concentrations of t-tau and p-tau proteins. Furthermore, the increased levels of Ng are linked to the gradual deterioration of cognitive abilities as time progresses. The impact of this increase is further influenced by the presence of A $\beta$  pathology [56]. However, plasma Ng levels do not correlate with those in the CSF. This lack of correlation is likely due to the production of the protein outside the brain [57]. Hence, it is improbable that Ng levels in the plasma will serve as a useful blood-based biomarker for AD. However,  $\beta$ -synuclein and synaptosomal-associated protein 25 (SNAP-25) are promising blood biomarkers for synaptic damage in AD.  $\beta$ -Synuclein is elevated in both CSF and blood of AD patients [58]. This increase is notably associated with brain atrophy in AD, distinguishing it from other conditions like frontotemporal lobar degeneration [59]. However, further research is needed to understand its role in preclinical stages of AD and its relationship with amyloid and tau pathologies [60]. SNAP-25 is a crucial presynaptic protein that belongs to the soluble N-ethylmaleimide-sensitive factor attachment receptor (SNARE) family. It plays a vital role in cognitive processes because of its involvement in the mechanism of vesicular exocytosis [61]. In the past, the levels of these proteins could only be determined in CSF, and they have been found to be elevated in individuals with AD [61]. A recent study has demonstrated increased SNAP-25 levels in the plasma of AD patients, showing a significant correlation with cognitive function and cortical atrophy [62]. Further studies are required to confirm these findings in separate groups and to investigate how plasma SNAP-25 levels relate to other biomarkers across a wide spectrum of neurodegenerative diseases.



## Conclusions

Blood-based biomarkers of AD have great promise in AD diagnosis, follow-up, prognostication, tracking treatment response, and population screening. Using highly sensitive assays, the levels of blood-based biomarkers are easily determined and may reliably be used in routine clinical settings. Much more effort should be on developing more robust assays and achieving as high performance as core CSF and PET biomarkers of AD. If individuals with pre-clinical AD can be caught via blood-based biomarkers, especially in primary care settings, more promising treatment options would appear on the horizon. Simplified protocols for blood biomarker assessment through finger prick testing are currently being developed [63].

**Research ethics:** Not applicable.

**Informed consent:** Not applicable.

**Author contributions:** All authors have accepted responsibility for the entire content of this manuscript and approved its submission.

**Conflicts of interest:** NJA has given lectures, produced educational materials, and participated in educational programs for Eli-Lily, BioArtic, Quanterix. HZ has served at scientific advisory boards and/or as a consultant for Abbvie, Acumen, Alector, Alzinova, ALZPath, Annexon, Apellis, Artery Therapeutics, AZTherapies, Cognito Therapeutics, CogRx, Denali, Eisai, Merry Life, Nervgen, Novo Nordisk, Optoceutics, Passage Bio, Pinteon Therapeutics, Prothena, Red Abbey Labs, reMYND, Roche, Samumed, Siemens Healthineers, Triplet Therapeutics, and Wave, has given lectures in symposia sponsored by Alzecure, Biogen, Cellectricon, Fujirebio, Lilly, and Roche, and is a co-founder of Brain Biomarker Solutions in Gothenburg AB (BBS), which is a part of the GU Ventures Incubator Program (outside submitted work).

**Research funding:** HZ is a Wallenberg Scholar supported by grants from the Swedish Research Council (#2023-00356; #2022-01018 and #2019-02397), the European Union's Horizon Europe research and innovation programme under grant agreement No 101053962, Swedish State Support for Clinical Research (#ALFGBG-71320), the Alzheimer Drug Discovery Foundation (ADDF), USA (#201809-2016862), the AD Strategic Fund and the Alzheimer's Association (#ADSF-21-831376-C, #ADSF-21-831381-C, and #ADSF-21-831377-C), the Bluefield Project, the Olav Thon Foundation, the Erling-Persson Family Foundation, Stiftelsen för Gamla Tjänarinnor, Hjärnfonden, Sweden (#FO2022-0270), the European Union's Horizon 2020 research and innovation programme under the Marie Skłodowska-Curie grant agreement No 860197

(MIRIADE), the European Union Joint Programme – Neurodegenerative Disease Research (JPND2021-00694), the National Institute for Health and Care Research University College London Hospitals Biomedical Research Centre, and the UK Dementia Research Institute at UCL (UKDRI-1003).

**Data availability:** Not applicable.

## References

- Nichols E, Steinmetz JD, Vollset SE, Fukutaki K, Chalek J, Abd-Allah F, et al. Estimation of the global prevalence of dementia in 2019 and forecasted prevalence in 2050: an analysis for the global burden of disease study 2019. *Lancet Public Health* 2022;7:e105–25.
- Blennow K, Zetterberg H. Biomarkers for Alzheimer's disease: current status and prospects for the future. *J Intern Med* 2018;284: 643–63.
- Jack CR, Jr., Bennett DA, Blennow K, Carrillo MC, Dunn B, Haeberlein SB, et al. NIA-AA research framework: toward a biological definition of Alzheimer's disease. *Alzheimer's Dementia* 2018;14:535–62.
- Zetterberg H, Blennow K. Moving fluid biomarkers for Alzheimer's disease from research tools to routine clinical diagnostics. *Mol Neurodegener* 2021;16:10.
- Ossenkoppele R, Hansson O. Towards clinical application of tau PET tracers for diagnosing dementia due to Alzheimer's disease. *Alzheimer's Dementia* 2021;17:1998–2008.
- Association As. Diagnostic criteria & guidelines. Available from: [https://www.alz.org/research/for\\_researchers/diagnostic-criteria-guidelines](https://www.alz.org/research/for_researchers/diagnostic-criteria-guidelines).
- Ashton NJ, Janelidze S, Mattsson-Carlgen N, Binette AP, Strandberg O, Brum WS, et al. Differential roles of Aβ42/40, p-tau231 and p-tau217 for Alzheimer's trial selection and disease monitoring. *Nat Med* 2022;28: 2555–62.
- Ashton NJ, Puig-Pijoan A, Milà-Alomà M, Fernández-Lebrero A, García-Escobar G, González-Ortiz F, et al. Plasma and CSF biomarkers in a memory clinic: head-to-head comparison of phosphorylated tau immunoassays. *Alzheimer's Dementia* 2023;19:1913–24.
- Zetterberg H, Schott JM. Blood biomarkers for Alzheimer's disease and related disorders. *Acta Neurol Scand* 2022;146:51–5.
- Benedet AL, Brum WS, Hansson O, Karikari TK, Zimmer ER, Zetterberg H, et al. The accuracy and robustness of plasma biomarker models for amyloid PET positivity. *Alzheimer's Res Ther* 2022;14:26.
- Ashton NJ, Pascoal TA, Karikari TK, Benedet AL, Lantero-Rodriguez J, Brinkmalm G, et al. Plasma p-tau231: a new biomarker for incipient Alzheimer's disease pathology. *Acta Neuropathol* 2021;141:709–24.
- Montoliu-Gaya L, Benedet AL, Tissot C, Vrillon A, Ashton NJ, Brum WS, et al. Mass spectrometric simultaneous quantification of tau species in plasma shows differential associations with amyloid and tau pathologies. *Nat Aging* 2023;3:661–9.
- Karikari TK, Pascoal TA, Ashton NJ, Janelidze S, Benedet AL, Rodriguez JL, et al. Blood phosphorylated tau 181 as a biomarker for Alzheimer's disease: a diagnostic performance and prediction modelling study using data from four prospective cohorts. *Lancet Neurol* 2020;19:422–33.
- Ashton NJ, Brum WS, Di Molfetta G, Benedet AL, Arslan B, Jonatis E, et al. Diagnostic accuracy of the plasma ALZpath pTau217 immunoassay to identify Alzheimer's disease pathology. *medRxiv* [Preprint] 2023. <https://doi.org/10.1101/2023.07.11.23292493>.

15. Arslan B, Zetterberg H. Neurofilament light chain as neuronal injury marker – what is needed to facilitate implementation in clinical laboratory practice? *Clin Chem Lab Med* 2023;61:1140–9.
16. Ashton NJ, Janelidze S, Al Khleifat A, Leuzy A, van der Ende EL, Karikari TK, et al. A multicentre validation study of the diagnostic value of plasma neurofilament light. *Nat Commun* 2021;12:3400.
17. Benedet AL, Milà-Alomà M, Vrillon A, Ashton NJ, Pascoal TA, Lussier F, et al. Differences between plasma and cerebrospinal fluid glial fibrillary acidic protein levels across the Alzheimer disease continuum. *JAMA Neurol* 2021;78:1471–83.
18. Olsson B, Lautner R, Andreasson U, Öhrfelt A, Portelius E, Bjerke M, et al. CSF and blood biomarkers for the diagnosis of Alzheimer's disease: a systematic review and meta-analysis. *Lancet Neurol* 2016;15: 673–84.
19. Janelidze S, Zetterberg H, Mattsson N, Palmqvist S, Vanderstichele H, Lindberg O, et al. CSF A $\beta$ 42/A $\beta$ 40 and A $\beta$ 42/A $\beta$ 38 ratios: better diagnostic markers of Alzheimer disease. *Ann Clin Transl Neurol* 2016;3:154–65.
20. Ashton N, Leuzy A, Karikari T, Mattsson-Carlsson N, Dodich A, Boccardi M, et al. The validation status of blood biomarkers of amyloid and phospho-tau assessed with the 5-phase development framework for AD biomarkers. *Eur J Nucl Med Mol Imaging* 2021;48:2140–56.
21. Palmqvist S, Janelidze S, Stomrud E, Zetterberg H, Karl J, Zink K, et al. Performance of fully automated plasma assays as screening tests for Alzheimer disease-related  $\beta$ -amyloid status. *JAMA Neurol* 2019;76: 1060–9.
22. Martínez-Dubarbíe F, Guerra-Ruiz A, López-García S, Lage C, Fernández-Matarrubia M, Infante J, et al. Accuracy of plasma A $\beta$ 40, A $\beta$ 42, and p-tau181 to detect CSF Alzheimer's pathological changes in cognitively unimpaired subjects using the Lumipulse automated platform. *Alzheimer's Res Ther* 2023;15:163.
23. Nakamura A, Kaneko N, Villemagne VL, Kato T, Doecke J, Doré V, et al. High performance plasma amyloid- $\beta$  biomarkers for Alzheimer's disease. *Nature* 2018;554:249–54.
24. Schindler SE, Bollinger JG, Ovod V, Mawuenyega KG, Li Y, Gordon BA, et al. High-precision plasma  $\beta$ -amyloid 42/40 predicts current and future brain amyloidosis. *Neurology* 2019;93:e1647–59.
25. Janelidze S, Teunissen CE, Zetterberg H, Allué JA, Sarasa L, Eichenlaub U, et al. Head-to-head comparison of 8 plasma amyloid- $\beta$  42/40 assays in Alzheimer disease. *JAMA Neurol* 2021;78:1375–82.
26. Zicha S, Bateman RJ, Shaw LM, Zetterberg H, Bannan AW, Horton WA, et al. Comparative analytical performance of multiple plasma A $\beta$ 42 and A $\beta$ 40 assays and their ability to predict positron emission tomography amyloid positivity. *Alzheimer's Dementia* 2023;19:956–66.
27. Milà-Alomà M, Salvadó G, Gispert JD, Vilor-Tejedor N, Grau-Rivera O, Sala-Vila A, et al. Amyloid beta, tau, synaptic, neurodegeneration, and glial biomarkers in the preclinical stage of the Alzheimer's continuum. *Alzheimer's Dementia* 2020;16:1358–71.
28. Brum WS, Docherty KF, Ashton NJ, Zetterberg H, Hansson O, McMurray JJV, et al. Effect of neprilysin inhibition on alzheimer disease plasma biomarkers: a secondary analysis of a randomized clinical trial. *JAMA Neurol* 2023;e234719. <https://doi.org/10.1001/jamaneurol.2023.4719>.
29. Cullen N, Janelidze S, Palmqvist S, Stomrud E, Mattsson-Carlsson N, Hansson O, et al. Association of CSF A $\beta$ 38 levels with risk of Alzheimer disease-related decline. *Neurology* 2022;98:e958–67.
30. Bayoumy S, Verberk IMW, den Dulk B, Hussaini Z, Zwan M, van der Flier WM, et al. Clinical and analytical comparison of six Simoa assays for plasma P-tau isoforms P-tau181, P-tau217, and P-tau231. *Alzheimer's Res Ther* 2021;13:198.
31. Palmqvist S, Janelidze S, Quiroz YT, Zetterberg H, Lopera F, Stomrud E, et al. Discriminative accuracy of plasma phospho-tau217 for Alzheimer disease vs other neurodegenerative disorders. *JAMA* 2020;324:772–81.
32. Lleó A, Zetterberg H, Pegueroles J, Karikari TK, Carmona-Iragui M, Ashton NJ, et al. Phosphorylated tau181 in plasma as a potential biomarker for Alzheimer's disease in adults with Down syndrome. *Nat Commun* 2021;12:4304.
33. Mattsson-Carlsson N, Janelidze S, Bateman RJ, Smith R, Stomrud E, Serrano GE, et al. Soluble P-tau217 reflects amyloid and tau pathology and mediates the association of amyloid with tau. *EMBO Mol Med* 2021; 13:e14022.
34. Salvadó G, Ossenkoppele R, Ashton NJ, Beach TG, Serrano GE, Reiman EM, et al. Specific associations between plasma biomarkers and postmortem amyloid plaque and tau tangle loads. *EMBO Mol Med* 2023;15:e17123.
35. Van Dyck CH, Swanson CJ, Aisen P, Bateman RJ, Chen C, Gee M, et al. Lecanemab in early Alzheimer's disease. *N Engl J Med* 2023;388:9–21.
36. Horie K, Salvadó G, Barthélemy NR, Janelidze S, Li Y, He Y, et al. CSF MTBR-tau243 is a specific biomarker of tau tangle pathology in Alzheimer's disease. *Nat Med* 2023;29:1954–63.
37. Horie K, Barthélemy NR, Sato C, Bateman RJ. CSF tau microtubule binding region identifies tau tangle and clinical stages of Alzheimer's disease. *Brain* 2021;144:515–27.
38. Mattsson N, Zetterberg H, Janelidze S, Insel PS, Andreasson U, Stomrud E, et al. Plasma tau in Alzheimer disease. *Neurology* 2016;87: 1827–35.
39. Andreasson U, Gobom J, Delatour V, Auclair G, Noam Y, Lee S, et al. Assessing the commutability of candidate reference materials for the harmonization of neurofilament light measurements in blood. *Clin Chem Lab Med* 2023;61:1245–54.
40. Wihersaari L, Ashton NJ, Reinikainen M, Jakkula P, Pettilä V, Hästbacka J, et al. Neurofilament light as an outcome predictor after cardiac arrest: a post hoc analysis of the COMACARE trial. *Intensive Care Med* 2021;47: 39–48.
41. Mattsson N, Andreasson U, Zetterberg H, Blennow K. Association of plasma neurofilament light with neurodegeneration in patients with Alzheimer disease. *JAMA Neurol* 2017;74:557–66.
42. Sims JR, Zimmer JA, Evans CD, Lu M, Ardayfio P, Sparks J, et al. Donanemab in early symptomatic Alzheimer disease: the TRAILBLAZER-ALZ 2 randomized clinical trial. *JAMA* 2023;330:512–27.
43. Gonzalez-Ortiz F, Turton M, Kac PR, Smirnov D, Premi E, Ghidoni R, et al. Brain-derived tau: a novel blood-based biomarker for Alzheimer's disease-type neurodegeneration. *Brain* 2023;146:1152–65.
44. Lantero-Rodriguez J, Tissot C, Snellman A, Servaes S, Benedet AL, Rahmouni N, et al. Plasma and CSF concentrations of N-terminal tau fragments associate with in vivo neurofibrillary tangle burden. *Alzheimer's Dementia* 2023;19:5343–54.
45. Woo MS, Tissot C, Lantero-Rodriguez J, Snellman A, Theriault J, Rahmouni N, et al. Plasma pTau-217 and N-terminal tau (NTA) enhance sensitivity to identify tau PET positivity in amyloid- $\beta$  positive individuals. *Alzheimer's Dementia* 2023;1–9. <https://doi.org/10.1002/alz.13528> [Epub ahead of print].
46. Pereira JB, Janelidze S, Smith R, Mattsson-Carlsson N, Palmqvist S, Teunissen CE, et al. Plasma GFAP is an early marker of amyloid- $\beta$  but not tau pathology in Alzheimer's disease. *Brain* 2021;144:3505–16.
47. Heller C, Foiani MS, Moore K, Convery R, Bocchetta M, Neason M, et al. Plasma glial fibrillary acidic protein is raised in progranulin-associated frontotemporal dementia. *J Neurol Neurosurg Psychiatry* 2020;91: 263–70.

48. Petzold A, Jenkins R, Watt H, Green A, Thompson E, Keir G, et al. Cerebrospinal fluid S100B correlates with brain atrophy in Alzheimer's disease. *Neurosci Lett* 2003;336:167–70.
49. Chaves ML, Camozzato AL, Ferreira ED, Piazenski I, Kochhann R, Dall'Igna O, et al. Serum levels of S100B and NSE proteins in Alzheimer's disease patients. *J Neuroinflammation* 2010;7:1–7.
50. Michetti F, D'Ambrosi N, Toesca A, Puglisi MA, Serrano A, Marchese E, et al. The S100B story: from biomarker to active factor in neural injury. *J Neurochem* 2019;148:168–87.
51. Mrak RE, Griffin WST. The role of activated astrocytes and of the neurotrophic cytokine S100B in the pathogenesis of Alzheimer's disease. *Neurobiol Aging* 2001;22:915–22.
52. Janelidze S, Hertz J, Zetterberg H, Landqvist Waldö M, Santillo A, Blennow K, et al. Cerebrospinal fluid neurogranin and YKL-40 as biomarkers of Alzheimer's disease. *Ann Clin Transl Neurol* 2016;3:12–20.
53. Llorens F, Thüne K, Tahir W, Kanata E, Diaz-Lucena D, Xanthopoulos K, et al. YKL-40 in the brain and cerebrospinal fluid of neurodegenerative dementias. *Mol Neurodegener* 2017;12:1–21.
54. Janelidze S, Mattsson N, Stomrud E, Lindberg O, Palmqvist S, Zetterberg H, et al. CSF biomarkers of neuroinflammation and cerebrovascular dysfunction in early Alzheimer disease. *Neurology* 2018;91:e867–7.
55. Terry RD, Masliah E, Salmon DP, Butters N, DeTeresa R, Hill R, et al. Physical basis of cognitive alterations in Alzheimer's disease: synapse loss is the major correlate of cognitive impairment. *Ann Neurol* 1991;30:572–80.
56. Kvartsberg H, Duits FH, Ingelsson M, Andreasen N, Öhrfelt A, Andersson K, et al. Cerebrospinal fluid levels of the synaptic protein neurogranin correlates with cognitive decline in prodromal Alzheimer's disease. *Alzheimer's Dementia* 2015;11:1180–90.
57. De Vos A, Jacobs D, Struyfs H, Fransen E, Andersson K, Portelius E, et al. C-terminal neurogranin is increased in cerebrospinal fluid but unchanged in plasma in Alzheimer's disease. *Alzheimer's Dementia* 2015;11:1461–9.
58. Mohaupt P, Pons M-L, Vialaret J, Delaby C, Hirtz C, Lehmann S.  $\beta$ -Synuclein as a candidate blood biomarker for synaptic degeneration in Alzheimer's disease. *Alzheimer's Res Ther* 2022;14:179.
59. Oeckl P, Anderl-Straub S, Danek A, Diehl-Schmid J, Fassbender K, Fließbach K, et al. Relationship of serum beta-synuclein with blood biomarkers and brain atrophy. *Alzheimer's Dementia* 2023;19:1358–71.
60. Oeckl P, Janelidze S, Halbgebauer S, Stomrud E, Palmqvist S, Otto M, et al. Higher plasma  $\beta$ -synuclein indicates early synaptic degeneration in Alzheimer's disease. *Alzheimer's Dementia* 2023;19:5095–102.
61. Nilsson J, Ashton NJ, Benedet AL, Montoliu-Gaya L, Gobom J, Pascoal TA, et al. Quantification of SNAP-25 with mass spectrometry and Simoa: a method comparison in Alzheimer's disease. *Alzheimer's Res Ther* 2022;14:1–10.
62. Sauer M, De Rocker C, Grötschel L, Goossens J, Benedet AL, Schöll M, et al. Blood-based SNAP-25 and VAMP-2 in Alzheimer's disease; relation to cognition, atrophy and synaptic density. *Alzheimer's Dementia* 2023;19:e083128.
63. Simrén J, Ashton NJ, Blennow K, Zetterberg H. Blood neurofilament light in remote settings: alternative protocols to support sample collection in challenging pre-analytical conditions. *Alzheimer's Dementia* 2021;13:e12145.